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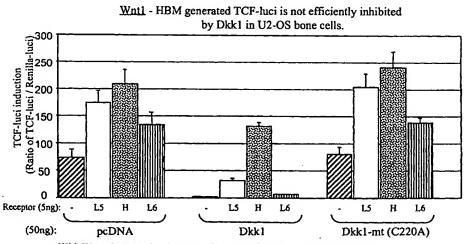
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(54) Title: REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS



- With Wnt1 the TCF-signal generated by LRP5 is greater than that of LRP6.
- LRP5/6 -Wnt1 induced TCF- is efficiently blocked byDkk1

(57) Abstract: The present invention provides reagents, compounds, compositions, and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk, including Dkk-1. The various nucleic acids, polypeptides, antibodies, assay methods, diagnostic methods, and methods of treatment of the present invention are related to and impact on Dkk, LRP5, LRP6, HBM, and Wnt signaling. Dkk, LRP5, LRP6, HBM, and Wnt are implicated in bone and lipid cellular signaling. Thus, the present invention provides reagents and methods for modulating lipid levels and/or bone mass and is useful in the treatment and diagnosis of abnormal lipid levels and bone mass disorders, such as osteoporosis.



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REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS

FIELD OF THE INVENTION

The present invention relates to signal transduction, bone development, bone loss disorders, modulation of lipid-related conditions, research reagents, methods of screening drug leads, drug development, treatments for bone and/or lipid disorders, screening and development of therapies, molecular, cellular, and animal models of bone and/or lipid development and maintenance, which are mediated by Dkk, including Dkk-1, and/or LRP5, LRP6, HBM or other members of the Wnt pathway.

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BACKGROUND OF THE INVENTION

Two of the most common types of osteoporosis are postmenopausal and senile osteoporosis. Osteoporosis affects both men and women, and, taken with other abnormalities of bone, presents an ever-increasing health risk for an aging population. The most common type of osteoporosis is that associated with menopause. Most women lose between 20-60% of the bone mass in the trabecular compartment of the bone within 3-6 years after the cessation of menses. This rapid bone loss is generally associated with an increase of bone resorption and formation. However, the resorptive cycle is more dominant and the result is a net loss of bone mass. Osteoporosis is a common and serious disease among postmenopausal women. There are an estimated 25 million women in the United States alone who are afflicted with this disease. The results of osteoporosis are personally harmful. and also account for a large economic loss due to its chronicity and the need for extensive and long-term support (e.g., hospitalization and nursing home care) from disease sequelae. This is especially true in elderly patients. Additionally, while osteoporosis is generally not thought of as a life-threatening condition, a 20-30% mortality rate is related to hip fractures in elderly women. A large percentage of this mortality rate can be directly associated with postmenopausal osteoporosis.

The most vulnerable tissue in the bone to the effects of postmenopausal osteoporosis is the trabecular bone. This tissue is often referred to as spongy bone and is particularly concentrated near the ends of the bone, near the joints, and in the vertebrae of the spine. The trabecular tissue is characterized by small structures which inter-connect with each other as well as the more solid and dense cortical tissue which makes up the outer surface and central shaft of the bone. This criscross network of trabeculae gives lateral support to the outer cortical structure and is critical to the biomechanical strength of the overall structure. In postmenopausal osteoporosis, it is primarily the net resorption and loss of the trabeculae which lead to the failure and fracture of the bone. In light of the loss of the trabeculae in postmenopausal women, it is not surprising that the most common fractures are those associated with bones which are highly dependent on trabecular support, e.g., the vertebrae, the neck of the femur, and the forearm. Indeed, hip fracture, Colle's fractures, and vertebral crush fractures are indicative of postmenopausal osteoporosis. Osteoporosis affects cortical as well as trabecular bone. Alterations in endosteal bone resorption and Haversian remodeling with age affect cortical thickness and structural integrity contributing the increased risk for fracture.

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One of the earliest generally accepted methods for treatment of postmenopausal osteoporosis was estrogen replacement therapy. Although this therapy frequently is successful, patient compliance is low, primarily due to the undesirable side-effects of chronic estrogen treatment. Frequently cited side-effects of estrogen replacement therapy include reinitiation of menses, bloating, depression, and, potentially, increased risk of breast or uterine cancer. In order to limit the known threat of uterine cancer in women who have not had a hysterectomy, a protocol of estrogen and progestin cyclic therapy is often employed. This protocol is similar to that used in birth control regimens, and often is not tolerated by women because of the side-effects characteristic of progestin. More recently, certain antiestrogens, originally developed for the treatment of breast cancer, have been shown in experimental models of postmenopausal osteoporosis to be efficacious. Among these agents is raloxifene (See, U.S. Patent No. 5,393,763; Black et al., J.

Clin. Invest., 93:63-69 (1994); and Ettinger et al., JAMA 282:637-45 (1999)). In addition, tamoxifen, a widely used clinical agent for treating breast cancer, has been shown to increase bone mineral density in post menopausal women suffering from breast cancer (Love et al., N. Engl. J. Med., 326:852-856 (1992)).

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Another therapy for the treatment of postmenopausal osteoporosis is the use of calcitonin. Calcitonin is a naturally occurring peptide which inhibits bone resorption and has been approved for this use in many countries (Overgaard *et al.*, *Br. Med. J.*, 305:556-561 (1992)). The use of calcitonin has been somewhat limited, however. Its effects are very modest in increasing bone mineral density, and the treatment is very expensive. Another therapy for the treatment of postmenopausal osteoporosis is the use of bisphosphonates. These compounds were originally developed for treating Paget's disease and malignant hypercalcemia. They have been shown to inhibit bone resorption. Alendronate, a bisphosphonate, has been approved for the treatment of postmenopausal osteoporosis. These agents may be helpful in the treatment of osteoporosis, but these agents also have potential liabilities which include osteomalacia, extremely long half-life in bone (greater than 2 years), and possible "frozen bone syndrome," *e.g.*, the cessation of normal bone remodeling.

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Senile osteoporosis is similar to postmenopausal osteoporosis in that it is marked by the loss of bone mineral density and resulting increase in fracture rate, morbidity, and associated mortality. Generally, it occurs in later life, *i.e.*, after 70 years of age. Historically, senile osteoporosis has been more common in females, but with the advent of a more elderly male population, this disease is becoming a major factor in the health of both sexes. It is not clear what, if any, role hormones such as testosterone or estrogen have in this disease, and its etiology remains obscure. Treatment of this disease has not been very satisfactory. Hormone therapy, estrogen in women and testosterone in men, has shown equivocal results; calcitonin and bisphosphonates may be of some utility.

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The peak mass of the skeleton at maturity is largely under genetic control.

Twin studies have shown that the variance in bone mass between adult monozygotic

twins is smaller than between dizygotic twins (Slemenda *et al.*, *J. Bone Miner. Res.*, 6: 561-567 (1991); Young *et al.*, *J. Bone Miner. Res.*, 6:561-567 (1995); Pocock *et al.*, *J. Clin. Invest.*, 80:706-710 (1987); Kelly *et al.*, *J. Bone Miner. Res.*, 8:11-17 (1993)). It has been estimated that up to 60% or more of the variance in skeletal mass is inherited (Krall *et al.*, *J. Bone Miner. Res.*, 10:S367 (1993)). Peak skeletal mass is the most powerful determinant of bone mass in elderly years (Hui *et al.*, *Ann. Int. Med.*, 111:355-361 (1989)), even though the rate of age-related bone loss in adult and later life is also a strong determinant (Hui *et al.*, *Osteoporosis Int.*, 1:30-34 (1995)). Since bone mass is the principal measurable determinant of fracture risk, the inherited peak skeletal mass achieved at maturity is an important determinant of an individual's risk of fracture later in life. Thus, study of the genetic basis of bone mass is of considerable interest in the etiology of fractures due to osteoporosis.

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Recently, a strong interest in the genetic control of peak bone mass has developed in the field of osteoporosis. The interest has focused mainly on candidate genes with suitable polymorphisms to test for association with variation in bone mass within the normal range, or has focused on examination of genes and gene loci associated with low bone mass in the range found in patients with osteoporosis. The vitamin D receptor locus (VDR) (Morrison et al., Nature, 367:284-287 (1994)), PTH gene (Howard et al., J. Clin. Endocrinol. Metab., 80:2800-2805 (1995); Johnson et al., J. Bone Miner. Res., 8:11-17 (1995); Gong et al., J. Bone Miner. Res., 10:S462 (1995)) and the estrogen receptor gene (Hosoi et al., J. Bone Miner. Res., 10:S170 (1995); Morrison et al., Nature, 367:284-287 (1994)) have figured most prominently in this work. These studies are difficult because bone mass (i.e., the phenotype) is a continuous, quantitative, polygenic trait, and is confounded by environmental factors such as nutrition, co-morbid disease, age, physical activity, and other factors. Also, this type of study design requires large numbers of subjects. In particular, the results of VDR studies to date have been confusing and contradictory (Garnero et al., J. Bone Miner. Res., 10:1283-1288 (1995); Eisman et al., J. Bone. Miner. Res., 10:1289-1293 (1995); Peacock, J. Bone Miner. Res., 10:1294-1297 (1995)).

Furthermore, thus far, the art has not determined the mechanism(s) whereby the genetic influences exert their effect on bone mass.

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While it is well known that peak bone mass is largely determined by genetic rather than environmental factors, studies to determine the gene loci (and ultimately the genes) linked to variation in bone mass are difficult and expensive. Study designs which utilize the power of linkage analysis, *e.g.*, sib-pair or extended family, are generally more informative than simple association studies, although the latter do have value. However, genetic linkage studies involving bone mass are hampered by two major problems. The first problem is the phenotype, as discussed briefly above. Bone mass is a continuous, quantitative trait, and establishing a discrete phenotype is difficult. Each anatomical site for measurement may be influenced by several genes, many of which may be different from site to site. The second problem is the age component of the phenotype. By the time an individual can be identified as having low bone mass, there is a high probability that their parents or other members of prior generations will be deceased and therefore unavailable for study, and younger generations may not have even reached peak bone mass, making their phenotyping uncertain for genetic analysis.

Thus, there is a need in the art for additional research tools for the elucidation of the molecular mechanism of bone modulation, for the screening and development of candidate drugs, and for treatments of bone development and bone loss disorders. The present invention is directed to these, as well as other, important ends.

In addition to bone modulation, the present invention relates to modulation of lipid levels. Cardiovascular disease is the most common cause of mortality in the United States, and atherosclerosis is the major cause of heart disease and stroke. It is widely appreciated that cholesterol plays an important role in atherogenesis. Normally, most cholesterol serves as a structural element in the walls of cells, whereas much of the rest is in transit through the blood or functions as the starting material for the synthesis of bile acids in the liver, steroid hormones in endocrine cells and vitamin D in skin. The transport of cholesterol and other lipids through the

circulatory system is facilitated by their packaging into lipoprotein carriers. These spherical particles comprise protein and phospholipid shells surrounding a core of neutral lipid, including unesterified ("free") or esterified cholesterol and triglycerides. Risk for atherosclerosis increases with increasing concentrations of low density lipoprotein (LDL) cholesterol, whereas risk is inversely proportional to levels of high-density lipoprotein (HDL) cholesterol. The receptor-mediated control of plasma LDL levels has been well-defined, and recent studies have now provided new insights into HDL metabolism.

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The elucidation of LDL metabolism began in 1974 by Michael Brown and Joseph Goldstein. In brief, the liver synthesizes a precursor lipoprotein (very low density lipoprotein, VLDL) that is converted during circulation to intermediate density lipoprotein (IDL) and then to LDL. The majority of the LDL receptors expressed in the body are on the surfaces of liver cells, although virtually all other tissues ("peripheral tissues") express some LDL receptors. After binding, the receptorlipoprotein complex is internalized by the cells via coated pits and vesicles, and the entire LDL particle is delivered to lysosomes, wherein it is dissembled by enzymatic hydrolysis, releasing cholesterol for subsequent cellular metabolism. This wholeparticle uptake pathway is called "receptor-mediated endocytosis." Cholesterolmediated feedback regulation of both the levels of LDL receptors and cellular cholesterol biosynthesis help ensure cellular cholesterol homeostasis. Genetic defects in the LDL receptor in humans results in familial hypercholesterolemia, a disease characterized by elevated plasma LDL cholesterol and premature atherosclerosis and heart attacks. One hypothesis for the deleterious effects of excess plasma LDL cholesterol is that LDL enters the artery wall, is chemically modified, and then is recognized by a special class of receptors called macrophage scavenger receptors, that mediate the cellular accumulation of the LDL cholesterol in the artery, eventually leading to the formation of an atherosclerotic lesion.

The major lipoprotein classes include intestinally derived chylomicrons that transport dietary fats and cholesterol, hepatic-derived VLDL, IDL, and LDL that can be atherogenic, and hepatic- and intestinally-derived HDL that are antiatherogenic.

Apoprotein B (ApoB) is necessary for the secretion of chylomicrons (ApoB48) and VLDL, IDL, and LDL (ApoB100). Plasma levels of VLDL triglycerides are determined mainly by the rates of secretion in LDL lipolytic activity. Plasma levels of LDL cholesterol are determined mainly by the secretion of ApoB100 into plasma, the efficacy with which VLDL are converted to LDL and by LDL receptor-mediated clearance. Regulation of HDL cholesterol levels is complex and is affected by rates of synthesis of its Apo proteins, rates of esterification of free cholesterol to cholesterol ester by LCAT, levels of triglyceride-rich lipoproteins and CETP-mediated transfer of cholesterol esters from HDL, and clearance from plasma of HDL lipids and Apo proteins.

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Normal lipoprotein transport is associated with low levels of triglycerides and LDL cholesterol and high levels of HDL cholesterol. When lipoprotein transport is abnormal, lipoprotein levels can change in ways that predispose individuals to atherosclerosis and arteriosclerosis (see Ginsburg, *Endocrinol. Metab. Clin. North Am.*, 27:503-19 (1998)).

Several lipoprotein receptors may be involved in cellular lipid uptake. These receptors include: scavenger receptors; LDL receptor-related protein/α2-macroglobulin receptor (LRP); LDL receptor; and VLDL receptor. With the exception of the LDL receptor, all of these receptors are expressed in atherosclerotic lesions while scavenger receptors are mostly expressed in macrophages, the LRP and VLDL receptors may play an important role in mediating lipid uptake in smooth muscle cells (Hiltunen *et al.*, *Atherosclerosis*, 137 suppl.:S81-8 (1998)).

A major breakthrough in the pharmacologic treatment of hypercholesterolemia has been the development of the "statin" class of 3-hydroxy-3-methylglutaryl-CoA reductase (HMG CoA reductase) inhibitory drugs. 3-hydroxy-3-methylglutaryl-CoA reductase is the rate controlling enzyme in cholesterol biosynthesis, and its inhibition in the liver stimulates LDL receptor expression. As a consequence, both plasma LDL cholesterol levels and the risk for atherosclerosis decrease. The discovery and analysis of the LDL receptor system has had a profound impact on cell biology, physiology, and medicine.

HDL is thought to remove unesterified, or "free" cholesterol (FC) from peripheral tissues, after which most of the cholesterol is converted to cholesterol ester (CE) by enzymes in the plasma. Subsequently, HDL cholesterol is efficiently delivered directly to the liver and steroidogenic tissues via a selective uptake pathway and the HDL receptor, SR-BI (class B type I scavenger receptor) or, in some species, transferred to other lipoproteins for additional transport in metabolism (see Krieger, *Proc. Natl. Acad. Sci. USA*, 95:4077-4080 (1998)).

These issues illustrate a need in the art for additional research tools for the elucidation of the molecular mechanism of lipid modulation, for the screening and development of candidate drugs, and for treatments of lipid levels and lipid level modulation disorders. The present invention is directed to these, as well as other, important ends.

SUMMARY OF THE INVENTION

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The present invention provides reagents, compounds, compositions and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk proteins. LRP5 is also referred to as Zmax1 or Zmax. Thus, when discussing methods, reagents, compounds, and compositions of the invention which relate to the interaction between Dkk and LRP5 (or Zmax1). the invention is also to be understood to encompass embodiments relating to interactions between Dkk and LRP6 and Dkk and HBM. Moreover, where Dkk is discussed herein, it is to be understood that the methods, reagents, compounds, and compositions of the present invention include the Dkk family members, including but not limited to Dkk-1, Dkk-2, Dkk-3, Dkk-4 and Soggy. Furthermore, the invention encompasses novel fragments of Dkk-1 which demonstrate a binding interaction between the ligand binding domain (LBD) of LRP5 and additional proteins and/or which can modulate an interaction between LRP5, or a variant or fragment thereof, and a Dkk protein. The invention provides assays, methods, compositions, and compounds relating to Dkk-Wnt signaling. Numerous Wnt proteins are compatible with the present invention, including Wnt1-Wnt19, and particularly, Wnt1, Wnt3,

Wnt3a, and Wnt10b. The present invention further provides reagents, compounds, compositions and methods modulating interactions between one or more other proteins and Dkk-1. The present invention also provides a series of peptide aptamers which bind to Dkk-1 or to LRP5 (or HBM and/or LRP6).

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The polypeptides of the invention, for example in the form of peptide oligomers, aptamers, proteins, and protein fragments as well as the nucleic acids of the invention, for example in the form of nucleic acids which encode the polypeptides of the invention as well as antisense, or complimentary nucleic acids, are useful as reagents for the study of bone mass and lipid level modulation. The polypeptides and nucleic acids of the invention are also useful as therapeutic and diagnostic agents.

The present invention provides useful reagents for the modulation of Dkk

present invention provides a series of peptide aptamers which bind Dkk-1 or LRP5,

proteins with LRP5, LRP6, and/or HBM, the modulation Dkk-1 and/or Dkk-1

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interacting protein activity, and modulation of LRP5/Dkk-1, LRP6/Dkk1 and HBM/Dkk-1 interactions and Dkk-1/Dkk-1 interacting protein interactions. The

LRP6, and/or HBM.

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An object of the invention is to provide for a method of regulating LRP5/LRP6/HBM/HBM-like activity in a subject comprising administering a therapeutically effective amount of a composition which modulates Dkk activity. The subject can be a vertebrate or an invertebrate organism, but more preferably the organism is a canine, a feline, an ovine, a primate, an equine, a porcine, a caprine, a camelid, an avian, a bovine, or a rodent organism. A more preferred organism is a human. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in trabecular

connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. The invention further provides such a method wherein the composition comprises a Dkk, Dkk-1 or a LRP5/LRP6/HBM binding fragment thereof, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The invention further provides such a method wherein the composition comprises one or more of the proteins which interact with Dkk, including Dkk-1, such as those depicted in Figure 5, or a Dkk-binding fragment thereof, or an antisense, siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding one or more Dkk interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises an LRP5/LRP6/Zmax1 antibody. Dkk antibody, a Dkk-1 antibody or an antibody to a Dkk-1 interacting protein. The invention further provides such a method wherein the compositions comprise an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188). or a mimetic of such an aptamer. The method further provides that invention further provides such a method wherein the compositions comprise an aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer.

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A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk to LRP5/LRP6/Zmax1, particularly Dkk-1, or the binding of Dkk-1 to a Dkk-1 interacting protein, such as those shown in Figure 5. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NOs:189-192) (particularly, peptide (SEQ ID NO:191) and 13 (including SEQ IDNOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise LRP5 antibodies.

Another aspect of the invention is to provide for a method of regulating Dkk-Wnt pathway activity in a subject comprising administering a therapeutically effective amount of a composition which modulates Dkk-Wnt pathway activity. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass

and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter. and an increase in inorganic bone content. In another preferred embodiment, the Wnt is Wnt1-Wnt19. In a particularly preferred embodiment, the Wnt is Wnt1, Wn3, Wnt3a, or Wnt10b. Preferred compositions comprise Dkk-modulating or Dkk-1modulating compounds or one or more Dkk interacting or Dkk-1 interacting proteins. or a Dkk-binding fragment thereof. Other preferred Dkk modulating compositions comprise a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. Also contemplated are antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk, including Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk, including Dkk-1, to LRP5, LRP6, or HBM or the binding of Dkk, including Dkk-1, to a Dkk interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein the composition comprises an aptamer of Dkk or Dkk-1, such as those depicted. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208). Preferred compositions of the present invention also comprise LRP5 antibodies.

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A further aspect of the invention is to provide for a method of modulating Wnt signaling in a subject comprising administering a therapeutically effective amount of a composition which modulates Dkk activity or modulates Dkk interaction with LRP5

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(or LRP6 or HBM). In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. In another preferred embodiment, the Wnt is Wnt1-Wnt19. In a particularly preferred embodiment, the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b. Preferred Wnt modulating compositions comprise one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active or LRP5/LRP6/HBM binding fragment thereof. Also contemplated are antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk or Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or blocking the binding of Dkk, including Dkk-1, to LRP5, LRP6, or HBM or the binding of Dkk or Dkk-1 to a Dkk interacting or Dkk-1 interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein compositions comprising an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such an aptamer. The invention further provides such a method wherein the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. The invention further provides such a method wherein compositions of an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NO:189-192

(particularly peptide (SEQ ID NO:191) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Additional preferred compositions of the present invention also comprise LRP5 antibodies.

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Additionally, the invention provides for a method of modulating bone mass and/or lipid levels in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5 in an amount effective to modulate bone mass and/or lipid levels, wherein bone mass and/or lipid levels are in need of modulation. In a preferred embodiment, the Dkk protein is Dkk-1. In a particularly preferred embodiment, Dkk-1 activity is decreased. In another embodiment, Dkk activity modulates bone mass and/or lipid levels. In a preferred embodiment, bone mass is increased and/or lipid levels are decreased. In another preferred embodiment, the modulation in bone mass is an increase in bone strength determined via one or more of a decrease in fracture rate, an increase in areal bone density, an increase in volumetric mineral bone density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density or thickness, an increase in bone diameter, and an increase in inorganic bone content. Preferred bone mass and/or lipid modulating compositions comprise one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active or LRP5/LRP6/HBM binding fragment thereof. Also contemplated are antisense. siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding one or more Dkk interacting or Dkk-1 interacting proteins. The invention further provides such a method wherein the composition comprises a biologically active or LRP5/LRP6/HBM binding fragment of Dkk, including Dkk-1, such as those depicted in Figure 6 or a mimetic of those fragments depicted in Figure 6. The Dkk modulating composition may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The invention further provides such a method wherein the composition comprises an aptamer of Dkk or Dkk-1, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such an aptamer. A composition of the present invention may modulate activity either by enhancing or inhibiting the binding of Dkk, including Dkk-1, to LRP5, LRP6,

or HBM or the binding of Dkk, including Dkk-1, to a Dkk interacting protein, such as those shown in Figure 5. The invention further provides such a method wherein the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figures 4 (SEQ ID NOs:189-192 (particularly peptide 13 (SEQ ID NO:191)) and 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise LRP5 antibodies. It is a further aspect of the invention that such lipid-modulated diseases include a cardiac condition, atherosclerosis, familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid levels due to dialysis and/or diabetes, and an elevated lipid level of unknown etiology.

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Bone disorders contemplated for treatment and/or diagnosis by the methods and compositions disclosed herein include a bone development disorder, a bone fracture, age related loss of bone, a chondrodystrophy, a drug-induced bone disorder, high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and rickets.

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It is a further object of the invention to provide a method of screening for compounds or compositions which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

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- (a) exposing Dkk or a LRP5/LRP6/HBM binding fragment thereof to a compound; and
- (b) determining whether said compound binds to Dkk or the LRP5/LRP6/HBM binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1. In a particularly preferred embodiment, the binding of Dkk-1 to LRP5/LRP6/HBM is decreased.

It is a further object of the invention to provide a method of screening compounds or compositions which modulate the interaction of DKK with LRP5, LRP6, HBM, or a DKK-finding fragment thereof comprising:

- (a) exposing DKK or a LRP5/LRP6/HBM binding fragment thereof to a compound; and,
- (b) determining whether said compound modulates the interaction of Dkk with LRP5, LRP6, or HBM, or the Dkk-binding fragment of LRP5/LRP6/HBM.

In a preferred embodiment, the Dkk is Dkk-1. In a particularly preferred embodiment, the interaction of Dkk-1 with LRP5/LRP6/HBM is decreased.

It is a further object of the invention to provide a method of screening for compounds or compositions which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

- (a) exposing Dkk or a LRP5/LRP6/HBM binding fragment thereof to a compound;
- (b) determining whether said compound binds to Dkk or the LRP5/LRP6/HBM binding fragment thereof; and,
- (c) further determining whether said compound modulates the interaction of Dkk with LRP5, LRP6, or HBM, or the Dkk-binding fragment of LRP5/LRP6/HBM.

In preferred embodiments of such methods, Dkk or a biologically active fragment thereof is attached to a solid substrate. In an alternative embodiment of the invention, LRP5/LRP6/HBM, or a biologically active fragment thereof (such as the ligand binding domain), is exposed to the compound. Another aspect of the invention provides for compounds and compositions identified by the disclosed methods. A preferred embodiment of the invention provides that the compound screened in an afore-mentioned method is one or more proteins which interact with Dkk, particularly Dkk-1, as depicted in Figure 5, or a LRP5/LRP6/HBM-binding fragment thereof. Another preferred embodiment provides that the compound comprises a Dkk or Dkk-1 peptide aptamer, such as those depicted in Figure 3 (SEQ

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ID NOs:171-188), or a mimetic of such aptamers. The compound may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The method further provides that the compound comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk-1 interacting protein. The invention further provides that the compound may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compounds of the present invention also comprise LRP5 antibodies.

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It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

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(a) exposing a Dkk interacting proteins or a Dkkbinding fragment thereof to a compound; and,

(b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

- (a) exposing Dkk interacting protein(s) or a Dkkbinding fragment thereof to a compounds; and,
- (b) determining whether said compound modulatesthe interaction of Dkk and Dkk interacting proteins.

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It is a further object of the invention to provide a method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:

> (a) exposing a Dkk interacting proteins or a Dkkbinding fragment thereof to a compound;

(b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof; and,

(c) further determining whether said compound modulates the interaction of Dkk and Dkk interacting proteins.

In a preferred embodiment, Dkk is Dkk-1.

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In preferred embodiments of such methods, the Dkk interacting proteins, particularly Dkk-1 interacting proteins, or a Dkk-binding fragment thereof are attached to a solid substrate. Another aspect of the invention provides for compounds and compositions identified by the disclosed methods. A preferred embodiment provides that the compound comprises a Dkk or Dkk-1 peptide aptamer, such as those depicted in Figure 3 (SEQ ID NOs:171-188), or a mimetic of such aptamers. The compound may also comprise a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The compound may also comprise an antibody to a Dkk interacting or Dkk-1 interacting protein.

It is another object of the invention to provide for a composition for treating bone mass disorders comprising a LRP5/LRP6/HBM modulating compound and a pharmaceutically acceptable excipient and/or carrier therefor. Preferred LRP5 (or LRP6 or HBM) modulating compounds include Dkk or Dkk-1 or a LRP5/LRP6/HBM binding fragment thereof. Also contemplated are compounds which comprise monoclonal or polyclonal antibodies or immunologically active fragments thereof which bind to Dkk, including Dkk-1, and a pharmaceutically acceptable excipient and/or carrier. Another preferred embodiment provides that the modulating compound comprises one or more Dkk interacting or Dkk-1 interacting proteins, or a biologically active fragment thereof. Also contemplated are compounds which comprise monoclonal or polyclonal antibodies or immunologically active fragments thereof which bind to Dkk interacting or Dkk-1 interacting proteins, or a biologically active fragment thereof, and a pharmaceutically acceptable excipient and/or carrier.

Another preferred embodiment provides that the modulating compound comprises an antisense, siRNA, and shRNA molecule which recognizes and binds to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. Another preferred embodiment provides that the modulating compound comprises a Dkk or Dkk-1 peptide aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. Another embodiment provides that the compound comprises an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compounds of the present invention also comprise LRP5 antibodies.

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It is a further object of the invention to provide a pharmaceutical composition for treating a Dkk-mediated disease or condition comprising a compound which modulates Dkk activity and a carrier therefor, including pharmaceutically acceptable excipients. Such compositions include those wherein the compound comprises an antisense, siRNA, and shRNA molecule or an antibody which binds to Dkk, including Dkk-1, and thereby prevents it from interacting with LRP5, LRP6, or HBM. Other such compositions include one or more of Dkk interacting or Dkk-1 interacting proteins, such as those depicted in Figure 5, or a Dkk-binding fragment thereof, or a monoclonal or polyclonal antibody, or immunologically active fragment thereof, which binds to a Dkk interacting or Dkk-1 interacting protein or Dkk-binding fragment thereof. Other contemplated compositions include antisense, siRNA, and shRNA molecules which recognize and bind to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. Further contemplated compositions include Dkk and Dkk-1 peptide aptamers, such as those depicted in Figure 3 (SEQ ID NOs;171-188). mimetics of such aptamers, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. Other contemplated compositions comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191))

and Figure 13 (including SEQ ID NO:204-214), or a mimetic of such an aptamer. Other preferred compositions of the present invention comprise LRP5 antibodies.

A further object of the invention to provide for a method of modulating the expression of a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein in an organism, such as those shown in Figure 5, comprising the step of administering to the organism an effective amount of composition which modulates the expression of a nucleic acid encoding a Dkk-1 interacting protein. In a preferred embodiment, said composition comprises an antisense, siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein.

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One aspect of the invention provides for a method of modulating at least one activity of Dkk or a Dkk-1 interacting protein comprising administering an effective amount of a composition which modulates at least one activity of Dkk or a Dkk-1 interacting protein. The invention provides for a composition comprising a Dkk interacting or Dkk-1 interacting protein, such as those shown in Figure 5, or a biologically active fragment thereof. Other agents contemplated for this method are antisense, siRNA, or shRNA molecules which recognize and bind to a nucleic acid encoding a Dkk interacting or Dkk-1 interacting protein. The method further provides that the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacing or Dkk-1 interacting protein. In another preferred embodiment, the composition comprises a Dkk or Dkk-1 peptide aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a peptide aptamer of a Dkk interacting or Dkk-1 interacting protein, or a mimetic of such an aptamer. The method provides that a composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NO:189-192) (particularly peptide including (SEQ ID NO:191)) and Figure including (SEQ ID NOs:204-214), or a mimetic of such an aptamer. Preferred compositions of the present invention also comprise LRP5 antibodies. In a further preferred embodiment, the modulated Dkk activity is lipid modulation or bone mass modulation.

In all of the testing/screening embodiments of the present invention discussed below to obtain compounds or compositions which ultimately impact LRP5/LRP6/HBM signaling, one skilled in the art will recognize that HBM can be used as a control in the absence of a test sample or compound. Further, the effect of a test sample of compound on Wnt signaling through the interaction of Dkk with LRP5/LRP6/HBM does not necessarily require a direct measurement of an association or interaction of Dkk and LRP5/LRP6/HBM. Other positive phenotypes/activities established by the High Bone Mass phenotype or by using HBM as a control.

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One aspect of the invention provides for a method of identifying binding partners for a Dkk protein comprising the steps of:

- (a) exposing the Dkk protein(s) or a LRP5/LRP6 binding fragment thereof to a potential binding partner; and
- (b) determining if the potential binding partner binds to a Dkk protein or the LRP5/LRP6 binding fragment thereof.

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In a preferred embodiment, the Dkk is Dkk-1.

Another aspect of the invention is to provide for a method of identifying a compound that effects Dkk-mediated activity comprising

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(a) providing a group of transgenic animals having (1) a regulatable one or more Dkk interacting protein genes, (2) a knock-out of one or more Dkk interacting protein genes, or (3) a knock-in of one or more Dkk interacting protein genes;

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(b) providing a second group of control animalsrespectively for the group of transgenic animals in step (a); and

(c) exposing the transgenic animal group and the control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and

(d) comparing the transgenic animal group and the control animal group and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.

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In a preferred embodiment, the Dkk is Dkk-1.

It is another aspect of the invention to provide for a method for determining whether a compound modulates a Dkk interacting protein, said method comprising the steps of:

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 (a) mixing the Dkk interacting protein or a Dkk-binding fragment thereof with the ligand binding domain of Dkk in the presence of said at least one compound;

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(b) measuring the amount of said binding domain of Dkk bound to said Dkk interacting protein or the Dkk-binding fragment thereof as compared to a control without said at least one compound; and

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(c) determining whether the compound reduces the amount of said binding domain of Dkk binding to said Dkk interacting protein or Dkk-binding fragment thereof.

In a preferred embodiment, the Dkk is Dkk-1.

In a preferred embodiment, the binding domain is attached to a solid substrate. The invention further provides for compounds identified by this method. In a preferred embodiment, the invention provides that the Dkk interacting or Dkk-1 interacting protein is detected by antibodies. In another preferred embodiment, the solid substrate is a microarray. Another preferred embodiment provides that the ligand binding domain of Dkk and/or Dkk interacting protein is fused or conjugated to a peptide or protein. The invention also provides that the compounds include Dkk

and Dkk-1 peptide aptamers, mimetics of Dkk and Dkk-1 peptide aptamers, Dkk and Dkk-1 interacting proteins peptide aptamers, or mimetics of such aptamers.

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An aspect of the invention provides a composition comprising one or more polypeptide sequences of one or more Dkk-1 interacting proteins, or a biologically active fragment thereof, one or more Dkk proteins, or a biologically active fragment thereof, or LRP5/LRP6/HBM polypeptide sequences or a biologically active fragment thereof (for example, the ligand binding domain) and a pharmaceutically acceptable excipient and/or carrier. Another aspect of the invention provides that the composition comprises a Dkk or Dkk-1 antibody or an antibody to a Dkk interacting or Dkk-1 interacting protein and a pharmaceutically acceptable excipient. A composition of the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. A composition of the present invention may comprise a Dkk peptide aptamer, for example as shown in Figure 3 (SEQ ID NOs:171-188). Preferred compositions of the present invention also comprise LRP5 antibodies.

Another aspect of the invention is to provide an antibody or immunologically active antibody fragment which recognizes and binds to a Dkk-1 amino acid sequence selected from the group consisting of: Asn34-His266 (SEQ ID NO:110), Asn34-Cys245 (SEQ ID NO:111), Asn34-Lys182 (SEQ ID NO:112), Cys97-His266 (SEQ ID NO:113), Val139-His266 (SEQ ID NO:114), Gly183-His266 (SEQ ID NO:115), Cys97-Cys245 (SEQ ID NO:116), or Val139-Cys245 (SEQ ID NO:117) of human Dkk-1. Additional antibodies may bind to any of the sequences depicted in Figures 3 (SEQ ID NOs:171-188) and Figure 4 (SEQ ID NOs:189-192). Another aspect of the invention is to provide for polyclonal antibodies to one or more amino acid sequences: Peptide 1 -GNKYQTIDNYQPYPC (SEQ ID NO:118), Peptide 2 - LDGYSRRTTLSSKMYHTKGQEG (SEQ ID NO:119), Peptide 3 - RIQKDHHQASNSSRLHTCQRH (SEQ ID NO:120), Peptide 4 - RGEIEETITESFGND (SEQ ID NO:121), and Peptide 5 - EIFQRCYCGEGLSCRIQKD (SEQ ID NO: 122).

It is a further object of the invention to provide a nucleic acid encoding a Dkk protein, e.g. Dkk-1, a Dkk interacting or Dkk-1 interacting protein aptamer, or an LRP5 aptamer comprising a nucleic acid encoding a scaffold protein in-frame with the activation domain of Gal4 or LexA that is in-frame with a nucleic acid which encodes for a Dkk or Dkk-1 or Dkk interacting or Dkk-1 interacting protein amino acid sequence. Preferably the scaffold protein is thioredoxin (trxA), S1 nuclease from Staphylococcus or M13. Other preferable embodiments include Dkk-1 amino acid sequences selected from Figure 6.

It is yet a further object of the invention to provide a composition comprising a polypeptide sequence of Figure 3 (SEQ ID NOs:171-188), Figure 4 (SEQ ID NO:189-192), or of Dkk-1 interacting proteins identified in Figure 5 and a pharmaceutically acceptable excipient and/or carrier.

Another aspect of the invention includes a method of detecting the modulatory activity of a compound on the binding interaction of a first peptide and a second peptide of a peptide binding pair that bind through extracellular interaction in their natural environment, comprising:

- (i) culturing at least one eukaryotic cell, wherein the eukaryotic cell comprises;
 - a) a nucleotide sequence encoding a first heterologous fusion protein comprising the first peptide or a segment thereof joined to a DNA binding domain of a transcriptional activation protein;
 - a nucleotide sequence encoding a second heterologous fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

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c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element produces a selected phenotype;

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- (ii) incubating a compound with the eukaryotic cell under conditions suitable to detect the selected phenotype; and
- (iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which produces the selected phenotype;

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wherein (1) said first peptide is a Dkk peptide and said second peptide is a peptide selected from LRP5, HBM, LRP6, and the Dkk-binding portion of LRP5/LRP6/HBM or (2) said first peptide is a Dkk-interacting protein or the Dkk-binding fragment thereof, and said second peptide is a Dkk peptide.

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In one embodiment, the eukaryotic cell is a yeast cell. In a preferred embodiment, the yeast cell is Saccharomyces. In a particularly preferred embodiment, the Saccharomyces cell is Saccharomyces cerevisiae. The invention further provides that the compound may comprise a Dkk interacting or Dkk-1 interacting protein, or a biologically active fragment thereof. In one embodiment, the Dkk interacting or Dkk-1 interacting protein, or a Dkk-binding fragment thereof, is added directly to the assay. In another embodiment, the Dkk interacting or Dkk-1 interacting protein, or a Dkk-binding fragment thereof, is recombinantly expressed by the eukaryotic cell in addition to the first and second peptides. In a preferred embodiment the compound comprises a Dkk or Dkk-1 aptamer, a mimetic of a Dkk or Dkk-1 peptide aptamer, a Dkk interacting or Dkk-1 interacting protein aptamer, or a mimetic of a Dkk-1 interacting protein aptamer. Other preferred embodiments provide that the compound comprises an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Alternatively, the present invention also provides that the compound may

comprise LRP5 antibodies or Dkk antibodies. In another embodiment, the yeast cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter element, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion. In another embodiment, the peptide binding pair comprises a ligand and a receptor to which the ligand binds. In one embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In another embodiment, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In one embodiment, the DNA binding domain comprises a heterologous DNA-binding domain of a transcriptional activation protein. In a preferred embodiment, the DNA binding protein is selected from the group consisting of a mammalian steroid receptor and bacterial LexA protein. In another embodiment, the reporter element is selected from the group consisting of lacZ, a polynucleotide encoding luciferase, a polynucleotide encoding green fluorescent protein (GFP), and a polynucleotide encoding chloramphenicol acetyltransferase. In a particularly preferred embodiment, the reporter element is lacZ

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The invention further provides for a rescue screen for detecting the activity of a compound for modulating the binding interaction of a first peptide and a second peptide of a peptide binding pair, comprising:

- (i) culturing at least one yeast cell, wherein the yeast cell comprises;
 - a) a nucleotide sequence encoding a first heterologous fusion
 protein comprising the first peptide or a segment thereof joined
 to a DNA binding domain of a transcriptional activation protein;
 - b) a nucleotide sequence encoding a second heterologous

fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

- a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter gene prevents exhibition of a selected phenotype;
- (ii) incubating a compound with the yeast cell under conditions suitable to detect the selected phenotype; and
- (iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which prevents exhibition of the selected phenotype,

wherein said first peptide is a Dkk peptide and said second peptide is a peptide selected from LRP5, HBM, LRP6 and a Dkk-binding fragment of LRP5/LRP6/HBM.

In a preferred embodiment, the invention provides that the yeast cell is *Saccharomyces*. In a particularly preferred embodiment, the *Saccharomyces* cell is *Saccharomyces cerevisiae*. In one embodiment, the compound comprises one or more Dkk interacting or Dkk-1 interacting proteins, or a Dkk-binding fragment thereof. Compounds used in the present invention may comprise an LRP5 peptide aptamer, such as OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NOs:189-192) (particularly peptide 13 (SEQ ID NO:191)) and Figure 13 (including SEQ ID NOs:204-214), or a mimetic of such an aptamer. Alternatively, the compound may comprise LRP5 antibodies or Dkk antibodies. In another embodiment, the yeast cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a

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transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter gene, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion. In another embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In one embodiment, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In another embodiment, the DNA binding domain is a heterologous DNA-binding domain of a transcriptional activation protein.

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The invention also provides for a rescue screen for detecting the modulatory activity of a compound on the binding interaction of a first peptide and a second peptide of a peptide binding pair, comprising:

culturing at least one yeast cell, wherein the yeast cell comprises; (i)

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a nucleotide sequence encoding a first heterologous fusion a) protein comprising the first peptide or a segment thereof joined to a DNA binding domain of a transcriptional activation protein;

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a nucleotide sequence encoding a second heterologous b) fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation domain of a transcriptional activation protein;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

- C) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element prevents exhibition of a selected phenotype:
- (ii) incubating a compound with the yeast cell under conditions suitable to detect the selected phenotype; and

(iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which prevents exhibition of the selected phenotype,

wherein said first peptide is a Dkk interacting or Dkk-1 interacting protein peptide and said second peptide is a Dkk or Dkk-1 peptide.

In a preferred embodiment of the rescue screen, the yeast cell is *Saccharomyces*. In a particularly preferred embodiment, the *Saccharomyces* cell is *Saccharomyces cerevisiae*. In another embodiment, the yeast cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter gene, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion. In one embodiment, the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor. In another embodiment of the rescue screen, at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid. In another embodiment, the DNA binding domain is a heterologous DNA-binding domain of a transcriptional activation protein.

The invention also provides for a method for identifying potential compounds which modulate Dkk activity comprising:

- a) measuring the effect on binding of one or more Dkk interacting protein, or a Dkk-binding fragment thereof, with Dkk or a LRP5/LRP6/HBM binding fragment thereof in the presence and absence of a compound; and
- b) identifying as a potential Dkk modulatory compound a compound which modulates the binding between one or more Dkk interacting proteins or Dkk-binding fragment thereof and Dkk or LRP5/LRP6/HBM fragment thereof.

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In a preferred embodiment, the Dkk is Dkk-1.

The invention further provides for any of the Dkk peptide aptamers of Figure 3 (SEQ ID NOs:171-188). The invention also provides for any of the LRP peptide aptamers of Figure 4 (SEQ ID NOs:189-192).

Another aspect of the invention provides for a method of identifying agents which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) injecting mRNA encoding Dkk and an agent into a *Xenopus* blastomere;
- (b) assessing axis duplication or analyzing marker gene expression; and
- (c) identifying agents which elicit changes in axis duplication or marker gene expression as agents which modulate the interaction of Dkk with the Wnt signaling pathway. Wherein the agent may be chosen from among mRNA encoding Dkk interacting proteins, fragments thereof, siRNA, shRNA, antisense nucleotides, and antibodies. In a preferred embodiment, Dkk is Dkk-1. In a further embodiment, mRNA of HBM, LRP5/6, any Wnt (including Wnt1-Wnt19, particularly Wnt1, Wnt3, Wnt3a, and Wnt10b), Wnt antagonist, or combination of these is co-injected into the *Xenopus* blastomere. In another embodiment, the marker gene analyzed could include Siamois, Xnr3, slug, Xbra, HNK-1, endodermin, Xlhbox8, BMP2, BMP4, XLRP6, EF-1, or ODC.

The present invention provides for a method for identifying agents which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) transfecting cells with constructs encoding Dkk and potential Dkk interacting proteins, mRNA fragments thereof, siRNA, shRNA, or antisense, antibodies to LRP5/HBM/LRP6/Dkk/Dkk-interacting protein;
- (b) assessing changes in expression of a reporter gene linked to a Wntresponsive promoter; and,
- (c) identifying as a Dkk interacting protein any protein which alters reporter gene expression compared with cells transfected with a Dkk construct alone. In a further preferred embodiment, the cells may be HOB-03-CE6, HEK293, or U2OS cells.

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In alternative embodiments, the Wnt-responsive promoter is TCF or LEF. In other preferred embodiments, the cells are co-transfected with CMV beta-galactosidase or tk-Renilla.

The present invention further provides for a LRP5/HBM monoclonal or polyclonal antibody to one or more peptides of amino acid sequences MYWTDWVETPRIE (SEQ ID NO:123), MYWTDWGETPRIE (SEQ ID NO:124), KRTGGKRKEILSA (SEQ ID NO:125), ERVEKTTGDKRTRIQGR (SEQ ID NO:126), or KQQCDSFPDCIDGSDE (SEQ ID NO:127).

Additionally, the present invention provides a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
- (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and
- (c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HMB using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag.

In one embodiment, the Dkk is Dkk-1. In a preferred embodiment, the epitope tag is alkaline phosphatase, histidine, myc, or a V5 tag.

Another embodiment of the present invention provides for a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) creating an LRP5, LRP6, or HBM fluorescent fusion protein using a first fluorescent tag;
- (b) creating a Dkk fusion protein comprising a second fluorescent tag;
- (c) adding a test compound; and,
- (d) assessing changes in the ratio of fluorescent tag emissions using Fluorescence Resonance Energy Transfer (FRET) or Bioluminescent Resonance Energy Transfer (BRET) to determine whether the compound modulates Dkk and LRP5/LRP6/HBM interactions.
- In a preferred embodiment, the Dkk is Dkk-1.

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The present invention also provides for a method of diagnosing low or high bone mass and/or low or high lipid levels in a subject comprising examining expression of Dkk, LRP5, LRP6, HBM or HBM-like variant in the subject and determining whether Dkk, LRP5, LRP6, or HBM or a HBM-like variant is over- or under-expressed to determine whether subject has (a) high or low bone mass and/or (b) high or low lipid levels.

The invention further provides for a transgenic animal wherein Dkk is knocked out in a tissue-specific fashion. In a preferred embodiment, the Dkk is Dkk-1. In one preferred embodiment, the tissue specificity is bone tissue. In another preferred embodiment, the tissue specificity is liver or other tissues or cells involved in regulating lipid metabolism or cancer tissue.

The present invention further provides a method of screening for compounds which modulate the interaction of Dkk with LRP5, LRP6, or HBM comprising:

- (a) exposing LRP5, LRP6, or HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM to a compound; and
- (b) determining whether said compound bound to LRP5, LRP6, or HBM or the Dkk-binding fragment of LRP5, LRP6, or HBM and further determining whether said compound modulates the interaction of Dkk and LRP5, LRP6, or HBM.

In one embodiment, the Dkk is Dkk-1. In a preferred embodiment, the compound comprises an LRP5 peptide aptamer. Other preferred compositions include the peptide aptamer, OST262 (SEQ ID NO:208), Figure 4 (SEQ ID NO:189-192) (particularly peptide 13 (SEQ ID NO:191) and Figure 13 (including SEQ ID NO:204-214), or a mimetic of such an aptamer, and an LRP5 antibody.

The present invention also provides a method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:

- (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
- (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and

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(c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HBM using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag. In a preferred embodiment, the epitope tag is alkaline phosphatase, histidine, myc or a V5 tag.

In a preferred embodiment, the Dkk is Dkk-1.

The invention also provides for a method for identifying compounds which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) transfecting cells with constructs containing Dkk and Wnt proteins;
- (b) assessing changes in expression of a reporter element linked to a Wntresponsive promoter; and
- (c) identifying as a Dkk/Wnt interaction modulating compound any compound which alters reporter gene expression compared with cells transfected with a Dkk construct alone.

In one embodiment, the Dkk is Dkk-1. In another embodiment, the Wnt is any of Wnt1-Wnt19. In a preferred embodiment, the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b. In a particularly preferred embodiment, the Wnt construct contains Wnt3a. In another particularly preferred embodiment, the Wnt construct contains Wnt1. In another preferred embodiment, the Wnt construct encodes for a Wnt that signals through the canonical Wnt pathway. In a particularly preferred embodiment, both Wnt3a and Wnt1 constructs are co-transfected into the cells. In another embodiment, the cells may be U2-OS, HOB-03-CE6, or HEK293 cells. In another embodiment, the reporter element used is TCF-luciferase, tk-Renilla, or a combination thereof.

The invention also provides for a method of testing compounds that modulate Dkk-mediated activity in a mammal comprising:

(a) providing a group of transgenic animals having (1) a regulatable one or more Dkk genes, (2) a knock-out of Dkk genes, or (3) a knock-in of one or more Dkk genes;

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(b) providing a second group of control animals respectively for the group of transgenic animals in step (a); and

- (c) exposing the transgenic animal group and control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and
- (d) comparing the transgenic animals and the control group of animals and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.

In a preferred embodiment, the Dkk is Dkk-1.

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The invention further provides variants of LRP5 which demonstrate HBM biological activity, i.e., that are "HBM-like." In preferred embodiments, variants G171F, M282V, G171K, G171Q, A65V, G171V, G171I, and A214V of LRP5 are provided. The invention further provides for the use any of these variants in the forgoing methods.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows a schematic of the components of the Wnt signal transduction pathway. Schematic obtained from:

http://www.stanford.edu/~rnusse/pathways/cell2.html

Figure 2 (A-C) show bait sequences (SEQ ID NOs:168-170) utilized in yeast two hybrid (Y2H) screens for protein-protein interactions.

Figure 3 shows a table of peptide aptamer insert sequences (SEQ ID NOs: 171-192) identified in Y2H screen with a Dkk-1 bait sequence.

Figure 4 shows a table of peptide aptamer insert sequences identified in a Y2H screen using a LRP5 ligand binding domain bait sequence.

Figure 5 shows a table of proteins identified in a Y2H screen using a Dkk-1 bait sequence. These proteins are identified by both their nucleic acid and amino acid accession numbers.

Figure 6 shows the results of a minimum interaction domain mapping screen of Dkk-1 with LRP5. At the top, a map of Dkk-1 showing the location of the signal

sequence, and cysteine rich domains 1 and 2. Below, the extent of domains examined using LRP5 LBD baits, LBD1 and LBD4, of Figure 2. To the right, scoring of the binding results observed in the experiment.

Figure 7 shows a diagram of the Xenopus Embryo Assay for Wnt activity.

Figure 8 shows the effects of Zmax/LRP5 and HBM on Wnt signaling in the *Xenopus* embryo assay.

Figure 9 shows the effects of Zmax/LRP5 and HBM on induction of secondary axis formation in the *Xenopus* embryo assay.

Figure 10 shows the effects of human Dkk-1 on the repression of the canonical Wnt pathway.

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Figure 11 shows the effects of human Dkk-1 on Zmax/LRP5 and HBM-mediated Wnt signaling.

Figure 12 shows pcDNA3.1 construct names with nucleotide sequences (including SEQ ID NOs:193-203) for LRP5-binding peptide aptamers, Dkk-1 peptides and control constructs.

Figure 13 shows the amino acid sequences (including SEQ ID NOs:204-214) for the corresponding LRP5-binding peptides, Dkk-1 peptide aptamers and control constructs in Figure 12.

Figure 14 shows the effects of Dkk-1 and Dkk-2 on Wnt1 signaling with coreceptors LRP5, HBM, and LRP6 in HOB03CE6 cells.

Figure 15 shows the effects of Dkk-1 and Dkk-2 on Wnt3a signaling with coreceptors LRP5, HBM, and LRP6 in HOB03CE6 cells.

Figure 16 demonstrates that the LRP5-LBD peptide aptamer 262 activates Wnt signaling in the presence of Wnt3a in U2OS cells.

Figure 17 shows the differential binding of an antibody generated to a sequence (a.a. 165-177) containing the HBM mutation in LRP5 in LRP5 and HBM virus-infected cells.

Figure 18 shows data generated from a Y2H interaction trap where a mutant Dkk-1 (C220A) is unable to bind to LRP5 and demonstrating the window of capability of detecting small molecule effects on LRP and Dkk interactions.

Figure 19 shows that Dkk-1 represses Wnt3a-mediated Wnt signaling in U2OS bone cells using the cell-based reporter gene assay for high throughput screening.

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Figure 20 demonstrates that Wnt1-HBM generated signaling is not efficiently inhibited by Dkk-1 in U2OS bone cells while LRP5 and LRP6-mediated signaling are using the cell-based reporter gene assay for high throughput screening.

Figure 21 shows that the TCF signal in the cell-based reporter gene assay for high throughput screening can be modulated by Dkk-1 and Dkk-1-AP without Wnt DNA transfection.

Figure 22 shows the morphological results in the Xenopus assay using aptamers 261 and 262 from the LRP5-LBD to activate Wnt signaling.

Figure 23 demonstrates that LRP5-LBD aptamers 261 and 262 induce Wnt signaling over other LRP5 aptamers.

Figure 24 shows that the mutation G171F in LRP5 produces a greater activation of the Wnt pathway than LRP5 which is consistent with HBM activity.

Figure 25 shows that the mutation M282V in LRP5 produces an activation of the Wnt pathway which is consistent with HBM activity in U2OS cells.

Figure 26 shows the amino acid sequence of the various peptides of dkk-1 selected to generate polyclonal antibodies, their relationship to the Dkk-1 amino acid sequence and identities of polyclonal antibodies generated.

Figure 27 shows a Western blot demonstrating that polyclonal antibody #5521 to amino acids 165-186 of Dkk-1 was able to detect Dkk1-V5 and Dkk1-AP from conditioned medium.

Figure 28 shows a Western blot demonstrating that polyclonal antibody #74397 to amino acids 147-161 was able to detect Dkk1-V5 in both conditioned medium and immunoprecipitated conditioned medium.

DETAILED DESCRIPTION OF THE INVENTION

1. <u>Definitions</u>

In general, terms in the present application are used consistent with the manner in which those terms are understood in the art. To aid in the understanding of the specification and claims, the following definitions are provided.

"Gene" refers to a DNA sequence that encodes through its template or messenger RNA a sequence of amino acids characteristic of a specific peptide. The term "gene" includes intervening, non-coding regions, as well as regulatory regions, and can include 5' and 3' ends.

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By "nucleic acid" is meant to include single stranded and double stranded nucleic acids including, but not limited to DNAs, RNAs (e.g., mRNA, tRNAs, siRNAs), cDNAs, recombinant DNA (rDNA), rRNAs, antisense nucleic acids, oligonucleotides, and oligomers, and polynucleotides. The term may also include hybrids such as triple stranded regions of RNA and/or DNA or double stranded RNA:DNA hybrids. The term also is contemplated to include modified nucleic acids such as, but not limited to biotinylated nucleic acids, tritylated nucleic acids, fluorophor labeled nucleic acids, inosine, and the like.

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"Gene sequence" refers to a nucleic acid molecule, including DNA which contains a non-transcribed or non-translated sequence, which comprises a gene. The term is also intended to include any combination of gene(s), gene fragment(s), non-transcribed sequence(s) or non-translated sequence(s) which are present on the same DNA molecule.

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The nucleic acid sequences of the present invention may be derived from a variety of sources including DNA, cDNA, synthetic DNA, synthetic RNA or combinations thereof. Such sequences may comprise genomic DNA which may or may not include naturally occurring introns. Moreover, such genomic DNA may be obtained in association with promoter regions and/or poly (A) sequences. The sequences, genomic DNA or cDNA may be obtained in any of several ways. Genomic DNA can be extracted and purified from suitable cells by means well

known in the art. Alternatively, mRNA can be isolated from a cell and used to produce cDNA by reverse transcription or other means.

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"cDNA" refers to complementary or copy DNA produced from an RNA template by the action of RNA-dependent DNA polymerase (reverse transcriptase). Thus, a "cDNA clone" means a duplex DNA sequence for which one strand is complementary to an RNA molecule of interest, carried in a cloning vector or PCR amplified. cDNA can also be single stranded after first strand synthesis by reverse transcriptase. In this form, it is a useful PCR template and does not need to be carried in a cloning vector. This term includes genes from which the intervening sequences have been removed. Thus, the term "gene", as sometimes used generically, can also include nucleic acid molecules comprising cDNA and cDNA clones.

"Recombinant DNA" means a molecule that has been engineered by splicing in vitro a cDNA or genomic DNA sequence or altering a sequence by methods such as PCR mutagenesis.

"Cloning" refers to the use of *in vitro* recombination techniques to insert a particular gene or other DNA sequence into a vector molecule. In order to successfully clone a desired gene, it is necessary to use methods for generating DNA fragments, for joining the fragments to vector molecules, for introducing the composite DNA molecule into a host cell in which it can replicate, and for selecting the clone having the target gene from amongst the recipient host cells.

"cDNA library" refers to a collection of recombinant DNA molecules containing cDNA inserts which together comprise the entire or a partial repertoire of genes expressed in a particular tissue or cell source. Such a cDNA library can be prepared by methods known to one skilled in the art and described by, for example, Cowell and Austin, "cDNA Library Protocols," *Methods in Molecular Biology* (1997).

"Cloning vehicle" refers to a plasmid or phage DNA or other DNA sequence which is able to replicate in a host cell. This term can also include artificial chromosomes such as BACs and YACs. The cloning vehicle is characterized by one or more endonuclease recognition sites at which such DNA sequences may be cut in

a determinable fashion without loss of an essential biological function of the DNA, which may contain a marker suitable for use in the identification of transformed cells.

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"Expression" refers to the process comprising transcription of a gene sequence and subsequent processing steps, such as translation of a resultant mRNA to produce the final end product of a gene. The end product may be a protein (such as an enzyme or receptor) or a nucleic acid (such as a tRNA, antisense RNA, or other regulatory factor). The term "expression control sequence" refers to a sequence of nucleotides that control or regulate expression of structural genes when operably linked to those genes. These include, for example, the lac systems, the trp system, major operator and promoter regions of the phage lambda, the control region of fd coat protein and other sequences known to control the expression of genes in prokaryotic or eukaryotic cells. Expression control sequences will vary depending on whether the vector is designed to express the operably linked gene in a prokaryotic or eukaryotic host, and may contain transcriptional elements such as enhancer elements, termination sequences, tissue-specificity elements and/or translational initiation and termination sites.

"Expression vehicle" refers to a vehicle or vector similar to a cloning vehicle but which is capable of expressing a gene which has been cloned into it, after transformation into a host. The cloned gene is usually placed under the control of (i.e., operably linked to) an expression control sequence.

"Operator" refers to a DNA sequence capable of interacting with the specific repressor, thereby controlling the transcription of adjacent gene(s).

"Promoter" refers to a DNA sequence that can be recognized by an RNA polymerase. The presence of such a sequence permits the RNA polymerase to bind and initiate transcription of operably linked gene sequences.

"Promoter region" is intended to include the promoter as well as other gene sequences which may be necessary for the initiation of transcription. The presence of a promoter region is sufficient to cause the expression of an operably linked gene sequence. The term "promoter" is sometimes used in the art to generically indicate a promoter region. Many different promoters are known in the art which direct

expression of a gene in a certain cell types. Tissue-specific promoters can comprise nucleic acid sequences which cause a greater (or decreased) level of expression in cells of a certain tissue type.

"Operably linked" means that the promoter controls the initiation of expression of the gene. A promoter is operably linked to a sequence of proximal DNA if upon introduction into a host cell the promoter determines the transcription of the proximal DNA sequence(s) into one or more species of RNA. A promoter is operably linked to a DNA sequence if the promoter is capable of initiating transcription of that DNA sequence.

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"Prokaryote" refers to all organisms without a true nucleus, including bacteria.

"Eukaryote" refers to organisms and cells that have a true nucleus, including mammalian cells.

"Host" includes prokaryotes and eukaryotes, such as yeast and filamentous fungi, as well as plant and animal cells. The term includes an organism or cell that is the recipient of a replicable expression vehicle.

The term "animal" is used herein to include all vertebrate animals, except humans. It also includes an individual animal in all stages of development, including embryonic and fetal stages. Preferred animals include higher eukaryotes such as avians, rodents (e.g., mice, rabbits, rats, chinchillas, guinea pigs, hamsters and the like), and mammals. Preferred mammals include bovine, equine, feline, canine, ovine, caprine, porcine, buffalo, humans, and primates.

A "transgenic animal" is an animal containing one or more cells bearing genetic information received, directly or indirectly, by deliberate genetic manipulation or by inheritance from a manipulated progenitor at a subcellular level, such as by microinjection or infection with a recombinant viral vector (e.g., adenovirus, retrovirus, herpes virus, adeno-associated virus, lentivirus). This introduced DNA molecule may be integrated within a chromosome, or it may be extra-chromosomally replicating DNA.

"Embryonic stem cells" or "ES cells" as used herein are cells or cell lines usually derived from embryos which are pluripotent meaning that they are un-

differentiated cells. These cells are also capable of incorporating exogenous DNA by homologous recombination and subsequently developing into any tissue in the body when incorporated into a host embryo. It is possible to isolate pluripotent cells from sources other than embryonic tissue by methods which are well understood in the art.

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Embryonic stem cells in mice have enabled researchers to select for transgenic cells and perform gene targeting. This allows more genetic engineering than is possible with other transgenic techniques. For example, mouse ES cells are relatively easy to grow as colonies *in vitro*. The cells can be transfected by standard procedures and transgenic cells clonally selected by antibiotic resistance. See, for example, Doetschman *et al..*, 1994, *Gene transfer in embryonic stem cells*. In Pinkert (Ed.) <u>Transgenic Animal Technology: A Laboratory Handbook</u>. Academic Press Inc., New York, pp.115-146. Furthermore, the efficiency of this process is such that sufficient transgenic colonies (hundreds to thousands) can be produced to allow a second selection for homologous recombinants. Mouse ES cells can then be combined with a normal host embryo and, because they retain their potency, can develop into all the tissues in the resulting chimeric animal, including the germ cells. The transgenic modification can then be transmitted to subsequent generations.

Methods for deriving embryonic stem (ES) cell lines *in vitro* from early preimplantation mouse embryos are well known. See for example, Evans *et al.*, 1981 *Nature* 29: 154-6 and Martin, 1981, *Proc. Nat. Acad. Sci. USA*, 78: 7634-8. ES cells can be passaged in an undifferentiated state, provided that a feeder layer of fibroblast cells or a differentiation inhibiting source is present.

The term "somatic cell" indicates any animal or human cell which is not a sperm or egg cell or is capable of becoming a sperm or egg cell. The term "germ cell" or "germ-line cell" refers to any cell which is either a sperm or egg cell or is capable of developing into a sperm or egg cell and can therefore pass its genetic information to offspring. The term "germ cell-line transgenic animal" refers to a transgenic animal in which the genetic information was incorporated in a germ line cell, thereby conferring the ability to transfer the information to offspring. If such

offspring in fact possess some or all of that information, then they, too, are transgenic animals.

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The genetic alteration of genetic information may be foreign to the species of animal to which the recipient belongs, or foreign only to the particular individual recipient. In the last case, the altered or introduced gene may be expressed differently than the native gene.

"Fragment" of a gene refers to any portion of a gene sequence. A
"biologically active fragment" refers to any portion of the gene that retains at least
one biological activity of that gene. For example, the fragment can perhaps
hybridize to its cognate sequence or is capable of being translated into a polypeptide
fragment encoded by the gene from which it is derived.

"Variant" refers to a gene that is substantially similar in structure and biological activity or immunological characteristics to either the entire gene or to a fragment of the gene. Provided that the two genes possess a similar activity, they are considered variant as that term is used herein even if the sequence of encoded amino acid residues is not identical. Preferentially, as used herein (unless otherwise defined) the variant is one of LRP5, HBM or LRP6. The variant preferably is one that yields an HBM-like phenotype (i.e., enhances bones mass and/or modulates lipid levels). These variants include missense mutations, single nucleotide polymorphisms (SNPs), mutations which result in changes in the amino acid sequence of the protein encoded by the gene or nucleic acid, and combinations thereof, as well as com in the exon domains of the HBM gene and mutations in LRP5 or LRP6 which result in an HBM like phenotype.

"Amplification of nucleic acids" refers to methods such as polymerase chain reaction (PCR), ligation amplification (or ligase chain reaction, LCR) and amplification methods based on the use of Q-beta replicase. These methods are well known in the art and described, for example, in U.S. Patent Nos. 4,683,195 and 4,683,202. Reagents and hardware for conducting PCR are commercially available. Primers useful for amplifying sequences from the HBM region are preferably complementary to, and hybridize specifically to sequences in the HBM region or in

regions that flank a target region therein. HBM sequences generated by amplification may be sequenced directly. Alternatively, the amplified sequence(s) may be cloned prior to sequence analysis.

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"Antibodies" may refer to polyclonal and/or monoclonal antibodies and fragments thereof, and immunologic binding equivalents thereof, that can bind to the HBM proteins and fragments thereof or to nucleic acid sequences from the HBM region, particularly from the HBM locus or a portion thereof. Preferred antibodies also include those capable of binding to LRP5, LRP6 and HBM variants. The term antibody is used both to refer to a homogeneous molecular entity, or a mixture such as a serum product made up of a plurality of different molecular entities. Proteins may be prepared synthetically in a protein synthesizer and coupled to a carrier molecule and injected over several months into rabbits. Rabbit sera is tested for immunoreactivity to the HBM protein or fragment. Monoclonal antibodies may be made by injecting mice with the proteins, or fragments thereof. Monoclonal antibodies will be screened by ELISA and tested for specific immunoreactivity with HBM protein or fragments thereof. Harlow et al., Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1988) and Using Antibodies: A Laboratory Manual, Harlow, Ed and Lane, David (Cold Spring Harbor Press, 1999). These antibodies will be useful in assays as well as pharmaceuticals. By "antibody" is meant to include but not limited to polyclonal, monoclonal, chimeric. human, humanized, bispecific, multispecific, primatized™ antibodies.

"HBM protein" refers to a protein that is identical to a Zmax1 (LRP5) protein except that it contains an alteration of glycine 171 to a valine. An HBM protein is defined for any organism that encodes a Zmax1 (LRP5) true homolog. For example, a mouse HBM protein refers to the mouse Zmax1 (LRP5) protein having the glycine 170 to valine substitution.

By "HBM-like" is meant a variant of LRP5, LRP6 or HBM which when expressed in a cell is capable of modulating bone mass, lipid levels, Dkk activity, and/or Wnt activity.

In one embodiment of the present invention, "HBM gene" refers to the genomic DNA sequence found in individuals showing the HBM characteristic or phenotype, where the sequence encodes the protein indicated by SEQ ID NO: 4. The HBM gene and the Zmax1 (LRP5) gene are allelic. The protein encoded by the HBM gene has the property of causing elevated bone mass, while the protein encoded by the Zmax1 (LRP5) gene does not. The HBM gene and the Zmax1 (LRP5) gene differ in that the HBM gene has a thymine at position 582, while the Zmax1 gene has a guanine at position 582. The HBM gene comprises the nucleic acid sequence shown as SEQ ID NO: 2. The HBM gene may also be referred to as an "HBM polymorphism." Other HBM genes may further have silent mutations, such as those discussed in Section 3 below.

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In alternative embodiments of the present invention, "HBM gene" may also refer to any allelic variant of Zmax1 (LRP5) or LRP6 which results in the HBM phenotype. Such variants may include alteration from the wild-type protein coding sequence as described herein and/or alteration in expression control sequences of Zmax1 (LRP5) or contains an amino acid mutation in LRP5 or LRP6, such that the resulting protein produces a phenotype which enhances bone mass and/or modulates lipid levels. A preferred example of such a variant is an alteration of the endogenous Zmax1 (LRP5) promoter region resulting in increased expression of the Zmax1 (LRP5) protein.

"Normal," "wild-type," "unaffected", "Zmax1", "Zmax", "LR3" and "LRP5" all refer to the genomic DNA sequence that encodes the protein indicated by SEQ ID NO: 3. LRP5 has also been referred to LRP7 in mouse. Zmax1, LRP5 and Zmax may be used interchangeably throughout the specification and are meant to be the same gene, perhaps only relating to the gene in a different organism. The Zmax1 gene has a guanine at position 582 in the human sequence. The Zmax1 gene of human comprises the nucleic acid sequence shown as SEQ ID NO: 1. "Normal," "wild-type," "unaffected", "Zmax1" and "LRP5" also refer to allelic variants of the genomic sequence that encodes proteins that do not contribute to elevated bone

mass. The *Zmax1* (*LRP5*) gene is common in the human population, while the *HBM* gene is rare.

"Bone development" generally refers to any process involved in the change of bone over time, including, for example, normal development, changes that occur during disease states, and changes that occur during aging. This may refer to structural changes and dynamic rate changes such as growth rates, resorption rates, bone repair rates, and etc. "Bone development disorder" particularly refers to any disorders in bone development including, for example, changes that occur during disease states and changes that occur during aging. Bone development may be progressive or cyclical in nature. Aspects of bone that may change during development include, for example, mineralization, formation of specific anatomical features, and relative or absolute numbers of various cell types.

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"Bone modulation" or "modulation of bone formation" refers to the ability to affect any of the physiological processes involved in bone remodeling, as will be appreciated by one skilled in the art, including, for example, bone resorption and appositional bone growth, by, *inter alia*, osteoclastic and osteoblastic activity, and may comprise some or all of bone formation and development as used herein.

Bone is a dynamic tissue that is continually adapting and renewing itself through the renewal of old or unnecessary bone by osteoclasts and the rebuilding of new bone by osteoblasts. The nature of the coupling between these processes is responsible for both the modeling of bone during growth as well as the maintenance of adult skeletal integrity through remodeling and repair to meet the everyday needs of mechanical usage. There are a number of diseases that result from an uncoupling of the balance between bone resorption and formation. With aging there is a gradual "physiologic" imbalance in bone turnover, which is particularly exacerbated in women due to menopausal loss of estrogen support, that leads to a progressive loss of bone. As bone mineral density falls below population norms there is a consequent increase in bone fragility and susceptibility to spontaneous fractures. For every 10 percent of bone that is lost, the risk of fracture doubles. Individuals with bone mineral density (BMD) in the spine or proximal femur 2.5 or

more standard deviations below normal peak bone mass are classified as osteoporotic. However, osteopenic individuals with BMD between 1 and 2.5 standard deviations below the norm are clearly at risk.

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Bone is measured by several different forms of X-ray absorptiometry. All of the instruments measure the inorganic or bone mineral content of the bone. Standard DXA measurements give a value that is an areal density, not a true density measurement by the classical definition of density (mass/unit volume). Nevertheless, this is the type of measurement used clinically to diagnose osteoporosis. However, while BMD is a major contributing factor to bone strength, as much as 40% of bone strength stems from other factors including: 1) bone size (i.e., larger diameters increase organ-level stiffness, even in the face of lower density); 2) the connectivity of trabecular structures; 3) the level of remodeling (remodeling loci are local concentrators of strain); and 4) the intrinsic strength of the bony material itself, which in turn is a function of loading history (i.e., through accumulated fatigue damage) and the extent of collagen cross-linking and level of mineralization. There is good evidence that all of these strength/fragility factors play some role in osteoporotic fractures, as do a host of extraskeletal influences as well (such as fall patterns, soft tissue padding, and central nervous system reflex responsiveness).

Additional analytical instruments can be used to address these features of bone. For example, the pQCT allows measurement of separate trabecular and cortical compartments for size and density and the μ CT provides quantitative information on architectural features such as trabecular connectivity. The μ CT also gives a true bone density measurement. With these tools, the important non-BMD parameters can be measured for diagnosing the extent of disease and the efficacy of treatments. Current treatments for osteoporosis are based on the ability of drugs to prevent or retard bone resorption. Although newer anti-resorptive agents are proving to be useful in the therapy of osteoporosis, they are viewed as short-term solutions to the more definitive challenge to develop treatments that will increase bone mass and/or the bone quality parameters mentioned above.

Thus, bone modulation may be assessed by measuring parameters such as bone mineral density (BMD) and bone mineral content (BMC) by pDXA X-ray methods. bone size, thickness or volume as measured by X-ray, bone formation rates as measured for example by calcien labeling, total, trabecular, and mid-shaft density as measured by pQCT and/or μ CT methods, connectivity and other histological parameters as measured by μ CT methods, mechanical bending and compressive strengths as preferably measured in femur and vertebrae respectively. Due to the nature of these measurements, each may be more or less appropriate for a given situation as the skilled practitioner will appreciate. Furthermore, parameters and methodologies such as a clinical history of freedom from fracture, bone shape, bone morphology, connectivity, normal histology, fracture repair rates, and other bone quality parameters are known and used in the art. Most preferably, bone quality may be assessed by the compressive strength of vertebra when such a measurement is appropriate. Bone modulation may also be assessed by rates of change in the various parameters. Most preferably, bone modulation is assessed at more than one age.

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"Normal bone density" refers to a bone density within two standard deviations of a Z score of 0 in the context of the HBM linkage study. In a general context, the range of normal bone density parameters is determined by routine statistical methods. A normal parameter is within about 1 or 2 standard deviations of the age and sex normalized parameter, preferably about 2 standard deviations. A statistical measure of meaningfulness is the P value which can represent the likelihood that the associated measurement is significantly different from the mean. Significant P values are P < 0.05, 0.01, 0.005, and 0.001, preferably at least P < 0.01.

"HBM" refers to "high bone mass" although this term may also be expressed in terms of bone density, mineral content, and size.

The "HBM phenotype" and "HBM-like phenotype" may be characterized by an increase of about 2 or more standard deviations, preferably 2, 2.5, 3, or more standard deviations in 1, 2, 3, 4, 5, or more quantitative parameters of bone modulation, preferably bone density and mineral content and bone strength

parameters, above the age and sex norm for that parameter. The HBM phenotype and HBM-like phenotype are characterized by statistically significant increases in at least one parameter, preferably at least 2 parameters, and more preferably at least 3 or more parameters. The HBM phenotype and the HBM-like phenotype may also be characterized by an increase in one or more bone quality parameters and most preferably increasing parameters are not accompanied by a decrease in any bone quality parameters. Most preferably, an increase in bone modulation parameters and/or bone quality measurements is observed at more than one age. The HBM phenotype and HBM-like phenotype also includes changes of lipid levels, Wnt activity and/or Dkk activity.

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The terms "isolated" and "purified" refer to a substance altered by hand of man from the natural environment. An isolated peptide may be for example in a substantially pure form or otherwise displaced from its native environment such as by expression in an isolated cell line or transgenic animal. An isolated sequence may for example be a molecule in substantially pure form or displaced from its native environment such that at least one end of said isolated sequence is not contiguous with the sequence it would be contiguous with in nature.

"Biologically active" refers to those forms of proteins and polypeptides, including conservatively substituted variants, alleles of genes encoding a protein or polypeptide fragments of proteins which retain a biological and/or immunological activity of the wild-type protein or polypeptide. Preferably the activity is one which induces a change in Dkk activity, such as inhibiting the interaction of Dkk with a ligand binding partner (e.g., LRP5 or LRP6 or Dkk-1 with a Dkk-1 interacting protein such as those shown in Figure 5). By biologically active is also meant to include any form which modulates Wnt signaling.

By "modulate" and "regulate" is meant methods, conditions, or agents which increase or decrease the wild-type activity of an enzyme, inhibitor, signal transducer, receptor, transcription activator, co-factor, and the like. This change in activity can be an increase or decrease of mRNA translation, mRNA or DNA transcription, and/or

mRNA or protein degradation, which may in turn correspond to an increase or decrease in biological activity.

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By "modulated activity" is meant any activity, condition, disease or phenotype which is modulated by a biologically active form of a protein. Modulation may be effected by affecting the concentration or subcellular localization of biologically active protein, *i.e.*, by regulating expression or degradation, or by direct agonistic or antagonistic effect as, for example, through inhibition, activation, binding, or release of substrate, modification either chemically or structurally, or by direct or indirect interaction which may involve additional factors.

By "effective amount" or "dose effective amount" or "therapeutically effective amount" is meant an amount of an agent which modulates a biological activity of the polypeptide of the invention.

By "immunologically active" is meant any immunoglobulin protein or fragment thereof which recognizes and binds to an antigen.

By "Dkk" is meant to refer to the nucleic acids and proteins of members of the Dkk (Dickkopf) family. This includes, but is not limited to, Dkk-1, Dkk-2, Dkk-3, Dkk-4, Soggy, and related Dkk proteins. Dkk-1 is a preferred embodiment of the present invention. However, the Dkk proteins have substantial homology and one skilled in the art will appreciate that all of the embodiments of the present invention utilizing Dkk-1 may also be utilized with the other Dkk proteins.

By "Dkk-1" is meant to refer to the Dkk-1 protein and nucleic acids which encode the Dkk-1 protein. Dkk-1 refers to Dickkopf-1, and in *Xenopus* it is related to at least Dkk-2, Dkk-3, and Dkk-4 (see Krupnik et al., Gene 238:301-313 (1999)). Dkk-1 was first identified in *Xenopus* (Glinka et al., Nature 391:357-62 (1998)). It was recognized as a factor capable of inducing ectopic head formation in the presence of inhibition of the BMP pathway. It was then also found to inhibit the axis-inducing activity of several *Xenopus* Wnt molecules by acting as an extracellular antagonist of Wnt signaling. Mammalian homologs have been found including Dkk-1, Dkk-2, Dkk-3, Dkk-4 and soggy (Fedi et al., 1999 and Krupnick et al. 1999). Human Dkk-1 was also referred to as sk (Fedi et al. 1999). As used herein, Dkk-1 is

meant to include proteins from any species having a Wnt pathway in which Dkk-1 interacts. Particularly preferred are mammalian species (e.g., murine, caprine, canine, bovine, feline, equine, primate, ovine, porcine and the like), with particularly preferred mammals being humans. Nucleic acid sequences encoding Dkk-1 include, but are not limited to human Dkk-1 (GenBank Accession Nos. AH009834, XM 005730, AF261158, AF261157, AF177394, AF127563 and NM_012242), Mus musculus dickkopf homolog 1 (GenBank Accession No. NM 010051), and Danio rerio dickkopf-1 (GenBank Accession Nos. AF116852 and AB023488). The genomic sequences with exon annotation are GenBank Accession Nos. AF261157 and AF261158. Also contemplated are homologs of these sequences which have Dkk-1 activity in the Wnt pathway. Dkk-1 amino acid sequences include, but are not limited to human dickkopf homolog 1 (GenBank Accession Nos. AAG15544, BAA34651, NP 036374, AAF02674, AAD21087, and XP 005730), Danio rerio (zebrafish) dickkopf1 (GenBank Accession Nos. BAA82135 and AAD22461) and murine dickkopf-1 (GenBank Accession Nos. 054908 and NP 034181). Variants and homologs of these sequences which possess Dkk-1 activity are also included when referring to Dkk-1.

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By "Dkk mediated" disorder, condition or disease is any abnormal state that involves Dkk activity. The abnormal state can be induced by environmental exposure or drug administration. Alternatively, the disease or disorder can be due to a genetic defect. Dkk mediated diseases, disorders and conditions include but are not limited to bone mass disorders or conditions and lipid disorders and conditions. For example, bone mass disorders/conditions/diseases, which may be mediated by Dkk, include but are not limited to age related loss of bone, bone fractures (e.g., hip fracture, Colle's fracture, vertebral crush fractures), chondrodystrophies, druginduced disorders (e.g., osteoporosis due to administration of glucocorticoids or heparin and osteomalacia due to administration of aluminum hydroxide, anticonvulsants, or glutethimide), high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and rickets.

Lipid disorders/diseases/conditions, which may be mediated by Dkk, include but are not limited to familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid levels due to dialysis and/or diabetes, and elevated lipid levels of unknown etiologies

The term "recognizes and binds," when used to define interactions of antisense nucleotides, siRNAs (small inhibitory RNA), or shRNA (short hairpin RNA) with a target sequence, means that a particular antisense, siRNA, or shRNA sequence is substantially complementary to the target sequence, and thus will specifically bind to a portion of an mRNA encoding polypeptide. As such, typically the sequences will be highly complementary to the mRNA target sequence, and will have no more than 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 base mismatches throughout the sequence. In many instances, it may be desirable for the sequences to be exact matches, i.e. be completely complementary to the sequence to which the oligonucleotide specifically binds, and therefore have zero mismatches along the complementary stretch. As such, highly complementary sequences will typically bind quite specifically to the target sequence region of the mRNA and will therefore be highly efficient in reducing, and/or even inhibiting the translation of the target mRNA sequence into polypeptide product.

Substantially complementary oligonucleotide sequences will be greater than about 80 percent complementary (or `% exact-match`) to the corresponding mRNA target sequence to which the oligonucleotide specifically binds, and will, more preferably be greater than about 85 percent complementary to the corresponding mRNA target sequence to which the oligonucleotide specifically binds. In certain aspects, as described above, it will be desirable to have even more substantially complementary oligonucleotide sequences for use in the practice of the invention, and in such instances, the oligonucleotide sequences will be greater than about 90 percent complementary to the corresponding mRNA target sequence to which the oligonucleotide specifically binds, and may in certain embodiments be greater than about 95 percent complementary to the corresponding mRNA target sequence to

which the oligonucleotide specifically binds, and even up to and including 96%, 97%, 98%, 99%, and even 100% exact match complementary to the target mRNA to which the designed oligonucleotide specifically binds.

Percent similarity or percent complementary of any of the disclosed sequences may be determined, for example, by comparing sequence information using the GAP computer program, version 6.0, available from the University of Wisconsin Genetics Computer Group (UWGCG). The GAP program utilizes the alignment method of Needleman and Wunsch (1970). Briefly, the GAP program defines similarity as the number of aligned symbols (i.e., nucleotides or amino acids) which are similar, divided by the total number of symbols in the shorter of the two sequences. The preferred default parameters for the GAP program include: (1) a unary comparison matrix (containing a value of 1 for identities and 0 for non-identities) for nucleotides, and the weighted comparison matrix of Gribskov and Burgess (1986), (2) a penalty of 3.0 for each gap and an additional 0.10 penalty for each symbol in each gap; and (3) no penalty for end gaps.

By "mimetic" is meant a compound or molecule that performs the same function or behaves similarly to the compound mimicked.

By "reporter element" is meant a polynucleotide that encodes a poplypeptide capable of being detected in a screening assays. Examples of polypeptides encoded by reporter elements include, but are not limited to, lacZ, GFP, luciferase, and chloramphenicol acetyltransferase.

2. <u>Introduction</u>

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A polymorphism in LRP5 (Zmax), G171V, designated as HBM, has been identified as conferring a high bone mass phenotype in a population of related subjects as described in co-pending applications International Patent Application PCT/US 00/16951, and U.S. Patent Application Nos. 09/543,771 and 09/544,398, which are hereby incorporated by reference in their entirety (Little *et al., Am J Hum Genet.* 70:11-19 (2002)). LRP5 is also described in International Patent Application WO 98/46743, which is incorporated by reference in its entirety. Loss of LRP5

function has been shown to have a deleterious effect on bone (Gong *et al., Cell* 107:513-523 (2001)). Additionally, the HBM polymorphism and LRP5 may also be important in cardiac health and lipid-mediated disorders. Thus, methods of regulating their activity can serve as methods of treating and/or preventing cardiac and lipid-mediated disorders.

Recent studies have indicated that LRP5 participates in the Wnt signal transduction pathway. The Wnt pathway is critical in limb early embryological development. A recently published sketch of the components of Wnt signaling is shown in Figure 1

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(Nusse, 2001 http://www.stanford.edu/~rnusse/pathways/cell2.html) (see also, Nusse, Nature 411:255-6 (2001); and Mao et al., Nature 411:321-5 (2001)). Briefly summarized, Wnt proteins are secreted proteins which interact with the transmembrane protein Frizzled (Fz). LRP proteins, such as LRP5 and LRP6, are believed to modulate the Wnt signal in a complex with Fz (Tamai et al., Nature 407:530-5 (2000)). The Wnt pathway acts intracellularly through the Disheveled protein (Dsh) which in turn inhibits glycogen synthetase kinase-3 (GSK3) from phosphorylating β-catenin. Phosphorylated β-catenin is rapidly degraded following ubiquitination. However, the stabilized β-catenin accumulates and translocates to the nucleus where it acts as a cofactor of the T-cell factor (TCF) transcription activator complex.

The protein dickkopf-1 (Dkk-1) is reported to be an antagonist of Wnt pathway. Dkk-1 is required for head formation in early development. Dkk-1 and its function in the Wnt pathway are described in e.g., Krupnik, et al., Gene 238:301-13 (1999); Fedi et al., J. Biol. Chem. 274:19465-72 (1999); see also for Dkk-1 and the Wnt pathway, Wu et al., Curr. Biol. 10:1611-4 (2000), Shinya et al., Mech. Dev. 98:3-17 (2000), Mukhopadhyay et al., Dev Cell 1:423-434 (2001) and in PCT Patent Application No. WO 00/52047, and in references cited in each. It has been known that Dkk-1 acts upstream of Dsh, however the nature of the mechanism of inhibition by Dkk-1 is just beginning to be elucidated. Dkk-1 is expressed in the mouse embryonic limb bud and its disruption results in abnormal limb morphogensis, among

other developmental defects (Gotewold et al., Mech. Dev. 89:151-3 (1999); and, Mukhopadhyay et al., Dev Cell 1:423-434 (2001)).

Related U.S. provisional application 60/291,311 disclosed a novel interaction between Dkk-1 (GenBank Accession No. XM 005730) and LRP5. The interaction between Dkk-1 and LRP5 was discovered by a yeast two hybrid (Y2H) screen for proteins which interact with the ligand binding domain of LRP5, as described in Example 1. The two-hybrid screen is a common procedure in the art, which is described, for example, by Gietz *et al.*, *Mol. Cell. Biochem.* 172:67-79 (1997); Young, *Biol. Reprod.* 58:302-11 (1998); Brent and Finley, *Ann. Rev. Genet.* 31:663-704 (1997); and Lu and Hannon, eds., <u>Yeast Hybrid Technologies</u>, Eaton Publishing, Natick MA, (2000). More recently, other studies confirm that Dkk-1 is a binding partner for LRP and modulates the Wnt pathway via direct binding with LRP (R. Nusse, *Nature* 411:255-256 (2001); A. Bafico *et al.*, *Nat. Cell Biol.* 3:683-686 (2001); M. Semënov, *Curr. Biol.* 11:951-961 (2001); B. Mao, *Nature* 411:321-325 (2001), Zorn, *Curr. Biol.* 11:R592-5 (2001)); and, L. Li *et al.*, *J. Biol Chem.* 277:5977-81 (2002)).

Mao and colleagues (2001) identified Dkk-1 as a ligand for LRP6. Mao *et al.* suggest that Dkk-1 and LRP6 interact antagonistically where Dkk proteins inhibit the Wnt coreceptor functions of LRP6. Using co-immunoprecipitation, the group verified that the Dkk-1/LRP6 interaction was direct. Dkk-2 was also found to directly bind LRP6. Contrary to data contained in provisional application 60/291,311, Mao *et al.* report that no interaction was detected between any Dkk protein and LRP5, as well as no interaction with LDLR, VLDLR, ApoER, or LRP). Additionally, Mao *et al.* demonstrated that LRP6 can titrate Dkk-1's effects of inhibiting Wnt signaling using the commercial TCF-luciferase reporter gene assay (TOPFLASH). A similar conclusion was drawn from analogous studies in *Xenopus* embryos. Deletion analyses of LRP6 functional domains revealed that EGF repeats (beta-propellers) 3 and 4 were necessary for Dkk-1 binding and that the ligand binding domains of LRP6 had no effect on Dkk-1 binding. The findings of Mao *et al.* contrast with data obtained by the present inventors indication that the ligand binding domains of LRP5

were necessary and sufficient for Dkk-1 binding in yeast. Using classical biochemical ligand-receptor studies, Mao *et al.* determined a Kd=0.34 nM for Dkk-1/LRP6 and a Kd=0.73 nM for Dkk-2/LRP6.

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Semenov *et al.* (2001) verified the Mao group's results and confirmed by coimmunoprecipitation that Dkk-1 does not directly bind to Wnt or Frizzled but rather interacts with LRP6. Their Scatchard analyses found a Kd=0.5 nM for Dkk-1/LRP6. Semenov *et al.* also demonstrated that Dkk-1 could abolish an LRP5/Frizzled8 complex implying that Dkk-1 can also repress Wnt signaling via interactions with LRP5. A Dkk-1 mutant where cysteine 220 was changed to alanine abolished LRP6 binding and was unable to repress Wnt signaling. Studies in *Xenopus* embryos confirmed the results and revealed a functional consequence of Dkk-1/LRP6: repression of Wnt signaling. Their *Xenopus* work also suggested that LRP6/Dkk-1 may be specific for the canonical, β-catenin-mediated, Wnt pathways as opposed to the Wnt Planar Cell Polarity pathway.

Bafico *et al.* (2001) employed a ¹²⁵I-labeled Dkk-1 molecule to identify LRP6 as its sole membrane receptor with a Kd=0.39 nM. Again, the functional consequences of the Dkk-1/LRP6 interaction was a repression of the canonical Wnt signaling even when Dkk-1 was added at extremely low concentrations (30 pM).

Not wishing to be bound by theory, it is believed that the present invention provides an explanation for the mechanism of Dkk-1 inhibition of the Wnt pathway and provides a mechanism whereby the Wnt pathway may be modulated. The present application and related provisional application 60/291,311 describe Dkk-1/LRP5 interactions and demonstrate that the interaction between LRP5/LRP6/HBM and Dkk can be used in a method as an intervention point in the Wnt pathway for an anabolic bone therapeutic or a modulator of lipid metabolism.

As detailed below, in the section "Methods to Identify Binding Partners" and Examples 6 and 7, Dkk-1 is able to repress LRP5-mediated Wnt signaling but not HBM-mediated Wnt signaling. This observation is of particular interest because the HBM mutation in LRP5 is a gain of function or activation mutation. That is, Wnt signaling, via the canonical pathway, is enhanced with HBM versus LRP5. The

present data suggest the mechanism of this functional activation: the inability of Dkk-1 to repress HBM-mediated Wnt signaling. Further investigations of other Wnt or Dkk family members show differential activities in the canonical Wnt pathway that demonstrate the complexity and variability in Wnt signaling that can be achieved depending on the LRP/Dkk/Wnt/Frizzled repertoire that is expressed in a particular cell or tissue. This may attest to the apparent bone specificity of the HBM phenotype in humans and in the HBM transgenic animals.

Furthermore, the present data reveal the importance and functional consequence for the potential structural perturbation of the first beta-propeller domain of LRP5. Our data identified the ligand binding domain of LRP5 as the interacting region with Dkk-1 while the Mao *et al.* publication demonstrated the functional role of propellers 3 and 4 in their LRP6/Dkk-1 studies. In the present invention, we implicate the first beta propeller domain, via the HBM mutation at residue 171, as having a functional consequence in the Dkk-1-mediated Wnt pathway. The involvement of position 171 of propeller 1 may be direct or indirect with Dkk-1. Direct involvement could arise from perturbations of the 3-dimensional structure of the HBM extracellular domain that render Dkk-1 unable to bind. Alternatively, residue 171 of propeller 1 may directly interact with Dkk-1; however, by itself, it is insufficient to bind and requires other LRP5 domains. Potential indirect candidate molecules may be among the proteins identified the Dkk-1 yeast-two-hybrid experiments.

It may be that the disruption of Dkk activity is not necessarily mediated by enhancing or preventing the binding of Dkk to LRP5/LRP6/HBM. More than one mechanism may be involved. Indeed, the inventors have observed that Dkk-1 binds LRP5, LRP6, and HBM. It is able to effectively inhibit LRP6, and to a slightly lesser extent, LRP5 activity. Further, has been observed that different members of the Dkk family differentially affect LRP5/LRP6/HBM activity. For example, Dkk-1 inhibits LRP5/LRP6/HBM activity while another Dkk may enhance LRP5/LRP6/HBM activity. An endpoint to consider is the modulation of the LRP5/LRP6/HBM activity, not simply binding.

The present disclosure shows that targeting the disruption of the Dkk-1/LRP5 interaction is a therapeutic intervention point for an HBM mimetic agent. A therapeutic agent of the invention may be a small molecule, peptide or nucleic acid aptamer, antibody, or other peptide/protein, etc. Methods of reducing Dkk-1 expression may also be therapeutic using methodologies such as: RNA interference, antisense oligonucleotides, morpholino oligonucleotides, PNAs, antibodies to Dkk-1 or Dkk-1 interacting proteins, decoy or scavenger LRP5 or LRP6 receptors, and knockdown of Dkk-1 or Dkk-1 interactor transcription.

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In an embodiment of the present invention, the activity of Dkk-1 or the activity of a Dkk-1 interacting protein may be modulated for example by binding with a peptide aptamer of the present invention. In another embodiment, LRP5 activity may be modulated by a reagent provided by the present invention (e.g., a peptide aptamer). In another embodiment, the Dkk-1/LRP5 interaction may be modulated by a reagent of the present invention (e.g., a Dkk-1 interacting protein such as those identified in Figure 5). In another embodiment, the Wnt signal transduction pathway may be modulated by use of one or more of the above methods. In a preferred embodiment of the present invention, the Dkk-1 mediated activity of the Wnt pathway may be specifically modulated by one or more of the above methods. In another preferred embodiment of the present invention, the Wnt signal transduction pathway may be stimulated by down-regulating Dkk-1 interacting protein activity; such down-regulation could, for example, yield greater LRP5 activity. In a more preferred embodiment, by stimulating LRP5 activity, bone mass regulation may be stimulated to restore or maintain a more optimal level. In another preferred embodiment, by stimulating LRP5 activity, lipid metabolism may be stimulated to restore or maintain a more optimal level. Alternative embodiments provide methods for screening candidate drugs and therapies directed to correction of bone mass disorders or lipid metabolism disorders. And, preferred embodiments of the present invention provide drugs and therapies developed by the use of the reagents and/or methods of the present invention. One skilled in the art will understand that the present invention provides important research tools to develop an effective model of

osteoporosis, to increase understanding of bone mass and lipid modulation, and to modulate bone mass and lipid metabolism.

Previous investigation of a large family in which high bone mass is inherited as a single gene (autosomal dominant) trait (HBM-1) has provided important insight into the mechanism by which bone density might be modulated. Members of this family have significantly increased spinal and hip BMD (>3 standard deviations above the norm) which affects young adults as well as elderly family members into the ninth decade. The bones of affected members, while appearing very dense radiographically, have normal external shape and outer dimensions. Cortical bone is thickened on endosteal surfaces and "affected" individuals are asymptomatic without any other phenotypic abnormalities. Assays of biochemical markers that reflect skeletal turnover suggest that the disorder is associated with a normal rate of bone remodeling. Affected individuals have achieved a balance in bone turnover at a density that is significantly greater than necessary for normal skeletal stresses. Importantly, the bones most affected are load-bearing bones which are subjected to the greatest mechanical and gravitational stresses (spine and hip). These are the most important bones to target fir therapeutic interventions in osteoporosis. The gene identified as being responsible for this phenotype. Zmax or LRP5, was not previously associated with bone physiology. The fact that modification of this gene, such as that produced by the polymorphism leading to the autosomal dominant inheritance of the HBM family phenotype, identifies Zmax/LRP5 and the pathway by which it is regulated, including Dkk/Wnt pathways discussed above, as an important target for developing modulators of bone density. Modulation of Zmax/LRP5 to mimic the gain in function provided by the HBM polymorphism would be expected to provide an important therapy for bone wasting conditions. Additionally, such modulation in young adults could enhance peak bone mass and prevent or delay fracture risk later in life. Alternatively, modulation to reduce function could be employed to treat conditions where bone is being inappropriately produced.

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3. Polypeptides

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Polypeptides contemplated for use in this invention include those which modulate Dkk and Dkk interacting protein activities. Preferred polypeptides and peptides include those which modulate the Wnt pathway. Examples of preferred sequences include the Y2H baits exemplified in Figure 2, peptide aptamers of Figure 3 (SEQ ID NOs:171-188) and Figure 4 (SEQ ID NOs:189-192), the polypeptides of the Dkk-1 interacting proteins identified in Figure 5, those polypeptides shown in Figure 6, the LRP binding domain of Dkk (amino acids 138-266 of hDkk1), the cysteine-rich domain 2 (a.a. 183-245 of hDkk-1), the cysteine-rich domain 1 (a.a. 97-138 of hDkk), and LRP5 binding aptamers of Figure 13 (including SEQ ID NOs:204-213). Although Dkk-1 is exemplified, the other Dkk proteins contain substantially similar regions and may also be used according to the present invention.

For example, the baits depicted in Figure 2 were used in a yeast two hybrid (Y2H) screen. The Y2H screen was performed as described in Example 2 to determine the minimum required binding domain for Dkk-1 to bind LRP5. The minimum binding domain constructs (i.e., residues 139-266 in bold below and residues 97-245 which are underlined, of Dkk-1) include the second cysteine rich domain which has sequence homology to a colipase fold.

20 mmalgaagat rvfvamvaaa lgghpllgvs atlnsvlnsn aiknlppplg gaaghpgsav 60 saapgilypg gnkyqtidny qpypcaedee cgtdey<u>casp trqqdagvqi clacrkrrkr</u> 120 <u>cmrhamccpg nyckngicvs sdqnhfrgei eetitesfqn dhstldgysr rttlsskmyh</u> 180 <u>tkgqegsvcl rssdcasqlc carhfwskic kpvlkeqqvc tkhrrkgshq leifqrcycg</u> 240 <u>eglscriqkd hhqasnssrl htcqrh</u> (GenBank Accession No. XP_005730) (SEQ ID NO:128).

This homology suggests a lipid-binding function and may facilitate Dkk-1 interactions at the plasma membrane (van Tilbeurgh, H., *Biochim. Biophys. Acta.* 1441:173-84 (1999)). An interaction domain of Dkk-1 that is able to interact with the ligand binding domain (LBD) of LRP5 is a useful reagent in the modulation of LRP5 activity

and modulation of Dkk-1/LRP5 complex formation. Similar screens can be prepared for Dkk-1 and Dkk-1 interacting proteins or polypeptides.

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A set of peptide aptamers was identified from a library of random peptides constrained and presented in a thioredoxin A (trxA) scaffold as described in Example 3. Peptide aptamers are powerful new tools for molecular medicine as reviewed by Hoppe-Seyler & Butz, J. Mol. Med., 78:426-430 (2000); Brody and Gold, Rev. Mol. Biotech., 74:5-13 (2000); and Colas, Curr. Opin. in Chem. Biol. 4:54-9 (2000) and the references cited therein. Briefly, peptide aptamers have been shown to be highly specific reagents capable of binding in vivo. As such, peptide aptamers provide a method of modulating the function of a protein and may serve as a substitute for conventional knock-out methods, knock-down or complete loss of function. Peptide aptamers are also useful reagents for the validation of targets for drug development and may be used as therapeutic compounds directly or provide the necessary foundation for drug design. Once identified, the peptide insert may be synthesized and used directly or incorporated into another carrier molecule. References reviewed and cited by Brody and Gold (2000, supra) describe demonstrated therapeutic and diagnostic applications of peptide aptamers and would be known to the skilled artisan.

The peptide aptamers of the present invention are useful reagents in the binding of Dkk-1 to its ligands and thereby modulation of the Wnt pathway and may be used to prevent Dkk-1 from inhibiting LRP5 modulation or Dkk-1 interacting protein modulation of the Wnt pathway. The sequence of these peptide aptamers is shown in Figure 3 (SEQ ID NOs:171-188). The peptide aptamers refers to the peptide constrained by the thioredoxin scaffold. The aptamers are also contemplated as therapeutic agents to treat Dkk-1 mediated diseases and conditions. Such aptamers are useful structural guides to chemists, for the design of mimetic compounds of the aptamers.

Peptide aptamers were likewise developed to the LRP5 ligand binding domain (LBD) bait sequences. The sequences of these peptide aptamers is shown in Figure 4 (SEQ ID NOs:189-192). These are useful reagents which may be used to disrupt

the Dkk-1/LRP5 binding interface while leaving Dkk-1 undisturbed. These can be used as comparative controls for Wnt signaling, thus, a control is provided for the specificity of any drug or therapy screened. The aptamers are also useful therapeutic agents to treat LRP mediated diseases and conditions. Such aptamers may also be used as structural guides to chemists, for the design of mimetic compounds of the aptamers.

Thirty proteins were identified which interact with Dkk-1, Dkk-1 interacting proteins, were identified in a yeast-two-hybrid screen using the Dkk-1 bait and are shown in Figure 5. It was noted that these results suggest an interaction of Dkk-1 with Notch-2. It has been suggested that cross-talk exists between the Wnt and Notch signaling pathways. For instance, Presenilin1 (Ps1) is required for Notch processing and inhibits the downstream Wnt pathway. The extracellular domain of Notch is thought to interact with Wnt. Furthermore, the Notch intracellular domain is thought to interact with disheveled and in signal induced processing, the intracellular domain is thought to interact with presenilin. (Soriano et al., J. Cell Biol. 152:785-94 (2001)). For additional information regarding the relationships between Notch and Wnt signaling, see Wesley, Mol. Cell. Biol. 19:5743-58 (1999) and Axelrod et al., Science 271:1826-32 (1996).

An interaction between Dkk-1 and chordin has also been noted; suggesting that cross-talk exists between the Wnt and TGF-beta/BMP signaling pathways (Letamendia *et al., J. Bone Joint Surg. Am.* 83A:S31 (2001); Labbe *et al., Proc. Natl. Acad. Sci. USA* 97:8358-63 (2000); Nishita *et al., Nature* 403:781-5 (2000); DeRobertis *et al., Int. J. Dev. Biol.* 45:1389-97 (2001); and Saint-Jeannet *et al., Proc. Natl. Acad. Sci. USA* 94:13713-8 (1997)). The BMP signaling pathway has an established role in bone and connective tissue development, repair and homeostasis (review in Rosen and Wozney "Bone Morphogenetic Proteins" In: Principles of Bone Biology, 2nd Edition, Eds. J. Bilezikian, L. Raisz and G. Rodan, Academic Press, pp. 919-28 (2002)). Chordin is an important molecule during development which also modulates BMP signaling in adults by sequestering BMPs in latent complexes (Piccolo *et al., Cell* 86:589-98 (1996) reviewed in Reddi, *Arthritis Res.* 3:1-5 (2001);

DeRobertis *et al.*, *Int. J. Dev. Biol.* 45:189-97 (2001)). It may be that Dkk effects bone mass modulation through both the Wnt signaling pathway via LRP and the BMP pathway via chordin.

Moreover, a number of putative growth factors, growth factor related proteins, and extracellular matrix proteins have been identified as Dkk-1 interacting proteins. Additional information regarding Dkk-1 interacting proteins identified in the Y2H assay may be obtained from publicly available databases such as PubMed via the use of the accession numbers provided in the present application. In a preferred embodiment of the invention, the amino acid sequences of these Dkk-1 interacting proteins or biologically active fragments thereof be used to modulate Dkk, Dkk-1, LRP5, LRP6, HBM, or Wnt activity. Although these proteins were identified as interacting with Dkk-1, due to the substantial homology between the various Dkk proteins, such interacting proteins are contemplated to interact with the other Dkk family members.

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4. Aptamer Mimetics

The present invention further provides for mimetics of Dkk, particularly Dkk-1, and LRP5 peptide aptamers. Such aptamers may serve as structural guides to chemists for the design of mimetic compounds of the aptamers. The aptamers and their mimetics are useful as therapeutic agents to treat LRP- or Dkk-mediated diseases and conditions.

5. Nucleic Acid Molecules

The present invention further provides nucleic acid molecules that encode polypeptides and proteins which interact with Dkk and Dkk interacting proteins, and/or LRP5 (also LRP6 and HBM) to modulate biological activities of these proteins. Preferred embodiments provide nucleic acids encoding for fragments of Dkk-1 protein, including the nucleic acids of Figure 7, the Dkk-1 interacting proteins listed in Figure 5, polypeptide aptamers of Dkk-1 (Figure 3 - SEQ ID NOs:171-188), LRP5 (Figure 4 - SEQ ID NOs:189-192), Figure 13 peptide aptamers (including SEQ

ID NO:204-214) encoded by Figure 12 polynucleotides (including SEQ ID NO:193-203), LRP6 and HBM and the related fusion proteins herein described, preferably in isolated or purified form. As used herein, "nucleic acid" is defined as RNA, DNA, or cDNA that encodes a peptide as defined above, or is complementary to a nucleic acid sequence encoding such peptides, or hybridizes to either the sense or antisense strands of the nucleic acid and remains stably bound to it under appropriate stringency conditions. The nucleic acid may encode a polypeptide sharing at least about 75% sequence identity, preferably at least about 80%, and more preferably at least about 85%, with the peptide sequences; at least about 90%, 95%, 96%, 97%, 98%, and 99% or greater are also contemplated. Specifically contemplated are genomic DNA, cDNA, mRNA, antisense molecules, enzymatically active nucleic acids (e.g., ribozymes), as well as nucleic acids based on an alternative backbone or including alternative bases, whether derived from natural sources or synthesized. Such hybridizing or complementary nucleic acids, however, are defined further as being novel and nonobvious over any prior art nucleic acid including that which encodes, hybridizes under appropriate stringency conditions, or is complementary to a nucleic acid encoding a protein according to the present invention.

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As used herein, the terms "hybridization" (hybridizing) and "specificity" (specific for) in the context of nucleotide sequences are used interchangeably. The ability of two nucleotide sequences to hybridize to each other is based upon the degree of complementarity of the two nucleotide sequences, which in turn is based on the fraction of matched complementary nucleotide pairs. The more nucleotides in a given sequence that are complementary to another sequence, the greater the degree of hybridization of one to the other. The degree of hybridization also depends on the conditions of stringency which include temperature, solvent ratios, salt concentrations, and the like. In particular, "selective hybridization" pertains to conditions in which the degree of hybridization of a polynucleotide of the invention to its target would require complete or nearly complete complementarity. The complementarity must be sufficiently high so as to assure that the polynucleotide of

the invention will bind specifically to the target nucleotide sequence relative to the binding of other nucleic acids present in the hybridization medium. With selective hybridization, complementarity will be about 90-100%, preferably about 95-100%, more preferably about 100%.

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"Stringent conditions" are those that (1) employ low ionic strength and high temperature for washing, for example: 0.015 M NaCl, 0.0015 M sodium titrate, 0.1% SDS at 50°C; or (2) employ during hybridization a denaturing agent such as formamide, for example, 50% (vol/vol) formamide with 0.1% bovine serum albumin, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 50 mM sodium phosphate buffer at pH 6.5 with 750 mM NaCl, 75 mM sodium citrate at 42°C. Another example is use of 50% formamide, 5X SSC (0.75 M NaCl, 0.075 M sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5X Denhardt's solution, sonicated salmon sperm DNA (50 μg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2X SSC and 0.1% SDS. A skilled artisan can readily determine and vary the stringency conditions appropriately to obtain a clear and detectable hybridization signal.

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As used herein, a nucleic acid molecule is said to be "isolated" or "purified" when the nucleic acid molecule is substantially separated from contaminant nucleic acid encoding other polypeptides from the source of nucleic acid. Isolated or purified is also meant to include nucleic acids which encode Dkk or fragments thereof which lack surrounding genomic sequences that flank the *Dkk* gene. Isolated or purified is further intended to include nucleic acids which encode Dkk interacting proteins or biologically active fragments thereof which lack surrounding genomic sequences that flank the Dkk interacting protein genes.

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The present invention further provides fragments of the encoding nucleic acid molecule. As used herein, a fragment of an encoding nucleic acid molecule refers to a small portion of the entire protein encoding sequence. The size of the fragment will be determined by the intended use. For example, if the fragment is chosen so as to encode an active portion of the protein, the fragment will need to be large enough to encode the functional region(s) of the protein. If the fragment is to be used as a

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nucleic acid probe or PCR primer, then the fragment length is chosen so as to obtain a relatively small number of false positives during probing/priming.

Fragments of the encoding nucleic acid molecules of the present invention (i.e., synthetic oligonucleotides) that are used as probes or specific primers for the polymerase chain reaction (PCR), or to synthesize gene sequences encoding proteins of the invention can easily be synthesized by chemical techniques, for example, the phosphotriester method of Matteucci et al. (J. Am. Chem. Soc. 103:3185-3191 (1981)) or using automated synthesis methods. In addition, larger DNA segments can readily be prepared by well known methods, such as synthesis of a group of oligonucleotides that define various modular segments of the gene, followed by ligation of oligonucleotides to build the complete modified gene.

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The polypeptide encoding nucleic acid molecules of the present invention may further be modified to contain a detectable label for diagnostic and probe purposes. A variety of such labels are known in the art and can readily be employed with the encoding molecules herein described. Suitable labels include, but are not limited to, biotin, radiolabeled nucleotides and the like. A skilled artisan can employ any of the art known labels to obtain a labeled encoding nucleic acid molecule.

Modifications to the primary structure itself by deletion, addition, or alteration of the amino acids incorporated into the protein sequence during translation can be made without destroying the activity of the protein. Such substitutions or other alterations result in proteins having an amino acid sequence encoded by a nucleic acid falling within the contemplated scope of the present invention.

Antisense molecules corresponding to the polypeptide coding or complementary sequence may be prepared. Methods of making antisense molecules which bind to mRNA, form triple helices or are enzymatically active and cleave TSG RNA and single stranded DNA (ssDNA) are known in the art. See, e.g., Antisense and Ribozyme Methodology:Laboratory Companion (Ian Gibson, ed., Chapman & Hall, 1997) and Ribozyme Protocols: Methods in Molecular Biology (Phillip C. Turner, ed., Humana Press, Clifton, NJ, 1997).

Also contemplated is the use of compounds which mediate postranscriptional gene silencing (PTGS), quelling and RNA interference (RNAi). These compounds typically are about 21 to about 25 nucleotides and are also known as short interfering RNAs or short inhibitory RNAs (siRNAs). The siRNAs are produced from an initiating double stranded RNA (dsRNA). Although the full mechanism by which the siRNAs function is not fully elucidated, it is known that these siRNAs transform the target mRNA into dsRNA, which is then degraded. Preferred forms are 5' phosphorylated siRNAs, however, hydroxylated forms may also be utilized. For additional background regarding the preparation and mechanism of siRNAs generally, see, e.g., Lipardi et al., Cell 107(3): 297-307 (2001); Boutla et al., Curr. Biol. 11(22): 1776-80 (2001); Djikeng et al., RNA 7(11): 1522-30 (2001); Elbashir et al., EMBO J. 20(23): 6877-88 (2001); Harborth et al., J. Cell. Sci. 114(Pt. 24): 4557-65 (2001); Hutvagner et al., Science 293(5531): 811-3 (2001); and Elbashir et al., Nature 411:494-98 (2001).

Also contemplated are short hairpin RNAs (shRNAs). shRNAs are a modification of the siRNA method described above. Instead of transfecting exogenously synthesized dsRNA into a cell, sequence-specific silencing can be achieved by stabling expressing siRNA from a DNA template as a fold-back stemloop, or hairpin. This approach is known as shRNA. This method permits the analysis of loss of function phenotypes due to sequence-specific gene silencing in mammalian cells by avoiding many of the problems associated with siRNAs, such as RNase degradation of the reagents, expensive chemical synthesis, etc. For additional background regarding the preparation and mechanism of shRNAs generally, see, e.g., Yu et al., PNAS 99:6047-6052 (2002); Paddison et al., Genes and Devel. 16:948-58 (2002); and Brummelkamp et al., Science 296:550-553 (2002). For additional background on the use of this method in mammalian gene knockdown methodologies, see Tuschl, Nature Biotech. 20:446-448 (2002) (and references therein).

In one preferred embodiment, the siRNA or shRNA is directed to a Dkk encoding mRNA, wherein a preferred Dkk is Dkk-1. In another embodiment, the

siRNA or shRNA is directed towards a protein which binds to and modulates the activity of or is modulated by a Dkk; these proteins include LRP5, LRP6 and HBM as well as other members of the Wnt pathway.

6. <u>Isolation of Other Related Nucleic Acid Molecules</u>

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The identification of the nucleic acid molecule of Dkk allows a skilled artisan to isolate nucleic acid molecules that encode other members of the Dkk family (see, Krupnik et al., 1999). Further, the presently disclosed nucleic acid molecules allow a skilled artisan to isolate nucleic acid molecules that encode Dkk-1-like proteins, in addition to Dkk-1. The presently disclosed Dkk-1 interacting proteins and their corresponding nucleic acid molecules allows a skilled artisan to further isolate other related protein family members which interact with Dkk-1.

A skilled artisan can readily use the amino acid sequence of Dkk and Dkk interacting proteins to generate antibody probes to screen expression libraries prepared from appropriate cells. Typically, polyclonal antiserum from mammals such as rabbits immunized with the purified protein (as described below) or monoclonal antibodies can be used to probe a mammalian cDNA or genomic expression library, such as a human macrophage library, to obtain the appropriate coding sequence for other members of the protein family. The cloned cDNA sequence can be expressed as a fusion protein, expressed directly using its own control sequences, or expressed by constructions using control sequences appropriate to the particular host used for expression of the desired protein.

Alternatively, a portion of the coding sequence herein described can be synthesized and used as a probe to retrieve DNA encoding a member of the protein family from any mammalian organism. Oligomers containing approximately 18-20 nucleotides (encoding about a 6-7 amino acid stretch) are prepared and used to screen genomic DNA or cDNA libraries to obtain hybridization under stringent conditions or conditions of sufficient stringency to eliminate an undue level of false positives.

Additionally, pairs of oligonucleotide primers can be prepared for use in a polymerase chain reaction (PCR) to selectively clone an encoding nucleic acid

molecule. A PCR denature/anneal/extend cycle for using such PCR primers is well known in the art and can readily be adapted for use in isolating other encoding nucleic acid molecules. For example, degenerate primers can be utilized to obtain sequences related to Dkk-1 or Dkk-1 interacting proteins. Primers can be designed that are not perfectly complementary and can still hybridize to a portion of a target sequence or flanking sequence and thereby provide for amplification of all or a portion of a target sequence. Primers of about 20 nucleotides or less, preferably have about one to three mismatches located at the 5' and/or 3' ends. Primers of about 20 to 30 nucleotides have up to about 30% mismatches and can still hybridize to a target sequence. Hybridization conditions for primers with mismatch can be determined by the method described in Maniatis et al., Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1982) or by reference to known methods. The ability of the primer to hybridize to a sequence of either Dkk-1, a Dkk-1 interacting protein, or a related sequence under varying conditions can be determined using this method. Because a target sequence is known, the effect of mismatches can be determined by methods known to those of skill in the art. Degenerate primers would be based on putative conserved amino acid sequences of the Dkk-1 and Dkk-1 interacting protein genes.

7. rDNA Molecules for Polypeptide Expression

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The present invention further provides recombinant DNA molecules (rDNAs) that contain a polypeptide coding sequence. As used herein, a rDNA molecule is a DNA molecule that has been subjected to molecular manipulation *in situ*. Methods for generating rDNA molecules are well known in the art, for example, see Sambrook *et al.*, Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989). In the preferred rDNA molecules, a coding DNA sequence is operably linked to expression control sequences and/or vector sequences.

The choice of vector and/or expression control sequences to which one of the protein family encoding sequences of the present invention is operably linked depends directly, as is well known in the art, on the functional properties desired, *e.g.*, protein

expression, and the host cell to be transformed. A vector contemplated by the present invention is at least capable of directing the replication and/or insertion into the host chromosome, and preferably also expression, of the structural gene included in the rDNA molecule.

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Expression control elements that are used for regulating the expression of an operably linked protein encoding sequence are known in the art and include, but are not limited to, inducible promoters, constitutive promoters, secretion signals, and other regulatory elements. Preferably, the inducible promoter is readily controlled, such as being responsive to a nutrient in the host cell's medium. Preferred promoters include yeast promoters, which include promoter regions for metallothionein, 3phosphoglycerate kinase or other glycolytic enzymes such as enolase or glyceraldehyde-3-phosphate dehydrogenase, enzymes responsible for maltose and galactose utilization, and others. Vectors and promoters suitable for use in yeast expression are further described in EP 73,675A. Appropriate non-native mammalian promoters might include the early and late promoters from SV40 (Fiers et al, Nature, 273:113 (1978)) or promoters derived from Moloney murine leukemia virus, mouse tumor virus, avian sarcoma viruses, adenovirus II, bovine papilloma virus or polyoma. In addition, the construct may be joined to an amplifiable gene (e.g., DHFR) so that multiple copies of the gene may be made. For appropriate enhancer and other expression control sequences, see also Enhancers and Eukaryotic Gene Expression (Cold Spring Harbor Press, Cold Spring Harbor, NY, 1983). Preferred bone related promoters include CMVbActin or type I collagen promoters to drive expression of the human HBM, Zmax1/LRP5 or LRP6 cDNA. Other preferred promoters for mammalian expression are from cytomegalovirus (CMV), Rous sarcoma virus (RSV), Simian virus 40 (SV40), and EF-1a (human elongation factor 1a-subunit).

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In one embodiment, the vector containing a coding nucleic acid molecule will include a prokaryotic replicon, *i.e.*, a DNA sequence having the ability to direct autonomous replication and maintenance of the recombinant DNA molecule extrachromosomally in a prokaryotic host cell, such as a bacterial host cell, transformed therewith. Such replicons are well known in the art. In addition, vectors with a

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prokaryotic replicon may also include a gene whose expression confers a detectable marker such as a drug resistance. Typical bacterial drug resistance genes are those that confer resistance to ampicillin or tetracycline.

Vectors that include a prokaryotic replicon can further include a prokaryotic or bacteriophage promoter capable of directing the expression (transcription and translation) of the coding gene sequences in a bacterial host cell, such as *E. coli*. A promoter is an expression control element formed by a DNA sequence that permits binding of RNA polymerase and transcription to occur. Promoter sequences compatible with bacterial hosts are typically provided in plasmid vectors containing convenient restriction sites for insertion of a DNA segment of the present invention. Typical of such vector plasmids are pUC8, pUC9, pBR322 and pBR329 available from Biorad Laboratories, (Richmond, CA), and pPL and pKK223 available from Pharmacia (Piscataway, NJ).

Expression vectors compatible with eukaryotic cells, preferably those compatible with vertebrate cells, can also be used to form a rDNA molecule that contains a coding sequence. Eukaryotic cell expression vectors are well known in the art and are available from several commercial sources. Typically, such vectors are provided containing convenient restriction sites for insertion of a desired DNA segment. Typical of such vectors are pSVL and pKSV-10 (Pharmacia), pBPV-1/pML2d (International Biotechnologies, Inc.), vector systems that include Histidine Tags and periplasmic secretion, or other vectors described in the art.

Eukaryotic cell expression vectors used to construct the rDNA molecules of the present invention may further include a selectable marker that is effective in an eukaryotic cell, preferably a drug resistance selection marker. A preferred drug resistance marker is the gene whose expression results in neomycin resistance, *i.e.*, the neomycin phosphotransferase (*neo*) gene (Southern *et al.*, *J. Mol. Anal. Genet.* 1:327-341 (1982)). Alternatively, the selectable marker can be present on a separate plasmid, and the two vectors introduced by co-transfection of the host cell, and selected by culturing in the appropriate drug for the selectable marker.

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8. <u>Host Cells Containing an Exogenously Supplied rDNA Nucleic Acid</u> Molecule

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The present invention further provides host cells transformed with a nucleic acid molecule that encodes a polypeptide or protein of the present invention. The host cell can be either prokaryotic or eukaryotic. Eukaryotic cells useful for expression of a protein of the invention are not limited, so long as the cell line is compatible with cell culture methods and compatible with the propagation of the expression vector and expression of the gene product. Preferred eukaryotic host cells include, but are not limited to, yeast, insect and mammalian cells, preferably vertebrate cells such as those from a mouse, rat, monkey or human cell line but also can include invertebrates with, for example, cartilage. Preferred eukaryotic host cells include but are not limited to Chinese hamster ovary (CHO) cells (ATCC No. CCL61), NIH Swiss mouse embryo cells NIH/3T3 (ATCC No. CRL 1658), baby hamster kidney cells (BHK), HOB-03-CE6 osteoblast cells, and other like eukaryotic tissue culture cell lines.

Any prokaryotic host can be used to express a rDNA molecule encoding a protein of the invention. A preferred prokaryotic host is *E. coli*.

Transformation of appropriate cell hosts with a recombinant DNA (rDNA) molecule of the present invention is accomplished by well known methods that typically depend on the type of vector used and host system employed. With regard to transformation of prokaryotic host cells, electroporation and salt treatment methods are typically employed; see, for example, Cohen *et al.*, *Proc. Natl. Acad. Sci. USA* 69: 2110 (1972); Maniatis *et al.* (1982); and Sambrook *et al.* (1989). With regard to transformation of vertebrate cells with vectors containing rDNAs, electroporation, cationic lipid or salt treatment methods are typically employed; see, for example, Graham *et al.*, *Virol.* 52: 456 (1973); Wigler *et al.*, *Proc. Natl. Acad. Sci. USA* 76: 1373-76 (1979).

Successfully transformed cells, *i.e.*, cells that contain a rDNA molecule of the present invention, can be identified by well known techniques including the selection for a selectable marker. For example, cells resulting from the introduction of an rDNA of the present invention can be cloned to produce single colonies. Cells from those

colonies can be harvested, lysed and their DNA content examined for the presence of the rDNA using a method such as that described by Southern, *J. Mol. Biol.* 98: 503 (1975), or Berent *et al.*, *Biotech.* 3: 208 (1985). Alternatively, the cells can be cultured to produce the proteins encoded by the rDNA and the proteins harvested and assayed, using for example, any suitable immunological method. See, *e.g.*, Harlow *et al.*, (1988).

Recombinant DNA can also be utilized to analyze the function of coding and non-coding sequences. Sequences that modulate the translation of the mRNA can be utilized in an affinity matrix system to purify proteins obtained from cell lysates that associate with the Dkk-1 or Dkk-1 interacting protein or expression control sequence. Synthetic oligonucleotides would be coupled to the beads and probed with the lysates, as is commonly known in the art. Associated proteins could then be separated using, for example, a two dimensional SDS-PAGE system. Proteins thus isolated could be further identified using mass spectroscopy or protein sequencing. Additional methods would be apparent to the skilled artisan.

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9. <u>Production of Recombinant Peptides and Proteins using a cDNA or Other Recombinant Nucleic Acids</u>

The invention also relates to nucleic acid molecules which encode a Dkk protein and polypeptide fragments thereof, and proteins and polypeptides which bind to Dkk (e.g., LRP5, LRP6 and HBM, Dkk interacting proteins such as the proteins of Figure 5) and molecular analogues. The polypeptides of the present invention include the full length Dkk and polypeptide fragments thereof, Dkk binding proteins and polypeptides thereof. Preferably these proteins are mammalian proteins, and most preferably human proteins and biologically active fragments thereof. Alternative embodiments include nucleic acid molecules encoding polypeptide fragments having a consecutive amino acid sequence of at least about 3, 5, 7, 8, 9, 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 125, 150, 175, or 200 amino acid residues from a common polypeptide sequence; amino acid sequence variants of a common polypeptide sequence wherein an amino acid residue has been inserted N- or C-terminal to, or within, the polypeptide sequence or its fragments; and amino acid sequence variants of the common

polypeptide sequence or its fragments, which have been substituted by another conserved residue. Recombinant nucleic acid molecules which encode polypeptides include those containing predetermined mutations by, e.g., homologous recombination. site-directed or PCR mutagenesis, and recombinant Dkk proteins or polypeptide fragments of other animal species, including but not limited to vertebrates (e.g., rabbit, rat, murine, porcine, camelid, reptilian, caprine, avian, fish, bovine, ovine, equine and non-human primate species) as well as invertebrates, and alleles or other naturally occurring variants and homologs of Dkk binding proteins of the foregoing species and of human sequences. Also contemplated herein are derivatives of the commonly known Dkk, Dkk interacting proteins, or fragments thereof, wherein Dkk, Dkk interacting proteins, or their fragments have been covalently modified by substitution. chemical, enzymatic, or other appropriate means with a moiety other than a naturally occurring amino acid (for example a detectable moiety such as an enzyme or radioisotope) and soluble forms of Dkk. It is further contemplated that the present invention also includes nucleic acids with silent mutations which will hybridize to the endogenous sequence and which will still encode the same polypeptide.

The nucleic acid molecules encoding Dkk binding proteins, the LRP5 binding domain fragment of Dkk, or other polypeptides of the present invention are preferably those which share a common biological activity (e.g., mediate Dkk activity such as its interaction with LRP5, HBM or LRP6). The polypeptides of the present invention include those encoded by a nucleic acid molecule with silent mutations, as well as those nucleic acids encoding a biologically active protein with conservative amino acid substitutions, allelic variants, and other variants of the disclosed polypeptides which maintain at least one Dkk activity.

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The amino acid compounds of the invention are polypeptides which are partially defined in terms of amino acid residues of designated classes. Polypeptide homologs would include conservative amino acid substitutions within the amino acid classes described below. Amino acid residues can be generally sub-classified into four major subclasses as follows:

Acidic: The residue has a negative charge due to loss of H⁺ ion at physiological pH, and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium, at physiological pH.

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Basic: The residue has a positive charge due to association with H⁺ ion at physiological pH, and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium at physiological pH.

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Neutral/non-polar: The residues are not charged at physiological pH, but the residue is repelled by aqueous solution so as to seek the inner positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium. These residues are also designated "hydrophobic."

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<u>Neutral/polar:</u> The residues are not charged at physiological pH, but the residue is attracted by aqueous solution so as to seek the outer positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

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It is understood, of course, that in a statistical collection of individual residue molecules some molecules will be charged, and some not, and there will be an attraction for or repulsion from an aqueous medium to a greater or lesser extent. To fit the definition of "charged", a significant percentage (at least approximately 25%) of the individual molecules are charged at physiological pH. The degree of attraction or repulsion required for classification as polar or nonpolar is arbitrary and, therefore, amino acids specifically contemplated by the invention have been classified as one or the other. Most amino acids not specifically named can be classified on the basis of known behavior.

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Amino acid residues can be further subclassified as cyclic or noncyclic, and aromatic or non-aromatic, self-explanatory classifications with respect to the side chain substituent groups of the residues, and as small or large. The residue is considered small if it contains a total of 4 carbon atoms or less, inclusive of the carboxyl carbon. Small residues are, of course, always nonaromatic.

The gene-encoded secondary amino acid proline, although technically within the group neutral/nonpolar/large/cyclic and nonaromatic, is a special case due to its known effects on the secondary conformation of peptide chains, and is not, therefore, included in this defined group.

Other amino acid substitutions of those encoded in the gene can also be included in peptide compounds within the scope of the invention and can be classified within this general scheme according to their structure.

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All of the compounds of the invention may be in the form of the pharmaceutically acceptable salts or esters. Salts may be, for example, Na⁺, K⁺, Ca⁺², Mg⁺² and the like; the esters are generally those of alcohols of 1-6 carbons.

The present invention further provides methods for producing a protein of the invention using nucleic acid molecules herein described. In general terms, the production of a recombinant form of a protein typically involves the following steps.

First, a nucleic acid molecule is obtained that encodes Dkk, such as a nucleic acid molecule encoding human Dkk or any other Dkk sequence, or that encodes a Dkk binding protein, a Dkk aptamer or a biologically active fragment thereof. Particularly for Dkk binding peptides, the nucleotides encoding the peptide are incorporated into a nucleic acid in the form of an in-frame fusion, insertion into or appended to a thioredoxin coding sequence. The coding sequence (ORF) is directly suitable for expression in any host, as it is not interrupted by introns.

These DNAs can be transfected into host cells such as eukaryotic cells or prokaryotic cells. Eukaryotic hosts include mammalian cells and vertebrate (e.g., osteoblasts, osteosarcoma cell lines, Drosophila S2 cells, hepatocytes, tumor cell lines and other bone cells of any mammal, as well as insect cells, such as Sf9 cells using recombinant baculovirus). For example, a DNA expressing an open reading frame (ORF) under control of a type I collagen promoter, or such osteoblast promoters as osteocalcin histone, type I collagen, TGFβ1, MSX2, cfos/cJun and Cbfa1, can be used to regulate the Dkk in animal cells. Alternatively, the nucleic acid can be placed downstream from an inducible promoter, which can then be placed into vertebrate or invertebrate cells or be used in creating a transgenic animal model.

Alternatively, proteins and polypeptides of the present invention can be expressed in an heterologous system. The human cell line GM637, SV-40 transformed human fibroblasts, can be transfected, with a plasmid containing a Dkk ligand binding domain coding sequence under the control of the chicken actin promoter (Reis *et al.*, EMBO J. 11: 185-193 (1992)). Such transfected cells could be used as a source of Dkk binding domain in functional assays. Alternatively, polypeptides encoding only a portion of Dkk or any of the disclosed Dkk binding peptides Dkk aptamers or a polypeptide encoding a Dkk interacting protein can be expressed alone or in the form of a fusion protein. For example, Dkk derived peptides can be expressed in bacteria (*e.g.*, *E. coli*) as GST- or His-Tag fusion proteins. These fusion proteins are then purified and can be used to generate polyclonal antibodies or can be used to identify other Dkk ligands.

The nucleic acid coding sequence is preferably placed in operable linkage with suitable control sequences, as described above, to form an expression unit containing the protein encoding open reading frame. The expression unit is used to transform a suitable host and the transformed host is cultured under conditions that allow the production of the recombinant protein. Optionally the recombinant protein is isolated from the medium or from the cells; recovery and purification of the protein may not be necessary in some instances where some impurities may be tolerated.

Each of the foregoing steps can be done in a variety of ways. For example, the desired coding sequences may be obtained from genomic fragments and used directly in appropriate hosts. The construction of expression vectors that are operable in a variety of hosts is accomplished using appropriate replicons and control sequences, as set forth above. The control sequences, expression vectors, and transformation methods are dependent on the type of host cell used to express the gene and were discussed in detail earlier. Suitable restriction sites can, if not normally available, be added to the ends of the coding sequence so as to provide an excisable gene to insert into these vectors. A skilled artisan can readily adapt any host/expression system known in the art for use with the nucleic acid molecules of the invention to produce recombinant protein.

10. <u>Methods to Identify Binding Partners</u>

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Another embodiment of the present invention provides methods for use in isolating and identifying binding partners of Dkk or Dkk interacting proteins. Dkk or a Dkk interacting protein or a polypeptide fragment thereof can be mixed with a potential binding partner or an extract or fraction of a cell under conditions that allow the association of potential binding partners with Dkk or with Dkk interacting proteins. After mixing, the peptides, polypeptides, proteins or other molecules that have become associated with Dkk or a Dkk interacting protein are separated from the mixture. The binding partner that bound to the polypeptide then can be purified and further analyzed. Determination of binding partners of Dkk and Dkk interacting proteins as well as agents which prevent the interaction of Dkk with one of its interacting proteins (e.g., LRP5, LRP6, HBM, or those proteins listed in Figure 5) can be performed using a variety of different competition assays as are known in the art. For example, the minimal sequence of Dkk, as described herein, can be used to identify antibodies which compete with LRP5 (or LRP6, HBM or other ligand binding partners) for binding to Dkk-1 and vice versa. The minimal Dkk sequence can be bound to the bottom of a 96-well plate (or other solid substrate), and antibodies or other potential binding agents (e.g., polypeptides, mimetics, homologs, antibody fragments and the like) can be screened in a competition assay to identify agents with binding affinities, for example, greater than the natural ligand binding partner of Dkk.

In the present invention, suitable cells are used for preparing assays, for the expression of a LRP and/or Dkk or proteins that interact therewith. The cells may be made or derived from mammals, yeast, fungi, or viruses. A suitable cell for the purposes of this invention is one that includes but is not limited to a cell that can exhibit a detectable Dkk-LRP (or HBM) interaction, and preferably, the differential interaction between Dkk-1-LRP5 and Dkk-1-HBM. For the desired assay, the cell type may vary. In several embodiments, bone cells are preferred, for example, a human osteoblast cell (e.g. hOB-03-CE6) or osteosarcoma cell (e.g. U2OS). Additional hOB cells are hOB-03-C5, hOB-02-02 and, an immortalized pre-osteocytic cell line referred to as hOB-01-C1-PS-09 cells (which are deposited with American Type Culture Collection in

Manassas, Va. with the designation PTA-785), Examples of osteosarcoma cells would include SaoS2, MG63 and HOS TE85. Immortalized refers to a substantially continuous and permanently established cell culture with substantially unlimited cell division potential. That is, the cells can be cultured substantially indefinitely, i.e., for at least about 6 months under rapid conditions of growth, preferably much longer under slower growth conditions, and can be propagated rapidly and continually using routine cell culture techniques. Alternatively stated, preferred cells can be cultured for at least about 100, 150 or 200 population doublings. These cells produce a complement of proteins characteristic of normal human osteoblastic cells and are capable of osteoblastic differentiation. They can be used in cell culture studies of osteoblastic cell sensitivity to various agents, such as hormones, cytokines, and growth factors, or in tissue therapy. Certain non bone cells such as HEK 293 cells that exhibit detectable Dkk-LRP (or HBM) interaction are also be useful for the assays of this invention.

To identify and isolate a binding partner, the entire Dkk protein (e.g., human Dkk-1, GenBank Accession No. BAA34651) or a Dkk interacting protein (Genbank Accession Nos. for some Dkk-1 interacting proteins are given in Figure 5) can be used. Alternatively, a polypeptide fragment of the protein can be used. Suitable fragments of the protein include at least about 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150 or more contiguous amino acid residues of any Dkk or Dkk interactor sequence. Preferable sequences of Dkk include portions or all of one or both of the cysteine rich domains (e.g., Cys-1 and Cys-2 of Dkk-1) or the conserved sequences at the amino terminus of Dkk-1 (See Krupnik et al., Gene 238: 301-313 (1999)). Alternatively, portions of LRP5, LRP6, HBM and other Dkk interacting proteins such as those in Figure 5 that interact with Dkk-1 can be used to identify and isolate agents which modulate Dkk activity. Alternatively, peptide aptamers of LRP5, LRP6, HBM, Dkk and other Dkk interacting proteins such as those in Figure 5 that interact with Dkk-1 can be used to identify and isolate agents which modulate Dkk activity.

As used herein, a cellular extract refers to a preparation or fraction which is made from a lysed or disrupted cell. A variety of methods can be used to obtain cell

extracts. Cells can be disrupted using either physical or chemical disruption methods. Examples of physical disruption methods include, but are not limited to, sonication and mechanical shearing. Examples of chemical lysis methods include, but are not limited to, detergent lysis and enzyme lysis. A skilled artisan can readily adapt methods for preparing cellular extracts in order to obtain extracts for use in the present methods.

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Once an extract of a cell is prepared, the extract is mixed with the protein of the invention under conditions in which association of the protein with the binding partner can occur. A variety of conditions can be used, the most preferred being conditions that closely resemble conditions found in the cytoplasm of a human cell. Features such as osmolarity, pH, temperature, and the concentration of cellular extract used, can be varied to optimize the association of the protein with the binding partner.

After mixing under appropriate conditions, the bound complex is separated from the mixture. A variety of techniques can be utilized to separate the mixture. For example, antibodies specific to a protein of the invention can be used to immunoprecipitate the binding partner complex. Alternatively, standard chemical separation techniques such as chromatography and density/sediment centrifugation can be used. For example, a protein of the invention is expressed with an affinity tag such as a His tag. The His labeled protein and any bound molecule may be retained and selectively eluted from a Ni-NTA column.

After removal of non-associated cellular constituents found in the extract, the binding partner can be dissociated from the complex using conventional methods. For example, dissociation can be accomplished by altering the salt concentration or pH of the mixture.

To aid in separating associated binding partner pairs from the mixed extract, the protein of the invention can be immobilized on a solid support. For example, the protein can be attached to a nitrocellulose matrix or acrylic beads. Attachment of the protein to a solid support aids in separating peptide/binding partner pairs from other constituents found in the extract. The identified binding partners can be either a single protein or a complex made up of two or more proteins.

Alternatively, the nucleic acid molecules of the invention can be used in a Y2H system. The Y2H system has been used to identify other protein partner pairs and can readily be adapted to employ the nucleic acid molecules herein described. Methods of performing and using Y2H systems are known. See, e.g., Finley et al., "Two-Hybrid Analysis of Genetic Regulatory Networks," in The Yeast Two-Hybrid System (Paul L. Bartel et al., eds., Oxford, 1997); Meijia Yang, "Use of a Combinatorial Peptide Library in the Two-Hybrid Assay," in The Yeast Two-Hybrid System (Paul L. Bartel et al., eds., Oxford, 1997); Gietz et al., "Identification of proteins that interact with a protein of interest: Applications of the yeast two-hybrid system," Mol. & Cell. Biochem. 172: 67-9 (1997); K. H. Young, "Yeast Two-Hybrid: So Many Interactions,(in) so Little Time," Biol. Reprod. 58: 302-311 (1998); R. Brent et al., "Understanding Gene and Allele Function with Two-Hybrid Methods," Annu. Rev. Genet. 31:663-704 (1997) and U.S. Patent No. 5,989,808. The Dkk-1 interacting proteins identified in Figure 5 were identified using the Y2H interacting system using Dkk-1 as bait.

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One preferred in vitro binding assay for Dkk modulators would comprise a mixture of a LRP binding domain of Dkk and one or more candidate binding targets or substrates. After incubating the mixture under appropriate conditions, one would determine whether Dkk or a fragment thereof bound with the candidate modulator present. For cell-free binding assays, one or more of the components usually comprises or is coupled to a label. The label may provide for direct detection, such as radioactivity, luminescence, optical or electron density, etc., or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods may be employed to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and the label thereafter detected. Fluorescence resonance energy transfer may be utilized to monitor the interaction of two labeled molecules. For example, a fluorescence label on Dkk and another label on LRP5 or a soluble fragment thereof such as the extracellular domain will exchange fluorescence resonance energy when in close proximity indicating that the two molecules are bound. A preferred binding partner for Dkk will

increase or decrease the affinity between Dkk and LRP5 which will be readily observable in a fluorescence spectrometer. Alternatively, an instrument, such as a surface plasmon resonance detector manufactured by BIAcore (Uppsala, Sweden), may be used to observe interactions with a fixed target. One skilled in the art knows of many other methods which may be employed for this purpose.

Thereby, the present invention provides methods for screening candidates including polypeptides of the present invention for activity which identifies these candidates as valuable drug leads. Other suitable methods are also known in the art and are suitable for use herein, including *Xenopus* oocyte injection studies and TCF luciferase assays.

Additional assays can be used to identify the activity of Dkk and Dkk interacting proteins in the Wnt pathway, as well as the impact of modulators of Dkk and Dkk interacting proteins on the Wnt pathway. These include, for example, a *Xenopus* embryo assay and a TCF-luciferase reporter gene assay to monitor Wnt signaling modulation.

Xenopus embryos are an informative *in vivo* assay system to evaluate the modulation of Wnt signaling. Ectopic expression of certain Wnts or other activators of the Wnt signaling pathway results in a bifurcation of the anterior neural plate. This bifurcation results in a duplicated body axis, which suggests a role for Wnt signaling during embryonic development (McMahon *et al.*, *Cell* 58: 1075-84 (1989); Sokol *et al.*, *Cell* 67: 741-52 (1991)). Since these original observations, the *Xenopus* embryo assay has been extensively used as an assay system for evaluating modulation of the Wnt signaling pathway. One preferred embodiment of the present invention is demonstrated in Example 6.

Constructs for *Xenopus* expression can be prepared as would be known in the art. For example, a variety of cDNAs have been engineered into the vector pCS2+ (Turner *et al.*, *Genes Devel.* 8: 1434-1447 (1994)) to facilitate the *in vitro* generation of mRNA for use in *Xenopus* embryo injection experiments. DNA inserts are subcloned in the sense orientation with respect to the vector SP6 promoter. Downstream of the insert, the vector provides an SV40 virus polyadenlylation signal and a T3 promoter

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sequence (i.e., for the generation of antisense mRNA). Constructs can be generated for various Dkk family members, LRP5, LRP6, HBM, Dkk-1 interactors, etc. Constructs could be generated in pCS2⁺ that contain the nucleic acid sequence encoding for the peptide aptamers that were identified in yeast screens. These sequences would be fused to a 5' synthetic translation initiation sequence followed by a canonical signal sequence to ensure that the peptide aptamer would be translated and secreted from the cell.

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Once these constructs are made then mRNA can be synthesized and injected into *Xenopus* oocytes. mRNA for microinjection into *Xenopus* embryos is generated by *in vitro* transcription using the cDNA constructs in the pCS2⁺ vector described above as template. Various amounts of RNA can be injected into the ventral blastomere of the 4-or 8-cell *Xenopus* embryo substantially as described in Moon *et al.*, *Technique-J. of Methods in Cell and Mol. Biol.* 1: 76-89 (1989), and Peng, *Meth. Cell. Biol.* 36: 657-62 (1991).

Previous data has shown that expression of LRP5, in the presence of Wnt5a, results in a Wnt-induced duplicated axis formation in *Xenopus* embryos (Tamai *et al.*, *Nature* 407: 530-535 (2000)). The roles of Dkk-1 and Dkk-2, and Dkk-1 interacting proteins, in modulating the LRP5-mediated Wnt response *in vivo* can be analyzed using, for example, the *Xenopus* embryo. In addition, the peptide aptamers, Dkk

interacting proteins, or combinations of the above can be evaluated in a similar manner.

Experiments can also be conducted wherein RNA is injected into the dorsal blastomere to ensure the specificity of the observed phenotypes. Lineage tracing experiments can be performed where a marker gene such as green fluorescent protein (GFP) or LacZ is co-injected with the experimental RNAs. Detecting marker gene expression would identify the targeted cells of the microinjection and aid in elucidating the mechanism of action. In addition to the Wnt signaling components listed above, the point at which HBM acts upon the Wnt pathway can also be analyzed. This can be done by co-injections of various dominant-negative constructs. For example, a dominant negative TCF-3 construct would be useful to demonstrate that the observed axis duplication (and Wnt activation) is mediated via the β-catenin-TCF response. If so,

such a construct would be expected to abolish the observed duplicated axis phenotype. Another example would include a dominant negative Dsh construct. Since Dsh is far upstream in the Wnt signaling pathway, a dominant negative construct should abolish the activation of the Wnt response and the observed axis duplication. If it does not, this would suggest that axis duplication is being induced via a different signaling pathway.

The marker genes of the injected *Xenopus* embryos can be analyzed as follows. Representative embryos are collected at stage 10.5 (11 hours post fertilization) for marker gene analysis. RNA is extracted and purified from the embryos following standard protocols (Sambrook *et al.*, 1989 at 7.16). Marker genes could include the following: Siamois (*i.e.*, Wnt responsive gene), Xnr3 (*i.e.*, Wnt responsive gene), slug (*i.e.*, neural crest marker), Xbra (*i.e.*, early mesoderm marker), HNK-1 (*i.e.*, ectodermal/neural marker), endodermin (*i.e.*, endoderm), Xlhbox8 (*i.e.*, pancreatic), BMP2 and BMP4 (*i.e.*, early mesoderm), XLRP6 (*i.e.*, maternal and zygotic expression, it is also the LRP6 homolog in the frog), EF-1 (*i.e.*, control) and ODC (*i.e.*, control). Induction of marker genes is analyzed and quantitated by RT-PCR/TagMan®.

This type of marker analysis is excellent to monitor changes in gene expression that result very early in the embryo as a direct result of signaling perturbation. Other experiments could be designed that would monitor changes in gene expression in a more tissue or spatially-restricted fashion. Examples would include the generation of a transgenic *Xenopus* model. For example, Zmax/LRP5 and HBM expression could be under the control of the brachyury or cardiac-actin promoters directing gene expression transiently in the mesoderm or in the somites, respectively. Phenotype analyses of these transgenic *Xenopus* animals would include marker gene analysis/transcriptional profiling (from a restricted tissue source) and histologic examination of the tissue.

A TCF-luciferase assay system such as that described in Example 7 can also be used to monitor Wnt signaling activity, Dkk activity and Dkk interacting protein activity. Constructs for the TCF-luciferase assays can be prepared as would be known in the art. For example, Dkk and Dkk interacting protein peptides, LRP5/LRP6, among others, can be expressed in pcDNA3.1, using Kozak and signal sequences to target peptides for secretion.

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Once constructs have been prepared, cells such as osteoblasts and HEK293 cells are seeded in well plates and transfected with construct DNA, CMV betagalactosidase plasmid DNA, and TCF-luciferase reporter DNA. The cells are then lysed and assayed for beta-galactosidase and luciferase activity to determine whether Dkk, Dkk interacting proteins, or other molecules such as antibodies affect Wnt signaling.

Additional assays for monitoring Wnt signaling activity, Dkk activity, and Dkk interacting protein activity include:

Modulation of another Wnt-responsive transcription factor, LEF, as visualized by a reporter gene activity. One example includes the activation of the LEF1 promoter region fused to the luciferase reporter gene (Hsu *et al.*, *Mol. Cell. Biol.* 18: 4807-18 (1999)).

Alterations in cell proliferation, cell cycle or apoptosis. There are numerous examples describing Wnt-mediated cellular transformations including Shimizu *et al.*, *Cell. Growth Differ*. 8: 1349-58 (1997).

Stabilization and cellular localization of de-phosphorylated β -catenin as an indicator of Wnt activation (Shimizu et al., 1997).

Additional methods of assaying Wnt signaling, through either the canonical or non-canonical pathways, would be apparent to the artisan of ordinary skill.

11. <u>Methods to Identify Agents that Modulate the Expression of a Nucleic Acid</u> <u>Encoding the Dkk and/or LRP5 Proteins and/or Dkk interacting proteins</u>

Another embodiment of the present invention provides methods for identifying agents that modulate the expression of a nucleic acid encoding Dkk. Such assays may utilize any available means of monitoring for changes in the expression level of the nucleic acids of the invention. As used herein, an agent is said to modulate the expression of Dkk, if it is capable of up- or down-regulating expression of the nucleic acid in a cell (e.g., mRNA).

In one assay format, cell lines that contain reporter gene fusions between the nucleic acid encoding Dkk (or proteins which modulate the activity of Dkk) and any

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assayable fusion partner may be prepared. Numerous assayable fusion partners are known and readily available, including but not limited to the firefly luciferase gene and the gene encoding chloramphenicol acetyltransferase (Alam *et al.*, *Anal. Biochem.* 188: 245-254 (1990)). Cell lines containing the reporter gene fusions are then exposed to the agent to be tested under appropriate conditions and time. Differential expression of the reporter gene between samples exposed to the agent and control samples identifies agents which modulate the expression of a nucleic acid encoding Dkk or other protein which modulates Dkk activity. Such assays can similarly be used to determine whether LRP5 and even LRP6 activity is modulated by regulating Dkk activity.

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Additional assay formats may be used to monitor the ability of the agent(s) to modulate the expression of a nucleic acid encoding Dkk, alone or Dkk and LRP5, and/or Dkk interacting proteins such as those identified in Figure 5. For instance, mRNA expression may be monitored directly by hybridization to the nucleic acids of the invention. Cell lines are exposed to the agent to be tested under appropriate conditions and time and total RNA or mRNA is isolated by standard procedures such those disclosed in Sambrook *et al.* (1989); Ausubel *et al.*, Current Protocols in Molecular Biology (Greene Publishing Co., NY, 1995); Maniatis *et al.*, Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1982); and Short Protocols in Molecular Biology; A Compendium of Methods from Current Protocols in Molecular Biology (Frederick M. Ausubel *et al.*, April 1999).

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Probes to detect differences in RNA expression levels between cells exposed to the agent and control cells may be prepared from the nucleic acids of the invention. It is preferable, but not necessary, to design probes which hybridize only with target nucleic acids under conditions of high stringency. Only highly complementary nucleic acid hybrids form under conditions of high stringency. Accordingly, the stringency of the assay conditions determines the amount of complementarity which should exist between two nucleic acid strands in order to form a hybrid. Stringency should be chosen to maximize the difference in stability between the probe:target hybrid and potential probe:non-target hybrids.

Probes may be designed from the nucleic acids of the invention through methods known in the art. For instance, the G+C content of the probe and the probe length can affect probe binding to its target sequence. Methods to optimize probe specificity are commonly available. See for example, Sambrook *et al.* (1989) or Ausubel *et al.* (Current Protocols in Molecular Biology, Greene Publishing Co., NY, 1995).

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Hybridization conditions are modified using known methods, such as those described by Sambrook et al. (1989) and Ausubel et al. (1995), as suitable for each probe. Hybridization of total cellular RNA or RNA enriched for polyA RNA can be accomplished in any available format. For instance, total cellular RNA or RNA enriched for polyA RNA can be affixed to a solid support and the solid support exposed to at least one probe comprising at least one, or part of one of the nucleic acid sequences of the invention under conditions in which the probe will specifically hybridize. Alternatively, nucleic acid fragments comprising at least one, or part of one of the sequences of the invention can be affixed to a solid support, such as a porous glass wafer. The glass or silica wafer can then be exposed to total cellular RNA or polyA RNA from a sample under conditions in which the affixed sequences will specifically hybridize. Such glass wafers and hybridization methods are widely available, for example, those disclosed by Beattie (WO 95/11755). By examining for the ability of a given probe to specifically hybridize to an RNA sample from an untreated cell population and from a cell population exposed to the agent, agents which up- or downregulate the expression of a nucleic acid encoding Dkk, a Dkk interacting protein, and/or LRP5 can be identified.

Microarray technology and transcriptional profiling are examples of methods which can be used to analyze the impact of putative Dkk or Dkk interacting protein modulating compounds. For transcriptional profiling, mRNA from cells exposed *in vivo* to a potential Dkk modulating agent, such as the Dkk interacting proteins identified in the present invention (e.g., those identified in Figure 5), agents which modulate Dkk interacting proteins, and mRNA from the same type of cells that were not exposed to the agent could be reverse transcribed and hybridized to a chip containing DNA from

numerous genes, to thereby compare the expression of genes in cells treated and not treated with the agent. If, for example a putative Dkk modulating agent down-regulates the expression of Dkk in the cells, then use of the agent may be undesirable in certain patient populations. For additional methods of transcriptional profiling and the use of microarrays, refer to, for example, U.S. Patent No. 6,124,120 issued to Lizardi (2000).

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Additional methods for screening the impact of Dkk and Dkk interacting protein modulating compounds or the impact of Dkk or Dkk interacting proteins on modulation of LRP5, LRP6, HBM or the Wnt pathway include the use of TaqMan PCR, conventional reverse transcriptase PCR (RT-PCR), changes in downstream surrogate markers (*i.e.*, Wnt responsive genes), and anti-Dkk Western blots for protein detection. Other methods would be readily apparent to the artisan of ordinary skill.

12. <u>Methods to Identify Agents that Modulate at Least One Activity of Dkk, a</u> <u>Dkk Interacting Protein, or LRP5/LRP6/HBM</u>

Another embodiment of the present invention provides methods for identifying agents that modulate at least one activity of Dkk, Dkk interacting proteins, and/or LRP5/LRP6/HBM proteins or preferably which specifically modulate an activity of a Dkk/Dkk interacting protein complex or an LRP5(or LRP6/HBM)/Dkk complex, or a biologically active fragment of Dkk (e.g., comprising the domain which binds LRP5/LRP6/HBM) or a Dkk interacting protein complex. Such methods or assays may utilize any means of monitoring or detecting the desired activity as would be known in the art (See, e.g., Wu et al., Curr. Biol. 10:1611-4 (2000); Fedi et al., J. Biol. Chem. 274:19465-72 (1991); Grotewold et al., Mech. Dev. 89:151-3 (1999); Shibata et al., Mech. Dev. 96:243-6 (2000); Wang et al., Oncogene 19:1843-8 (2000); and Glinka et al., Nature 391:357-62 (1998)). Potential agents which modulate Dkk include, for example, p53, the tumor suppressor protein, which can induce Dkk-1. Damage to DNA has also been observed to up-regulate Dkk-1 expression via a stabilization and activation of p53 (Wang et al., Oncogene 19:1843-48 (2000)); and, Shou et al., Oncogene 21:878-89 (2002)). Additionally, Fedi et al. (1999) purportedly showed that Dkk-1 can block the Wnt2-induced oncogenic transformation of NIH-3T3 cells.

Furthermore, it has been suggested that Dkk expression can be modulated by BMP signaling in the developing skeleton (Mukhopadhyay et al., Dev. Cell. 1:423-34 (2001); and Grotewold et al., EMBO J. 21:966-75 (2002)). Grotewold et al. additionally describe altered Dkk expression levels in response to stress signals including UV irradiation and other genotoxic stimuli. They propose that Dkk expression is proapoptotic. In animals expressing HBM constructs conferring high bone mass, a reduced osteoblast apoptosis effect was observed. Thus, HBM and HBM-like variants may control/alter Dkk's role in programmed cell death. Other agents which potentially modulate Dkk activity include the Dkk interacting proteins identified in Figure 5.

In one embodiment, the relative amounts of Dkk or a Dkk interacting protein of a cell population that has been exposed to the agent to be tested is compared to an unexposed control cell population. Antibodies can be used to monitor the differential expression of the protein in the different cell populations. Cell lines or populations are exposed to the agent to be tested under appropriate conditions and time. Cellular lysates may be prepared from the exposed cell line or population and a control, unexposed cell line or population. The cellular lysates are then analyzed with the probe, as would be known in the art. See, e.g., Ed Harlow and David Lane, Antibodies: A Laboratory Manual (Cold Spring Harbor, NY, 1988) and Ed Harlow and David Lane, Using Antibodies: A Laboratory Manual (Cold Spring Harbor, NY, 1998).

For example, N- and C- terminal fragments of Dkk can be expressed in bacteria and used to search for proteins which bind to these fragments. Fusion proteins, such as His-tag or GST fusion to the N- or C-terminal regions of Dkk (or to biologically active domains of Dkk-1) or a whole Dkk protein can be prepared. These fusion proteins can be coupled to, for example, Talon or Glutathione-Sepharose beads and then probed with cell lysates to identify molecules which bind to Dkk. Prior to lysis, the cells may be treated with purified Wnt proteins, RNA, or drugs which may modulate Wnt signaling or proteins that interact with downstream elements of the Wnt pathway. Lysate proteins binding to the fusion proteins can be resolved by SDS-PAGE, isolated and identified by, for example protein sequencing or mass spectroscopy, as is known in the art. See, e.g., Protein Purification Applications: A Practical Approach (Simon Roe, ed., 2nd ed.

Oxford Univ. Press, 2001) and "Guide to Protein Purification" in *Meth. Enzymology* vol. 182 (Academic Press, 1997).

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The activity of Dkk, a Dkk interacting protein, or a complex of Dkk with LRP5/LRP6/HBM may be affected by compounds which modulate the interaction between Dkk and a Dkk interacting protein (such as those shown in Figure 5) and/or Dkk and LRP5/LRP6/HBM. The present invention provides methods and research tools for the discovery and characterization of these compounds. The interaction between Dkk and a Dkk interacting protein and/or Dkk and LRP5/6/HBM may be monitored in vivo and in vitro. Compounds which modulate the stability of a Dkk -LRP5/LRP6/HBM complex are potential therapeutic compounds. Example in vitro methods include: Binding LRP5/6/HBM, Dkk, or a Dkk interacting protein to a sensor chip designed for an instrument such are made by Biacore (Uppsala, Sweden) for the performance of an plasmon resonance spectroscopy observation. In this method, the chip with one of Dkk, a Dkk interacting protein, or LRP5/6 is first exposed to the other under conditions which permit them to form the complex. A test compound is then introduced and the output signal of the instrument provides an indication of any effect exerted by the test compound. By this method, compounds may be rapidly screened. Another, in vitro, method is exemplified by the SAR-by-NMR methods (Shuker et al., Science. 274:1531-4 (1996)). Briefly, a Dkk-1 binding domain and/or LRP 5 or 6 LBD are expressed and purified as ¹⁵N labeled protein by expression in labeled media. The labeled protein(s) are allowed to form the complex in solution in an NMR sample tube. The heteronuclear correlation spectrum in the presence and absence of a test compound provides data at the level of individual residues with regard to interactions with the test compound and changes at the protein-protein interface of the complex. One of skill in the art knows of many other protocols, e.g. affinity capillary electrophoresis (Okun et al. J Biol Chem 276:1057-62 (2001); Vergun and Chu. Methods, 19:270-7 (1999)), fluorescence spectroscopy, electron paramagnetic resonance, etc. which can monitor the modulation of a complex and/or measure binding affinities for complex formation.

In vitro protocols for monitoring the modulation of a Dkk/LRP5/LRP6/HBM complex include the yeast two hybrid protocol. The yeast two hybrid method may be used to monitor the modulation of a complex in vivo by monitoring the expression of genes activated by the formation of a complex of fusion proteins of Dkk and LRP ligand binding domains. Nucleic acids according to the invention which encode the interacting Dkk and LRP LBD domains are incorporated into bait and prey plasmids. The Y2H protocol is performed in the presence of one or more test compounds. The modulation of the complex is observed by a change in expression of the complex activated gene. It will be appreciated by one skilled in the art that test compounds can be added to the assay directly or, in the case of proteins, can be coexpressed in the yeast with the bait and prey compounds. Similarly, fusion proteins of Dkk and Dkk interacting proteins can also be used in a Y2H screen to identify other proteins which modulate the Dkk/Dkk interacting protein complex.

Assay protocols such as these may be used in methods to screen for compounds, drugs, treatments which modulate the Dkk/Dkk interacting protein and/or Dkk/LRP5/6 complex, whether such modulation occurs by competitive binding, or by altering the structure of either LRP 5/6 or Dkk at the binding site, or by stabilizing or destablizing the protein-protein interface. It may be anticipated that peptide aptamers may competitively bind, although induction of an altered binding site structure by steric effects is also possible.

12.1 Antibodies and Antibody Fragments

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Polyclonal and monoclonal antibodies and fragments of these antibodies which bind to Dkk or LRP5/LRP6/HBM can be prepared as would be known in the art. For example, suitable host animals can be immunized using appropriate immunization protocols and the peptides, polypeptides or proteins of the invention. Peptides for use in immunization are typically about 8-40 residues long. If necessary or desired, the polypeptide immunogens can be conjugated to suitable carriers. Methods for preparing immunogenic conjugates with carriers such as bovine serum albumin (BSA), keyhole limpet hemocyanin (KLH), or other carrier proteins are well known in the art (See,

Harlow *et al.*, 1988). In some circumstances, direct conjugation using, for example, carbodiimide reagents, may be effective; in other instances linking reagents such as those supplied by Pierce Chemical Co., Rockford, IL, may be desirable to provide accessibility to the polypeptide or hapten. The hapten peptides can be extended at either the amino or carboxy terminus with a cysteine residue or interspersed with cysteine residues, for example, to facilitate linking to a carrier. Administration of the immunogens is conducted generally by injection over a suitable time period and with use of suitable adjuvants, as is generally understood in the art. During the immunization schedule, titers of antibodies are taken to determine adequacy of antibody formation.

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Anti-peptide antibodies can be generated using synthetic peptides, for example, the peptides derived from the sequence of any Dkk, including Dkk-1, or LRP5/LRP6/HBM. Synthetic peptides can be as small as 2-3 amino acids in length, but are preferably at least 3, 5, 10, or 15 or more amino acid residues long. Such peptides can be determined using programs such as DNAStar. The peptides are coupled to KLH using standard methods and can be immunized into animals such as rabbits. Polyclonal anti-Dkk or anti-LRP5/LRP6/HBM peptide antibodies can then be purified, for example using Actigel beads containing the covalently bound peptide.

While the polyclonal antisera produced in this way may be satisfactory for some applications, for pharmaceutical compositions, use of monoclonal preparations is preferred. Immortalized cell lines which secrete the desired monoclonal antibodies may be prepared using the standard method of Kohler and Milstein or modifications which effect immortalization of lymphocytes or spleen cells, as is generally known (See, e.g., Harlow *et al.*, 1988 and 1998). The immortalized cell lines secreting the desired antibodies can be screened by immunoassay in which the antigen is the peptide hapten, polypeptide or protein. When the appropriate immortalized cell culture secreting the desired antibody is identified, the cells can be cultured either *in vitro* or by production in ascites fluid.

The desired monoclonal antibodies are then recovered from the culture supernatant or from the ascites supernatant. Fragments of the monoclonal antibodies

which contain the immunologically significant portion can be used as agonists or antagonists of Dkk activity. Use of immunologically reactive fragments, such as the Fab, scFV, Fab', of F(ab')₂ fragments are often preferable, especially in a therapeutic context, as these fragments are generally less immunogenic than the whole immunoglobulin.

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The antibodies or fragments may also be produced, using current technology, by recombinant means. Regions that bind specifically to the desired regions of Dkk or LRP5/LRP6/HBM can also be produced in the context of chimeras with multiple species origin. Antibody reagents so created are contemplated for use diagnostically or as stimulants or inhibitors of Dkk activity.

In one embodiment, antibodies against Dkk, bind Dkk with high affinity, i.e., ranging from 10⁻⁵ to 10⁻⁹ M. Preferably, the anti-Dkk antibody will comprise a chimeric, primate, Primatized®, human or humanized antibody. Also, the invention embraces the use of antibody fragments, *e.g.*, Fab's, Fv's, Fab's, F(ab)₂, and aggregates thereof.

Another embodiment contemplates chimeric antibodies which recognize Dkk or LRP5/LRP6/HBM. A chimeric antibody is intended to refer to an antibody with non-human variable regions and human constant regions, most typically rodent variable regions and human constant regions.

A "primatized® antibody" refers to an antibody with primate variable regions, e.g., CDR's, and human constant regions. Preferably, such primate variable regions are derived from an Old World monkey.

A "humanized antibody" refers to an antibody with substantially human framework and constant regions, and non-human complementarity-determining regions (CDRs). "Substantially" refers to the fact that humanized antibodies typically retain at least several donor framework residues (*i.e.*, of non-human parent antibody from which CDRs are derived).

Methods for producing chimeric, primate, primatized®, humanized and human antibodies are well known in the art. See, e.g., U.S. Patent 5,530,101, issued to Queen et al.; U.S. Patent 5,225,539, issued to Winter et al.; U.S. Patents 4,816,397 and

4,816,567, issued to Boss *et al.* and Cabilly *et al.* respectively, all of which are incorporated by reference in their entirety.

The selection of human constant regions may be significant to the therapeutic efficacy of the subject anti-Dkk or LRP5/LRP6/HBM antibody. In a preferred embodiment, the subject anti-Dkk or LRP5/LRP6/HBM antibody will comprise human, gamma 1, or gamma 3 constant regions and, more preferably, human gamma 1 constant regions.

Methods for making human antibodies are also known and include, by way of example, production in SCID mice, and *in vitro* immunization.

The subject anti-Dkk or LRP5/LRP6/HBM antibodies can be administered by various routes of administration, typically parenteral. This is intended to include intravenous, intramuscular, subcutaneous, rectal, vaginal, and administration with intravenous infusion being preferred.

The anti-Dkk or LRP5/LRP6/HBM antibody will be formulated for therapeutic usage by standard methods, e.g., by addition of pharmaceutically acceptable buffers, e.g., sterile saline, sterile buffered water, propylene glycol, and combinations thereof.

Effective dosages will depend on the specific antibody, condition of the patient, age, weight, or any other treatments, among other factors. Typically effective dosages will range from about 0.001 to about 30 mg/kg body weight, more preferably from about 0.01 to 25 mg/kg body weight, and most preferably from about 0.1 to about 20 mg/kg body weight.

Such administration may be effected by various protocols, e.g., weekly, biweekly, or monthly, depending on the dosage administered and patient response. Also, it may be desirable to combine such administration with other treatments.

Antibodies to Dkk-1 interacting proteins, such as those identified in Figure 5, are also contemplated according to the present invention, and can be used similarly to the Dkk-1 antibodies mentioned in the above methodology.

The antibodies of the present invention can be utilized in experimental screening, as diagnostic reagents, and in therapeutic compositions.

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12.2 Chemical Libraries

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Agents that are assayed by these methods can be randomly selected or rationally selected or designed. As used herein, an agent is said to be randomly selected when the agent is chosen randomly without considering the specific sequences involved in the association of Dkk-1 alone, Dkk-1 interacting proteins alone, or with their associated substrates, binding partners, etc. An example of randomly selected agents is the use of a chemical library or a peptide combinatorial library, or a growth broth of an organism.

The agents of the present invention can be, as examples, peptides, small molecules, vitamin derivatives, as well as carbohydrates. A skilled artisan can readily recognize that there is no limit as to the structural nature of the agents of the present invention.

12.3 Peptide Synthesis

The peptide agents of the invention can be prepared using standard solid phase (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides may be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production of polypeptides using solid phase peptide synthesis is necessitated if non-nucleic acid-encoded amino acids are to be included.

13. <u>Uses for Agents that Modulate at Least One Activity of Dkk, a Dkk</u> <u>Interacting Protein, a Dkk/Dkk Interacting Protein Complex, or a Dkk/LRP5 or Dkk/LRP6 Complex</u>

The proteins and nucleic acids of the invention, such as the proteins or polypeptides containing an amino acid sequence of LRP5, Dkk, and Dkk interacting proteins are involved in bone mass modulation and lipid modulation of other Wnt pathway mediated activity. Agents that modulate (*i.e.*, up and down-regulate) the expression of Dkk or Dkk interacting proteins, or agents, such as agonists and

antagonists respectively, of at least one activity of Dkk or a Dkk interacting protein may be used to modulate biological and pathologic processes associated with the function and activity of Dkk or a Dkk interacting protein.

As used herein, a subject can be preferably any mammal, so long as the mammal is in need of modulation of a pathological or biological process modulated by a protein of the invention. The term "mammal" means an individual belonging to the class *Mammalia*. The invention is particularly useful in the treatment of human subjects.

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As used herein, a biological or pathological process modulated by Dkk or a Dkk interacting protein may include binding of Dkk to a Dkk interacting protein, Dkk to LRP5 or LRP6 or release therefrom, inhibiting or activating Dkk or a Dkk interacting protein mRNA synthesis or inhibiting Dkk or Dkk interacting protein modulated inhibition of LRP5 or LRP6 mediated Wnt signaling. Further bone-related markers may be observed such as alkaline phosphatase activity, osteocalcin production, or mineralization.

Pathological processes refer to a category of biological processes which produce a deleterious effect. For example, expression or up-regulation of expression of LRP5 or LRP6 and/or Dkk and/or a Dkk interacting protein may be associated with certain diseases or pathological conditions. As used herein, an agent is said to modulate a pathological process when the agent statistically significantly (p < 0.05) alters the process from its base level in the subject. For example, the agent may reduce the degree or severity of the process mediated by that protein in the subject to which the agent was administered. For instance, a disease or pathological condition may be prevented, or disease progression modulated by the administration of agents which reduce or modulate in some way the expression or at least one activity of a protein of the invention.

As LRP5/6 and Dkk are involved both directly and indirectly in bone mass modulation, one embodiment of this invention is to use Dkk or Dkk interacting protein expression as a method of diagnosing a bone condition or disease. Certain markers are associated with specific Wnt signaling conditions (e.g., TCF/LEF activation). Diagnostic tests for bone conditions may include the steps of testing a sample or an

extract thereof for the presence of Dkk or Dkk interacting protein nucleic acids (*i.e.*, DNA or RNA), oligomers or fragments thereof or protein products of TCF/LEF regulated expression. For example, standard *in situ* hybridization or other imaging techniques can be utilized to observe products of Wnt signaling.

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This invention also relates to methods of modulating bone development or bone loss conditions. Inhibition of bone loss may be achieved by inhibiting or modulating changes in the LRP5/6 mediated Wnt signaling pathway. For example, absence of LRP5 activity may be associated with low bone mass. Increased activity LRP5 may be associated with high bone mass. Therefore, modulation of LRP5 activity will in turn modulate bone development. Modulation of the Dkk/LRP5/6 or Dkk/Dkk interacting protein complex via agonists and antagonists is one embodiment of a method to regulate bone development. Such modulation of bone development can result from inhibition of the activity of, for example, a Dkk/LRP(5/6) protein complex, a Dkk/Dkk interacting protein complex, upregulated transcription of the *LRP5* gene or inhibited translation of Dkk or Dkk interacting protein mRNA.

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The agents of the present invention can be provided alone, or in combination with other agents that modulate a particular pathological process. As used herein, two agents are said to be administered in combination when the two agents are administered simultaneously or are administered independently in a fashion such that the agents will act at the same time.

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The agents of the present invention can be administered via parenteral, subcutaneous (sc), intravenous (iv), intramuscular (im), intraperitoneal (ip), transdermal or buccal routes. Alternatively, or concurrently, administration may be by the oral route. The dosage administered will be dependent upon the age, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect desired.

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The present invention further provides compositions containing one or more agents which modulate expression or at least one activity of a protein of the invention. While individual needs vary, determination of optimal ranges of effective amounts of each component is within the skill of the art. Typical dosages of the active agent which

mediate Dkk or Dkk interacting protein activity comprise from about 0.0001 to about 50 mg/kg body weight. The preferred dosages comprise from about 0.001 to about 50 mg/kg body weight. The most preferred dosages comprise from about 0.1 to about 1 mg/kg body weight. In an average human of 70 kg, the range would be from about 7 µg to about 3.5 g, with a preferred range of about 0.5 mg to about 5 mg.

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In addition to the pharmacologically active agent, the compositions of the present invention may contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically for delivery to the site of action. Suitable formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form, for example, water-soluble salts. In addition, suspensions of the active compounds as appropriate oily injection suspensions may be administered. Suitable lipophilic solvents or vehicles include fatty oils, for example, sesame oil, or synthetic fatty acid esters, (e.g., ethyl oleate or triglycerides). Aqueous injection suspensions may contain substances which increase the viscosity of the suspension include, for example, sodium carboxymethyl cellulose, sorbitol and/or dextran. Optionally, the suspension may also contain stabilizers. Liposomes and other non-viral vectors can also be used to encapsulate the agent for delivery into the cell.

The pharmaceutical formulation for systemic administration according to the invention may be formulated for enteral, parenteral, or topical (top) administration. Indeed, all three types of formulations may be used simultaneously to achieve systemic administration of the active ingredient.

Suitable formulations for oral administration include hard or soft gelatin capsules, pills, tablets, including coated tablets, elixirs, suspensions, syrups or inhalations and controlled release forms thereof.

Potentially, any compound which binds Dkk or a Dkk interacting protein or modulates the Dkk/LRP5 or Dkk/LRP6 or Dkk/Dkk interacting protein complex may be a therapeutic compound. In one embodiment of the invention, a peptide or nucleic acid aptamer according to the invention is used in a therapeutic composition. Such compositions may comprise an aptamer, or a LRP5 or LRP6 fragment unmodified or

modified. In another embodiment, the therapeutic compound comprises a Dkk-1 interacting protein, or biologically active fragment thereof.

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Nucleic acid aptamers have been used in compositions for example by chemical bonding to a carrier molecule such as polyethylene glycol (PEG) which may facilitate uptake or stabilize the aptamer. A di-alkylgylcerol moiety attached to an RNA will embed the aptamer in liposomes, thus stabilizing the compound. Incorporating chemical substitutions (i.e. changing the 2'OH group of ribose to a 2'NH in RNA confers ribonuclease resistance) and capping, etc. can prevent breakdown. Several such techniques are discussed for RNA aptamers in Brody and Gold (Rev. Mol. Biol. 74:3-13 (2000)).

Peptide aptamers may by used in therapeutic applications by the introduction of an expression vector directing aptamer expression into the affected tissue such as for example by retroviral delivery, by encapsulating the DNA in a delivery complex or simple by naked DNA injection. Or, the aptamer itself or a synthetic analog may be used directly as a drug. Encapsulation in polymers and lipids may assist in delivery. The use of peptide aptamers as therapeutic and diagnostic agents is reviewed by Hoppe-Syler and Butz (*J. Mol. Med.* 78:426-430 (2000)).

In another aspect of the invention. The structure of a constrained peptide aptamer of the invention may be determined such as by NMR or X-ray crystallography. (Cavanagh et al., Protein NMR Spectroscopy: Principles and Practice, Academic Press, 1996; Drenth, Principles of Protein X-Ray Crystallography, Springer Verlag, 1999) Preferably the structure is determined in complex with the target protein. A small molecule analog is then designed according to the positions of functional elements of the 3D structure of the aptamer. (Guidebook on Molecular Modeling in Drug Design, Cohen, Ed., Academic Press, 1996; Molecular Modeling and Drug Design (Topics in Molecular and Structural Biology), Vinter and Gardner Eds., CRC Press, 1994) Thus the present invention provides a method for the design of effective and specific drugs which modulate the activity of Dkk, Dkk interacting proteins, Dkk/Dkk interacting protein complex and the Dkk/LRP complex. Small molecule mimetics of the peptide aptamers of the present invention are encompassed within the scope of the invention.

In practicing the methods of this invention, the compounds of this invention may be used alone or in combination, or in combination with other therapeutic or diagnostic agents. In certain preferred embodiments, the compounds of this invention may be coadministered along with other compounds typically prescribed for these conditions according to generally accepted medical practice. For example, the compounds of this invention can be administered in combination with other therapeutic agents for the treatment of bone loss. Bone loss mediating agents include bone resorption inhibitors such as bisphosphonates (e.g., alendronic acid, clodronic acid, etidronic acid, pamidronic acid, risedronic acid and tiludronic acid), vitamin D and vitamin D analogs, cathepsin K inhibitors, hormonal agents (e.g., calcitonin and estrogen), and selective estrogen receptor modulators or SERMs (e.g., raloxifene). And bone forming agents such as parathyroid hormone (PTH) and bone morphogenetic proteins (BMP).

Additionally contemplated are combinations of agents which regulate Dkk-1 and agents which regulate lipid levels such as HMG-CoA reductase inhibitors (*i.e.*, statins such as Mevacor®, Lipitor® and other inhibitors such as Baycol®, Lescol®, Pravachol® and Zocor®), bile acid sequestrants (e.g., Colestid® and Welchol®), fibric acid derivatives (Atromid-S®, Lopid®, Tricor®), and nicotinic acid.

[0001] The compounds of this invention can be utilized *in vivo*, ordinarily in vertebrates and preferably in mammals, such as humans, sheep, horses, cattle, pigs, dogs, cats, rats and mice, or *in vitro*.

14. <u>Transgenic Animals</u>

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Transgenic animal models can be created which conditionally express Dkk and/or LRP5 or LRP6 and/or Dkk interacting proteins, such as those shown in Figure 5. These animals can be used as research tools for the study of the physiological effects of the Dkk-1/Dkk-1 interacting protein interaction and/or the LRP5 / Dkk interaction. Alternatively, transgenic animals can be created which express a transgenic form of Dkk alone or in addition to a transgenic form of HBM or express Dkk interacting proteins alone or in addition to a transgenic form of Dkk. Transgenic animals expressing HBM or LRP5 can be crossed with transgenic animals expressing Dkk or

Dkk interacting proteins to obtain heterozygote as well as homozygote animals which express both desired genes.

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Animal models may be created to directly modulate the Dkk/Dkk interacting protein or Dkk/ LRP5 interaction activity in vivo to serve as a research tool for determining the efficacy of candidate compounds which modulate the Dkk/Dkk interacting protein or LRP5 / Dkk interaction activity in vitro. Animals, such as transgenic mice, can be created using the techniques employed to make transgenic mice that express for example, human Dkk or a Dkk interacting protein, or knockouts (KO), which may be conditional, of the gene encoding mouse Dkk or Dkk interacting protein. Knock-in animals include animals wherein genes have been introduced and animals wherein a gene that was previously knocked-out is reintroduced into the animal. Other transgenic animals can be created with inducible forms of Dkk or a Dkk interacting protein to study the effects of the gene on bone mass development and loss as well as lipid level regulation. These animals can also be used to study long term effects of Dkk or Dkk interacting protein modulation. Transgenic animals may be created to express peptide aptamers, or produce RNA aptamers. The transgenic vectors may direct expression in a tissue specific manner by the use of tissue specific promoters. In a preferred embodiment, a peptide aptamer fusion protein is expressed using a bone specific promoter. Such systems can provide a tissue specific knock-out of Dkk or Dkk interacting protein activity.

General methods for creating transgenic animals are known in the art, and are described in, for example, Strategies in Transgenic Animal Science (Glenn M. Monastersky and James M. Robl eds., ASM Press; Washington, DC, 1995);

Transgenic Animal Technology: A Laboratory Handbook (Carl A. Pinkert ed., Academic Press 1994); Transgenic Animals (Louis Marie Houdebine, ed., Harwood Academic Press, 1997); Overexpression and Knockout of Cytokines in Transgenic Mice (Chaim O. Jacob, ed., Academic Press 1994); Microinjection and Transgenesis; Strategies and Protocols (Springer Lab Manual) (Angel Cid-Arregui and Alejandro Garcia-Carranca, eds., Springer Verlag 1998); and Manipulating the Mouse Embryo: A Laboratory Manual (Brigid Hogan et al., eds., Cold Spring Harbor Laboratory Press 1994).

15. Peptide and Nucleotide Aptamers and Peptide Aptamer Mimetics

Another embodiment contemplates the use of peptide and nucleotide aptamer technology to screen for agents which interact with Dkk, which block Dkk from interacting with LRP5 or LRP6, or which block any other Dkk ligand interaction, or which interact with Dkk interacting proteins, such as those shown in Figure 5. Peptide aptamers are molecules in which a variable peptide domain is displayed from a scaffold protein. Thioredoxin A (trxA) is commonly used for a scaffold. The peptide insert destroys the catalytic site of trxA. It is recognized that numerous proteins may also be used as scaffolding proteins to constrain and/or present a peptide aptamer. Other scaffold proteins that could display a constrained peptide aptamer could include staphylococcal nuclease, the protease inhibitor eglin C, the *Streptomyces tendea* alphaamylase inhibitor Tendamistat, Sp1, and green fluorescent protein (GFP) (reviewed in Hoppe-Seyler *et al.*, *J. Steroid Biochem Mol. Biol.* 78:105-11 (2001)), and the S1 nuclease from *Staphylococcus* or M13 for phage display. Any molecule to which the aptamer could be anchored and presented in its bioactive conformation would be suitable.

Aptamers can then specifically bind to a given target protein *in vitro* and *in vivo* and have the potential to selectively block the function of their target protein. Peptide aptamers are selected from randomized expression libraries on the basis of their *in vivo* binding capacity to the desired target protein. Briefly, a target protein (e.g., Dkk, a Dkk interacting protein, or LRP5/6) is linked to a heterologous DNA binding domain (BD) and expressed as bait in a yeast test strain. Concomitantly, a library coding for different peptides (e.g., 16-mers) of randomized sequence inserted in a scaffold protein sequence, which are linked to a heterologous transcriptional activation domain (AD) is expressed as prey. If a peptide binds to a target protein, a functional transcription factor is reconstituted, in which the BD and AD are bridged together by interacting proteins. This transcription factor is then able to activate the promoter of a marker gene which can be monitored by colorimetric enzymatic assays or by growth selection. Additional variation, methods of preparing and screening methodologies are described in, for example, Hoppe-Seyler *et al.*, *J. Mol. Med.* 78: 426-430 (2000).

Nucleotide aptamers are described for example in Brody et al., Trends Mol. Biotechnol. 74: 5-13 (2000). Additional methods of making and using nucleotide aptamers include SELEX, i.e., Systematic Evolution of Ligands by Exponential Enrichment. SELEX is a process of isolating oligonucleotide ligands of a chosen target molecule (see Tuerk and Gold, Science 249:505-510 (1990); U.S. Pat. Nos. 5,475,096, 5,595,877, and 5,660,985). SELEX, as described in Tuerk and Gold, involves admixing the target molecule with a pool of oligonucleotides (e.g., RNA) of diverse sequences; retaining complexes formed between the target and oligonucleotides; recovering the oligonucleotides bound to the target; reverse-transcribing the RNA into DNA; amplifying the DNA with polymerase chain reactions (PCR); transcribing the amplified DNA into RNA; and repeating the cycle with ever increasing binding stringency. Three enzymatic reactions are required for each cycle. It usually takes 12-15 cycles to isolate aptamers of high affinity and specificity to the target. An aptamer is an oligonucleotide that is capable of binding to an intended target substance but not other molecules under the same conditions.

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In another reference, Bock et al., Nature 355:564-566 (1990), describe a different process from the SELEX method of Tuerk and Gold in that only one enzymatic reaction is required for each cycle (i.e., PCR) because the nucleic acid library in Bock's method is comprised of DNA instead of RNA. The identification and isolation of aptamers of high specificity and affinity with the method of Bock et al. still requires repeated cycles in a chromatographic column.

Other nucleotide aptamer methods include those described by Conrad *et al.*, *Meth. Enzymol.* 267:336-367 (1996). Conrad *et al.* describe a variety of methods for isolating aptamers, all of which employ repeated cycles to enrich target-bound ligands and require a large amount of purified target molecules. More recently described methods of making and using nucleotide aptamers include, but are not limited to those described in U.S. Patent Nos. 6,180,348; 6,051,388; 5,840,867; 5,780,610, 5,756,291 and 5,582,981.

Potentially, any compound which binds Dkk or a Dkk interacting protein or modulates the Dkk/Dkk interacting protein or Dkk/LRP5 or Dkk/LRP6 complex may be

a therapeutic compound. In one embodiment of the invention, a peptide or nucleic acid aptamer according to the invention is used in a therapeutic composition. Such compositions may comprise an aptamer, or a LRP5 or LRP6 fragment unmodified or modified.

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Nucleic acid aptamers have been used in compositions for example by chemical bonding to a carrier molecule such as polyethylene glycol (PEG) which may facilitate uptake or stabilize the aptamer. A di-alkylglycerol moiety attached to an RNA will embed the aptamer in liposomes, thus stabilizing the compound. Incorporating chemical substitutions (*i.e.*, changing the 2'-OH group of ribose to a 2'-NH in RNA confers ribonuclease resistance) and capping, etc. can prevent breakdown. Several such techniques are discussed for RNA aptamers in Brody and Gold *Rev. Mol. Biol.* 74:3-13 (2000).

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Peptide aptamers may by used in therapeutic applications by the introduction of an expression vector directing aptamer expression into the affected tissue such as for example by retroviral delivery, by encapsulating the DNA in a delivery complex or simple by naked DNA injection. Or, the aptamer itself or a synthetic analog may be used directly as a drug. Encapsulation in polymers and lipids may assist in delivery. The use of peptide aptamers as therapeutic and diagnostic agents is reviewed by Hoppe-Syler and Butz *J. Mol. Med.* 78:426-430 (2000).

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In another aspect of the invention, the structure of a constraine'd peptide aptamer of the invention may be determined such as by NMR or X-ray crystallography. (Cavanagh et al., Protein NMR Spectroscopy: Principles and Practice, Academic Press, 1996; Drenth, Principles of Protein X-Ray Crystallography, Springer Verlag, 1999) Preferably the structure is determined in complex with the target protein. A small molecule analog is then designed according to the positions of functional elements of the 3D structure of the aptamer. (Guidebook on Molecular Modeling in Drug Design, Cohen, Ed., Academic Press, 1996; Molecular Modeling and Drug Design (Topics in Molecular and Structural Biology), Vinter and Gardner Eds., CRC Press, 1994) Thus, a method is provided for the design of effective and specific drugs which modulate the activity of Dkk, Dkk interacting proteins, Dkk/Dkk interacting protein

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complex, and the Dkk/LRP complex. Small molecule mimics of the peptide aptamers of the present invention are also encompassed within the scope of the invention.

16. <u>Alternative Variants of LRP5/LRP6 Having HBM Activity</u>

A structural model of the LRP5/Zmax1 first beta-propeller module was generated based on a model prediction in Springer et al., (1998) *J. Molecular Biology*, 283:837-862. Based on the model, certain amino acid residues were identified as important variants of LRP5/HBM/Zmax1. The following three categories provide examples of such variants:

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The shape of the beta-propeller resembles a disk with inward-sloping sides and a hole down the middle. Residue 171 is in a loop on the outer or top surface of the domain in blade 4 of propeller module 1. Thus, variants comprising changed residues in structurally equivalent positions in other blades; as well as residues that are slightly more interior to the binding pocket, but still accessible to the surface, are important embodiments of the present invention for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention. The following are examples of such variants:

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A214V (a position equivalent to 171 in blade 5; alanine is not conserved in other propellers),

E128V (a position equivalent to 171 in blade 3; glutamate is not conserved in other propellers),

A65V (a position equivalent to 171 in blade 2; alanine is conserved in propellers 1-3 but not 4),

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G199V (an accessible interior position in blade 5; glycine is conserved in propellers 1-3 but not 4), and M282V (accessible interior position in blade 1; methionine is conserved in propellers 1-3 but not 4).

LRP5/Zmax1 has four beta-propeller structures; the first three beta-propeller modules conserve a glycine in the position corresponding to residue 171 in human

LRP5/Zmax1. Therefore, variants bearing a valine in the equivalent positions in the other propellers are important embodiments of the present invention. The following variants are useful for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention: G479V, G781V, and Q1087V.

The G171V HBM polymorphism results in "occupied space" of the beta-propeller 1, with the side-chain from the valine residue sticking out into an open binding pocket and potentially altering a ligand/protein interaction. The glycine residue is conserved in LRP5/Zmax1 propellers 1, 2 and 3 but is a glutamine in propeller 4. Therefore, the following variants of LRP5/HBM are important embodiments of the present invention for the study of bone mass modulation by LRP5/HBM, for the development of pharmaceuticals and treatments of bone mass disorders, and for other objectives of the present invention:

G171K (which introduces a charged side-chain),

G171F (which introduces a ringed side-chain),

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G1711 (which introduces a branched side-chain), and

G171Q (which introduces the propeller 4 residue).

Furthermore, LRP6 is the closest homolog of LRP5/Zmax1. LRP6 has a betapropeller structure predicted to be similar, if not identical to Zmax1. The position
corresponding to glycine 171 in human LRP5/Zmax1 is glycine 158 of human LRP6.
Thus, corresponding variants of LRP6 are an important embodiment of the present
invention for the study of the specificity of LRP5/Zmax1 versus its related family
member, for the development of pharmaceuticals and treatments of bone mass
disorders, and for other objectives of the present invention. Specifically, for example, a
glycine to valine substitution at the structurally equivalent position, residue 158, of
human LRP6 and similar variants of other species' LRP6 homologs represent important
research tools.

Site-directed mutants of LRP5 were generated in the full-length human LRP5 cDNA using the QuikChange XL-Site-Directed Mutagenesis Kit (catalog #200516,

Stratagene, La Jolla, CA) following the manufacturer's protocol. The mutant sequences were introduced using complementary synthetic oligonucleotides:

A65V: TGGTCAGCGGCCTGGAGGATGTGGCCGCAGTGGACTTCC (SEQ ID NO:129) and 5 GGAAGTCCACTGCGGCCACATCCTCCAGGCCGCTGACCA (SEQ ID NO:130) E128V: AAGCTGTACTGGACGGACTCAGTGACCAACCGCATCGAGG (SEQ ID NO:131) and CCTCGATGCGGTTGGTCACTGAGTCCGTCCAGTACAGCTT (SEQ ID 10 NO:132) G171K: ATGTACTGGACAGACTGGAAGGAGACGCCCCGGATTGAGCG (SEQ ID NO: 133) and CGCTCAATCCGGGGCGTCTCCTTCCAGTCTGTCCAGTACAT (SEQ ID NO:134) 15 G171F: ATGTACTGGACAGACTGGTTTGAGACGCCCCGGATTGAGCG (SEQ ID NO:135) and CGCTCAATCCGGGGCGTCTCAAACCAGTCTGTCCAGTACAT (SEQ ID NO:136) G1711: ATGTACTGGACAGACTGGATTGAGACGCCCCGGATTGAGCG (SEQ 20 ID NO:137) and CGCTCAATCCGGGGCGTCTCAATCCAGTCTGTCCAGTACAT (SEQ ID NO:138) G171Q: ATGTACTGGACAGACTGGCAGGAGACGCCCCGGATTGAGCG (SEQ ID NO:139) and 25 CGCTCAATCCGGGGCGTCTCCTGCCAGTCTGTCCAGTACAT (SEQ ID NO:140) G199V: CGGACATTTACTGGCCCAATGTACTGACCATCGACCTGGAGG (SEQ ID NO:141) and

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NO:142)

CCTCCAGGTCGATGGTCAGTACATTGGGCCAGTAAATGTCCG (SEQ ID

A214V: AGCTCTACTGGGCTGACGTCAAGCTCAGCTTCATCCACCG (SEQ ID NO: 143) and CGGTGGATGAAGCTGAGCTTGACGTCAGCCCAGTAGAGCT (SEQ ID NO:144) 5 M282V: GAGTGCCCTCTACTCACCCGTGGACATCCAGGTGCTGAGCC (SEQ ID NO:145) and GGCTCAGCACCTGGATGTCCACGGGTGAGTAGAGGGCACTC (SEQ ID NO:146) G479V: CATGTACTGGACAGACTGGGTAGAGAACCCTAAAATCGAGTGTGC 10 (SEQ ID NO:147) and GCACACTCGATTTTAGGGTTCTCTACCCAGTCTGTCCAGTACATG (SEQ ID NO:148) G781V: CATCTACTGGACCGAGTGGGTCGGCAAGCCGAGGATCGTGCG (SEQ ID NO:149) and 15 CGCACGATCCTCGGCTTGCCGACCCACTCGGTCCAGTAGATG (SEQ ID NO:150) Q1087V: GTACTTCACCAACATGGTGGACCGGGCAGCCAAGATCGAACG (SEQ ID NO:151) and CGTTCGATCTTGGCTGCCCGGTCCACCATGTTGGTGAAGTAC (SEQ ID 20 NO:152) LRP6 G158V: GTACTGGACAGACTGGGTAGAAGTGCCAAAGATAGAACGTGC (SEQ ID NO:153) and GCACGTTCTATCTTTGGCACTTCTACCCAGTCTGTCCAGTAC (SEQ ID 25 NO:154). All constructs were sequence verified to ensure that only the engineered modification was present in the gene. Once verified, each variant was functionally evaluated in the TCF-luciferase assay in U2OS cells (essentially as described in Example 7. Other functional evaluations could also be performed, such as the Xenopus

embryo assay (essentially as described in Example 6), or other assays to evaluate Wnt

signaling, Dkk modulation, or anabolic bone effect. Binding of these mutants to Dkk, LRP-interacting proteins, Dkk-interacting proteins, or peptide aptamers to any of the preceding could also be investigated in a variety of ways such as in a two-hybrid system (such as in yeast as described in this application), or other methods.

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Figure 24 shows the effects of the G171F mutation in propeller 1 of LRP5. This mutation is at the same position as HBM's G171V substitution. Expression of G171F results in an HBM effect. That is, in the presence of Wnt, G171F is able to activate the TCF-luciferase reporter construct. In fact, it may activate the reporter to a greater extent than either LRP5 or HBM. Furthermore, in the presence of Dkk1 and Wnt1, G171F is less susceptible than LRP5 to modulation by Dkk. These data exemplify that the G171F variant modulates Wnt signaling in a manner similar to HBM. In addition, this data confirms that HBM's valine residue at 171 is not the only modification at 171 that can result in an HBM effect. Together these data support an important role for LRP5 propeller 1 in modulating Wnt pathway activity; in responding to Dkk modulation; and, in the ability to generate an HBM effect.

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Figure 25 shows the effects of the M282V mutation in propeller 1 of LRP5. M282 expression results in an HBM-effect. That is, in the presence of Wnt, M282 is able to activate the TCF-luciferase reporter construct. Furthermore, in the presence of Dkk1 and Wnt1, M282V is less susceptible than LRP5 to modulation by Dkk. These data show that the M282V variant modulates Wnt signaling in a manner similar to HBM. In addition, this data confirms that modifications of other residues in propeller 1 of LRP5 can result in an HBM effect.

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These data support an "occupied space" model of the HBM mutation in propeller 1 and show that multiple mutations of propeller 1 are capable of generating an HBM effect; the original G171V HBM mutation is not unique in this ability. Moreover, various perturbations in propeller 1 can modulate Dkk activity.

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These data illustrate the molecular mechanism of Dkk modulation of LRP signaling. Using the methods disclosed herein and in U.S. Application 60/290,071, generation of a comprehensive mutant panel will reveal residues in LRP that function in Dkk modulation of Wnt signaling. Such variants of LRP5 and LRP6 that modulate Dkk

activity and the residues which distinguish them from LRP5 and LRP6 are points for therapeutic intervention by small molecule compound, antibody, peptide aptamer, or other agents. Furthermore, models of each HBM-effect mutation/polymorphism may be used in rational drug design of an HBM mimetic agent.

These are only a few illustrative examples presented to better describe the present invention. Variants of LRP5 which have demonstrated HBM activity in assays include G171F, M282V, G171K, G171Q and A214V. Clearly, other variants may be contemplated within the scope of the present invention. Furthermore, wherever HBM is recited in the methods of the invention, it should be understood that any such alternative variant of LRP which demonstrates HBM biological activity is also encompassed by those claims.

17. <u>Screening Assays</u>

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The two-hybrid system is extremely useful for studying protein:protein interactions. See, e.g., Chien et al., Proc. Natl Acad. Sci. USA 88:9578-82 (1991); Fields et al., Trends Genetics 10:286-92 (1994); Harper et al., Cell 75:805-16 (1993); Vojtek et al., Cell 74:205-14 (1993); Luban et al., Cell 73:1067-78 (1993); Li et al., FASEB J. 7:957-63 (1993); Zang et al., Nature 364:308-13 (1993); Golemis et al., Mol. Cell. Biol. 12:3006-14 (1992); Sato et al., Proc. Natl Acad. Sci. USA 91:9238-42 (1994); Coghlan et al., Science 267:108-111 (1995); Kalpana et al., Science 266:2002-6 (1994); Helps et al., FEBS Lett. 340:93-8 (1994); Yeung et al., Genes & Devel. 8:2087-9 (1994); Durfee et al., Genes & Devel. 7:555-569 (1993); Paetkau et al., Genes & Devel. 8:2035-45; Spaargaren et al., 1994 Proc. Natl. Acad. Sci. USA 91:12609-13 (1994); Ye et al., Proc. Natl Acad. Sci. USA 91:12629-33 (1994); and U.S. Patent Nos. 5,989,808; 6,251,602; and 6,284,519.

Variations of the system are available for screening yeast phagemid (see, e.g., Harper, Cellular Interactions and Development: A Practical Approach, 153-179 (1993); Elledge et al., Proc. Natl Acad. Sci. USA 88:1731-5 (1991)) or plasmid (Bartel, 1993 and Bartel, Cell 14:920-4 (1993)); Finley et al., Proc. Natl Acad. Sci. USA 91:12980-4

(1994)) cDNA libraries to clone interacting proteins, as well as for studying known protein pairs.

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The success of the two-hybrid system relies upon the fact that the DNA binding and polymerase activation domains of many transcription factors, such as GAL4, can be separated and then rejoined to restore functionality (Morin *et al.*, *Nuc. Acids Res.* 21:2157-63 (1993)). While these examples describe two-hybrid screens in the yeast system, it is understood that a two-hybrid screen may be conducted in other systems such as mammalian cell lines. The invention is therefore not limited to the use of a yeast two-hybrid system, but encompasses such alternative systems.

Yeast strains with integrated copies of various reporter gene cassettes, such as

for example GAL.fwdarw.LacZ, GAL.fwdarw.HIS3 or GAL.fwdarw.URA3 (Bartel, in Cellular Interactions and Development: A Practical Approach, 153-179 (1993); Harper et al., Cell 75:805-16 (1993); Fields et al., Trends Genetics 10:286-92 (1994)) are cotransformed with two plasmids, each expressing a different fusion protein. One plasmid encodes a fusion between protein "X" and the DNA binding domain of, for example, the GAL4 yeast transcription activator (Brent et al., Cell 43:729-36 (1985); Ma et al., Cell 48:847-53 (1987); Keegan et al., Science 231:699-704 (1986)), while the other plasmid encodes a fusion between protein "Y" and the RNA polymerase activation domain of GAL4 (Keegan et al., 1986). The plasmids are transformed into a strain of the yeast that contains a reporter gene, such as lacZ, whose regulatory region contains GAL4 binding sites. If proteins X and Y interact, they reconstitute a functional GAL4 transcription activator protein by bringing the two GAL4 components into sufficient proximity to activate transcription. It is well understood that the role of bait and prey

Either hybrid protein alone must be unable to activate transcription of the reporter gene, the DNA-binding domain hybrid, because it does not provide an activation function, and the activation domain hybrid, because it cannot localize to the GAL4 binding sites. Interaction of the two test proteins reconstitutes the function of GAL4 and results in expression of the reporter gene. The reporter gene cassettes

proteins may be alternatively switched and thus the embodiments of this invention

contemplate and encompass both alternative arrangements.

consist of minimal promoters that contain the GAL4 DNA recognition site (Johnson *et al.*, *Mol. Cell. Biol.* 4:1440-8 (1984); Lorch *et al.*, *J. Mol. Biol.* 186:821-824 (1984)) cloned 5' to their TATA box. Transcription activation is scored by measuring either the expression of β-galactosidase or the growth of the transformants on minimal medium lacking the specific nutrient that permits auxotrophic selection for the transcription product, *e.g.*, URA3 (uracil selection) or HIS3 (histidine selection). See, *e.g.*, Bartel, 1993; Durfee *et al.*, *Genes & Devel.* 7:555-569 (1993); Fields *et al.*, *Trends Genet.* 10:286-292 (1994); and U.S. Pat. No. 5,283,173.

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Generally, these methods include two proteins to be tested for interaction which are expressed as hybrids in the nucleus of a yeast cell. One of the proteins is fused to the DNA-binding domain (DBD) of a transcription factor and the other is fused to a transcription activation domain (AD). If the proteins interact, they reconstitute a functional transcription factor that activates one or more reporter genes that contain binding sites for the DBD. Exemplary two-hybrid assays which have been used for Dkk-1/LRP5 are presented in the Examples below.

Additional methods of preparing two hybrid assay systems for Dkk-1 interactors would be evident to one of ordinary skill in the art. See for example, Finley *et al.*, "Two-Hybrid Analysis of Genetic Regulatory Networks," in The Yeast Two-Hybrid System (Paul L. Bartel et al., eds., Oxford, 1997); Meijia Yang, "Use of a Combinatorial Peptide Library in the Two-Hybrid Assay," in The Yeast Two-Hybrid System (Paul L. Bartel et al., eds., Oxford, 1997); Gietz *et al.*, "Identification of proteins that interact with a protein of interest: Applications of the yeast two-hybrid system," *Mol. & Cell. Biochem.* 172:67-9 (1997); K. H. Young, "Yeast Two-Hybrid: So Many Interactions,(in) so Little Time," *Biol. Reprod.* 58:302-311 (1998); R. Brent *et al.*, "Understanding Gene and Allele Function with Two-Hybrid Methods," *Annu. Rev. Genet.* 31:663-704 (1997). It will be appreciated that protein networks can be elucidated by performing sequential screens of activation domain-fusion libraries.

Without further description, it is believed that one of ordinary skill in the art can, using the preceding description and the following illustrative examples, make and utilize the compounds of the present invention and practice the claimed methods. The

following working examples therefore, specifically point out preferred embodiments of the present invention, and are not to be construed as limiting in any way the remainder of the disclosure.

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EXAMPLES

The present invention is described by reference to the following Examples, which are offered by way of illustration and are not intended to limit the invention in any manner. Standard techniques well-known in the art or the techniques specifically described below were utilized.

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For routine practice of the protocols referenced below, one of skill in the art is directed to the references cited in this application as well as the several <u>Current Protocol</u> guides, which are continuously updated, widely available and published by John Wiley and Sons, (New York). In the life sciences, <u>Current Protocols</u> publishes comprehensive manuals in Molecular Biology, Immunology, Human Genetics, Protein Science, Cytometry, Neuroscience, Pharmacology, Cell Biology, Toxicology, and Nucleic Acid Chemistry. Additional sources are known to one of skill in the art.

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Example 1

Yeast Two Hybrid Screen Using LRP5 Ligand Binding Domain (LBD) Bait Sequences

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In a screen against human osteoblast library (*i.e.*, HOB03C5, a custom Gibco generated Y2H compatible cDNA library from a human osteoblast cell line as described by Bodine and Komm, *Bone* 25:535-43 (1999)), an interaction with Dkk-1 was identified. The LRP5 ligand binding domain (LBD) baits used for this screen are depicted in Figures 2B and C. The basic protocol is as follows:

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An overnight culture of the yeast strain containing the bait of interest is grown in 20 ml of appropriate selective medium containing 2% glucose at 30°C. The overnight culture is diluted by a 10 fold factor into YPDmedia supplemented with 40 mg/l of adenine, and grown for 4 hours at 30°C.

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For each mating event, an aliquot of the frozen prey library is grown in 150 ml YAPD medium for 5 hours at 30°C.

Appropriate volumes calculated by measuring the OD600 of each culture are combined into a tube. The number of diploids to be screened is typically ten times the number of clones originally present in the prey library of interest. Assuming a mating efficiency of 20% minimum, fifty times (*i.e.*, ten times coverage multiplied by 20% mating efficiency) as many haploid cells containing the bait and as many cells containing the prey are used in any given mating event. The mixture is filtered over a 47 mm, 0.45 mm sterile Metricel filter membrane (Gelman).

Using sterile forceps, the filter is transferred onto a 100 mm² YAPD agar plate with the cell side up, removing all air bubbles underneath the filter. The plate is incubated overnight at room temperature.

The filter is transferred into a 50 ml Falcon tube using sterile forceps and 10 ml SD medium containing 2% glucose are added to resuspend the cells. The filter, once free of cells, is removed and the cell suspension is spun for 5 min. at 2,000 xg.

The cells are resuspended in 10 ml SD medium containing 2% glucose. An aliquot of 100 μ l is set aside for titration.

The cells are plated onto large square plates containing appropriate selective media and incubated at 30°C for three to five days.

To calculate the mating efficiency and to determine the total number of diploid cells screened, the 100 μ l aliquot set aside for titration is diluted and plated onto different selective media. The mating efficiency is calculated by dividing the number of diploids/ml by the lowest number of haploids/ml, either bait or prey, and multiplied by 100. For example, if 2 million diploids were obtained by mating 10 million of haploids containing a bait and 12 million of haploids containing a prey, then the mating efficiency is calculated by dividing 2 million by 10 million, which equals 0.2 and multiplied by 100 which equals 20%. Typical mating efficiencies under the above conditions are within about 20 to about 40%. The total number of diploids screened in a mating event is obtained by multiplying the number of diploids/ml by the total number of ml plated, typically about 10.

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Isolation of colonies containing pairs of interacting proteins.

Yeast colonies from the interaction selection (large square) plates are picked with a sterile toothpick and patched onto plates containing the appropriate selective media and incubated at 30°C for two days.

To further ensure purity of the yeast, the plates are replicated onto another plate containing the same media and incubated at 30°C for another two days.

Yeast patches are scraped using a sterile toothpick and placed into a 96-well format plate containing 100 μ l SD –L –W –H with 2% glucose liquid medium.

Half the volume of the plate is transferred to a 96-well plate containing 50 μ l of 40% glycerol for storage. The other half is set aside for replication and galactosidase-activity assay (see below).

Cells are replicated onto a SD –L –W –H plate with 2% glucose plate to create a master plate, and incubated two days at 30°C. The master plate is replicated onto different selective media to score the strength of each interaction.

Cells are also replicated onto media selecting for the prey vector only for colony PCR and incubated two days at 30°C.

Galactosidase activity assay

Ten microliters from the 96-well plate (set aside from above) are transferred into another 96-well plate containing 100 μ l SD and 2% glucose media. The cell density is measured at OD₆₀₀ using a spectrophotometer, the OD₆₀₀ is usually between 0.03 and 0.1. Fifty microliters of Galactosidase reaction mixture (Tropix) are added to microplates (Marsh) specifically designed for the luminometer (Hewlett Packard Lumicount). Fifty microliters of the diluted cells are then added and mixed by pipetting. The reaction is incubated sixty to one hundred twenty minutes at room temperature. Relative Light Units (RLUs) are read by the luminometer. Each plate contains a negative control, constituted by diploid yeast containing the bait of interest and an empty prey vector. To be scored as positive, the diploids tested have to have an RLU number at least twice as high as the negative control.

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Example 2

Minimum interaction domain mapping

Further analysis of yeast two hybrid (Y2H) interacting proteins includes the dissection of protein motifs responsible for the interaction. Sequence alignment of multiple clones identified in the Y2H screens can help identify the smallest common region responsible for the interaction. In the absence of appropriate clones, deletion mapping of interacting domains is necessary.

PCR primers containing restriction sites suitable for cloning are designed to cover multiple sub-domains of the protein of interest (bait or prey). The methods involved in cloning, sequencing, yeast transformation, mating, and scoring of interactions are readily performed by one of ordinary skill in the art of molecular biology and genetic engineering.

Materials and Methods

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Minimum interaction domain: primers were designed for PCR of the Dkk-1 clone isolated by screening a primary osteoblast cell strain (HOB03C5) library with pooled Zmax1/LRP5 ligand binding domain (LBD) baits: LBD1 (Leu969-Pro1376) and LBD4 (Arg1070-Pro1376). The primers, which are presented in 5' to 3' orientation, were as follows:

	SEQ ID NO	<u>Primer</u>	Sequence
25	155	Forward 1	TTTTTTGTCGACCAATTCCAACGCTATCAAG
	156	Forward 2	TTTTTTGTCGACCTGCGCTAGTCCCACCCGC
	157	Forward 3	TTTTTTGTCGACCGTGTCTTCTGATCAAAATC
	158	Forward 4	TTTTTTGTCGACCGGACAAGAAGGTTCTGTTTG
	159	Reverse 1	TTTTTTGCGGCCGCTTATTTGGTGTGATACATTTTTG
	160	Reverse 2	TTTTTTGCGGCCGCTTAGCAAGACAGACCTTCTCC
	161	Reverse 3	TTTTTTGCGGCCGCTTAGTGTCTCTGACAAGTGTG

PCR was performed using PfuTurbo® polymerase (Stratagene). The PCR products were gel purified, digested with *Sall/Not*l and ligated to pPC86 (Gibco/BRL) which had been linearized with *Sall/Not*l. Clones were recovered and sequenced to ascertain that the structure was as expected and that the Gal4 activation domain and Dkk-1 were in-frame. The ORF of Dkk-1 was Met1-His266, as in human Dkk-1 (GenBank Accession No. XM_005730).

The clones used were as follows: D5 (F1/R3: Asn34-His266), D4 (F1/R2: Asn34-Cys245), D3 (F1/R1: Asn34-Lys182), D9 (F2/R3: Cys97-His266), D12 (F3/R3, val139-His266), D14 (F4/R3: Gly183-His266), D8 (F2/R2: Cys97-Cys245), and D11 (F3/R2: Val139-Cys245). F1, F2, F3 and F4 refer respectively to Forward primers 1, 2, 3 and 4. R1, R2 and R3 refer respectively to reverse primers 1, 2 and 3.

These clones were transformed into yeast and mated with each of three yeast strains containing pDBleu (Gibco/BRL), pDBleuLBD1, and pDBleuLBD4. Positive interactions were detected by growth of the hybrids on appropriate selective media.

Results

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Minimum interaction domain: Figure 6 shows that while growth was observed in diploids of D4, D5, D8, D9, and D12, no growth was observed in hybrids of D3, D11, and D12. Carboxy terminal (C-terminal) deletions indicated that while the C-terminal amino acids of Dkk-1 containing the potential N-glycosylation site (Arg246-His266) are not required for interaction with Zmax1/LRP5 LBD baits, the Cys2 domain, Gly183-Cys245, is required. N-terminal deletions also demonstrated that the region between the two cysteine domains, *i.e.* Val139 to Lys182, is also required. Two minimum interaction domain constructs were isolated: D12 (Val139-His266) and D8 (Cys97-Cys245). Similar constructs could be prepared for Dkk-1 interactors.

Example 3

Yeast-2 Hybrid screen for peptide aptamer sequences to Dkk-1
Peptide aptamer library construction

A peptide aptamer library, Tpep, was constructed, which provides a means to identify chimeric proteins that bind to a protein target (or bait) of interest using classic yeast two hybrid (Y2H) assays. The Tpep library is a combinatorial aptamer library composed of constrained random peptides, expressed within the context of the disulfide loop of *E. coli* thioredoxin (trxA), and as C-termini fusion to the *S. cerevisiae* Gal4 activation domain. The Tpep library was generated using a restriction enzyme modified recombinant Y2H prey vector, pPC86 (Gibco), which contains the trxA scaffold protein.

Generation of aptamer-encoding sequences

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Aptamer-encoding sequences were produced as follows. DNA encoding random stretches of approximately sixteen amino acids surrounded by appropriate restriction sites were generated by semi-random oligonucleotide synthesis. The synthetic oligonucleotides were PCR-amplified, restriction digested, and cloned into the permissive sites within the trxA scaffold protein. The cloning strategy was to insert the random oligonucleotide sequence is in-frame with the scaffold protein coding sequence, resulting in expression of a scaffold protein-aptamer chimera. The scaffold protein is itself in-frame with the activation domain of Gal4, within the pPC86 vector that is appropriate for the aptamer to be expressed and functional in a regular Y2H assay. Additional methods of preparing aptamers would be apparent to the skilled artisan.

Generation of a permissive recombinant pPC86 vector containing the trxA coding sequence

First the *Rsr*II restriction site located within the Gal4 activation domain of pPC86 (Gibco) was eliminated by site-directed mutagenesis (Quickchange™ kit, Stratagene). The amino acid sequence of the Gal4 activation domain was unchanged by this modification. The strength of different control interactions was verified to be unchanged by the modification.

Second, the *E. coli* trxA coding sequence was cloned into the *Sal*I and *Not*I sites of the *Rsr*II-modified pPC86. *EcoR*I and *Spe*I sites were then introduced within the trxA

RsrII site. The oligonucleotides encoding the peptide aptamers were cloned into the EcoRI and Spel sites of the resulting vector.

Example 4

Yeast-2 Hybrid screen for Dkk-1 interacting proteins

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A Dkk-1 bait sequence was utilized in a yeast two hybrid screen to identify Dkk-1 interacting proteins. The procedure for the Y2H was carried out similarly to that employed in Example 1, except that the Dkk-1 bait from Figure 2C was used instead of LRP baits. The screen was performed using Hela and fetal brain libraries (Invitrogen Corporation, Carlsbad, CA). Multiple libraries were used to identify additional Dkk-1 interacting proteins and to confirm interactions found in other libraries.

The list of Dkk-1 interacting proteins uncovered in these Y2H screens are listed in Figure 5.

The interacting proteins identified in the Dkk-1 bait screen can be used in other Y2H screens with LRP baits and other Dkk-1 interacting proteins to determine more complex interactions which may modulate Dkk-1/LRP interactions and/or Wnt signaling.

Example 5

Generation of antibodies

In each of the following antibody-generating examples, the synthesis of these linear peptides is followed by injection into two New Zealand Rabbits. Subsequent boosts and bleeds are taken according to a standard ten-week protocol. The end-user receives back 5 mgs of peptide, aliquots of pre-bleeds, roughly 80 ml of crude sera from each of the two rabbits and, and ELISA titration data is obtained.

Generation of LRP5 Polymorphism-specific antibodies

Antibodies were generated to the following peptides to obtain antibodies which distinguish the HBM polymorphism versus wild-type LRP5/Zmax: MYWTDWVETPRIE

(SEQ ID NO:123) (mutant peptide) and MYWTDWGETPRIE (SEQ ID NO:124) (wild-type peptide for negative selection). Immunofluorescence data confirmed that the antibody, after affinity purification, is specific for HBM and does not recognize LRP5 (Figure 17).

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Generation of LRP5 Monospecific antibodies

LRP5 monospecific polyclonal antibodies were generated to the following amino acid sequences of LRP5: Peptide 1 (a.a. 265-277) - KRTGGKRKEILSA (SEQ ID NO:125), Peptide 2 (a.a. 1178-1194) - ERVEKTTGDKRTRIQGR (SEQ ID NO:126), and Peptide 3 (a.a. 1352-1375) - KQQCDSFPDCIDGSDE (SEQ ID NO:127). Immunofluorescence confirmed that the antibody generated detects LRP5.

Generation of Dkk-1 monospecific polyclonal antibodies

Dkk-1 monospecific polyclonal antibodies were generated to the following amino acid sequences of Dkk-1: Peptide 1 (a.a. 71-85) - GNKYQTIDNYQPYPC (SEQ ID NO:118), Peptide 2 (a.a. 165-186) - LDGYSRRTTLSSKMYHTKGQEG (SEQ ID NO:119), Peptide 3 (a.a. 246-266) - RIQKDHHQASNSSRLHTCQRH (SEQ ID NO:120), Peptide 4 (a.a. 147-161) - RGEIEETITESFGND (SEQ ID NO:121), and Peptide 5 (232-250) - EIFQRCYCGEGLSCRIQKD (SEQ ID NO:122) of human Dkk-1. Figure 26 shows the location of the various peptides selected, their relationship to the Dkk-1 amino acid sequence and polyclonal antibodies generated.

Western blots demonstrated that the antibodies generated against peptides 2 (Antibody #5521) (Figure 27) and 4 (Antibody #74397) (Figure 28) are specific toward Dkk-1. Figure 27 shows Western blots using 500 μ l of conditioned medium (CM) from non-transfected 293 cells or from 293 cells transfected with Dkk1-V5 that were immunoprecipitated by anti-V5 antibody. Bead elutes were separated by non-reducing SDS-PAGE (lanes #4, 5 of Figure 27). 20 μ l of conditioned medium from both samples (lanes #2, 3 of Figure 27) and from Dkk1-AP transfected 293 cells (lane #6 of Figure 27) were additionally separated on the gel. The Western was performed using

antibodies Anti-V5/AP (1:10,000) and Ab#5521 (10 μ g/ml). Ab#5521 detected Dkk1-V5 and Dkk1-AP from conditioned medium.

Figure 28 shows Western blot results using Ab#74397. Anti-V5/AP was tested at a 1:4000 dilution and Ab#74397 was tested at a 1:500 dilution. Ab#74397 was able to detect Dkk1-V5 in both conditioned medium and immunoprecipitated conditioned medium.

The results obtained with antibodies #5521 and #74397 are summarized in the following table:

Rabbit No.	Peptide Position	Peptide Sequence	Purified (Y/N)	Western	Immuno- precipitation	Location
5521	165-186	LDGYSR RTTLSSK MYHTKG QEG	Y (Protein G purified)	Y	N/A	Between Cy1 and Cys2 domain
74397	147-161	RGEIEETI TESFGN D	N	Y	N/A	Between Cy1 and Cys2 domain

15 <u>Example 6</u>

Effects of exogenous Dkk-1 on Wnt-mediated signaling in the Xenopus embryo assay

Xenopus embryos are an informative and well-established *in vivo* assay system to evaluate the modulation of Wnt signaling (McMahon *et al.*, *Cell* 58: 1075-84 (1989); Smith and Harland, 1991; reviewed in Wodarz and Nusse 1998).

Modification of the Wnt signaling pathway can be visualized by examining the embryos for a dorsalization phenotype (duplicated body axis) after RNA injection into the ventral blastomere at the 4- or 8-cell stage. On the molecular level, phenotypes can be analyzed by looking for expression of various marker genes in stage 10.5 embryos. Such markers would include general endoderm, mesoderm, and ectoderm markers as well as a variety of tissue-specific transcripts.

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Analysis can be done by RT-PCR/TaqMan® and can be done on whole embryo tissue or in a more restricted fashion (microdissection). Because this system is very flexible and rapid, by injecting combinations of transcripts, such as HBM and different Wnts or Wnt antagonists, the mechanism of HBM in the Wnt pathwaycan thereby be dissected. Furthermore, investigations are conducted to determine whether Zmax/LRP5 and HBM differentially modulate Wnt signaling either alone, or in combination with other components. Previous studies have demonstrated that LRP6 alone or LRP5 + Wnt5a were able to induce axis duplication (dorsalization) in this system (Tamai et al., Nature 407: 530-35 (2000)).

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Constructs for Xenopus Expression (Vector pCS2*)

Constructs were prepared using the vector pCS2*. DNA inserts were subcloned in the sense orientation with respect to the vector SP6 promoter. The pCS2* vector contains an SV40 virus polyadenylation signal and T3 promoter sequence (for generation of antisense mRNA) downstream of the insert.

Full length Zmax/LRP5 and HBM ORF cDNA: Insert cDNA was isolated from the full length cDNA retrovirus constructs (with optimized Kozak sequences) by *Bg/II-EcoRI* digestion and subcloned into the *BamHI-EcoRI* sites of the pCS2⁺ vector.

Full length XWnt8: This cDNA was PCR amplified from a Xenopus embryo cDNA library using oligos 114484 (SEQ ID NO:162) (5'-CAGTGAATTCACCATGCAAAACACCACTTTGTTC-3') and 114487 (SEQ ID NO:163) (5'-CAGTTGCGGCCGCTCATCTCCGGTGGCCTCTG-3'). The oligos were designed to amplify the ORF with a consensus Kozak sequence at the 5' end as determined from GenBank #X57234. PCR was carried out using the following conditions: 96°C, 45 sec.; 63°C, 45 sec.; 72°C, 2 min. for 30 cycles. The resulting PCR product was purified, subcloned into pCRII-TOPO (Invitrogen Corp.), sequence verified, and digested with BamHI/Xhol. This insert was subcloned into the vector at the BamHI-Xhol sites.

<u>Full length Wnt5a:</u> A murine Wnt5a cDNA clone was purchased from Upstate Biotechnology (Lake Placid, NY) and subcloned into the *EcoRI* site of the vector. Sequencing confirmed insert orientation.

Full length human Dkk-1: A human cDNA with GenBank accession number AF127563 was available in the public database. Using this sequence, PCR primers were designed to amplify the open reading frame with a consensus Kozak sequence immediately upstream of the initiating ATG. Oligos 117162 (SEQ ID NO:164) (5'-CAATAGTCGACGAATTCACCATGGCTCTGGGCGCAGCGG-3') and 117163 (SEQ ID NO:165) (5'-GTATTGCGGCCGCTCTAGATTAGTGTCTCTGACAAGTGTGAA-3') were used to screen a human uterus cDNA library by PCR. The resulting PCR product was purified, subcloned into pCRII-TOPO (Invitrogen Corp.), sequence verified, and digested with *EcoRI/XhoI*. This insert was subcloned into the pCS2⁺ vector at the *EcoRI-XhoI* sites.

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Full length human Dkk-2: A full length cDNA encoding human Dkk-2 was isolated to investigate the specificity of the Zmax/LRP5/HBM interaction with the Dkk family of molecules. Dkk-1 was identified in yeast as a potential binding partner of Zmax/LRP5/HBM. Dkk-1 has also been shown in the literature to be an antagonist of the Wnt signaling pathway, while Dkk-2 is not (Krupnik et al., 1999). The Dkk-2 full length cDNA serves as a tool to discriminate the specificity and biological significance of Zmax/LRP5/HBM interactions with the Dkk family (e.g., Dkk-1, Dkk-2, Dkk-3, Dkk-4, Soggy, their homologs and variant, etc.). A human cDNA sequence for Dkk-2 (GenBank Accession No. NM 014421) was available in the public database. Using this sequence, PCR primers were designed to amplify the open reading frame with a consensus Kozak sequence immediately upstream of the initiating ATG. Oligos 51409 (SEQ ID NO:166) (5'- CTAACGGATCCACCATGGCCGCGTTGATGCGG-3') and 51411 (SEQ ID NO:167) (5'-GATTCGAATTCTCAAATTTTCTGACACACATGG-3') were used to screen human embryo and brain cDNA libraries by PCR. The resulting PCR product was purified, subcloned into pCRII-TOPO, sequence verified, and digested with BamHI/EcoRI. This insert was subcloned into the pCS2+vector at the BamHI-EcoRI sites.

Full length LRP6 was isolated from the pED6dpc4 vector by *Xhol-Xbal* digestion. The full length cDNA was reassembled into the *Xhol-Xbal* sites of pCS2⁺. Insert orientation was confirmed by DNA sequencing.

mRNA Synthesis and Microinjection Protocol

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mRNA for microinjection into *Xenopus* embryos is generated by *in vitro* transcription using the cDNA constructs in the pCS2⁺ vector described above as template. RNA is synthesized using the Ambion mMessage mMachine high yield capped RNA transcription kit (Cat. #1340) following the manufacturer's specifications for the Sp6 polymerase reactions. RNA products were brought up to a final volume of 50 µl in sterile, glass-distilled water and purified over Quick Spin Columns for Radiolabelled RNA Purification G50-Sephadex (Roche, Cat. #1274015) following the manufacturer's specifications. The resulting eluate was finally extracted with phenol:chloroform:isoamyl alcohol and isopropanol precipitated using standard protocols (Sambrook et al., 1989). Final RNA volumes were approximately 50 µl. RNA concentration was determined by absorbance values at 260 nm and 280 nm. RNA integrity was visualized by ethidium bromide staining of denaturing (formaldehyde) agarose gel electrophoresis (Sambrook et al., 1989). Various amounts of RNA (2 pg to 1 ng) are injected into the ventral blastomere of the 4- or 8-cell Xenopus embryo. These protocols are described in Moon et al., Technique-J. of Methods in Cell and Mol. Biol. 1: 76-89 (1989), and Peng, Meth. Cell. Biol. 36: 657-62 (1991).

Screening for Duplicated Body Axis

In vitro transcribed RNA is purified and injected into a ventral blasomere of the 4-or 8-cell *Xenopus* embryo (approx. 2 hours post-fertilization). At stage 10.5 (approx. 11 hours post-fertilization), the injected embryos are cultured for a total of 72 hours and then screened for the presence of a duplicated body axis (dorsalization) (Figure 7). Using XWnt8-injected (2-10 pg) as a positive control (Christian et al. (1991)) and water-injected or non-injected embryos as negative controls, we replicated the published observation that Zmax(LRP5) + Wnt5a (500 and 20 pg, respectively) could induce axis duplication. Wnt5a (20 pg) alone could not induce axis duplication (as previously reported by Moon *et al.* (1993)). We have also injected GFP RNA (100-770 pg) as a negative control to show that the amount of RNA injected is not perturbing embryo development (not shown). Strikingly, HBM + Wnt5a (500 and 20 pg, respectively)

yielded an approximately 3.5 fold more robust response of the phenotype (p=0.043 by Fisher's exact test) compared to Zmax(LRP5) + Wnt5a, suggesting that the HBM mutation is activating the Wnt pathway (Figures 8 and 9). The HBM/Wnt5a embryos also appear to be more "anteriorized" than the Zmax(LRP5)/Wnt5a embryos, again suggestive of a gain-of-function mutation.

The role of Dkk-1 as a modulator of Zmax/LRP5- and HBM-mediated Wnt signaling was investigated. Literature reports have previously characterized *Xenopus* and murine Dkk-1 as antagonists of the canonical Wnt pathway in the *Xenopus* system (Glinka *et al.*, *Nature* 391:357-362 (1998)). Using the human Dkk-1 construct, a doseresponse assay was performed to confirm that our construct was functional and to identify the optimal amount of RNA for microinjection. Using 250 pg/embryo of hDkk-1 RNA, over 90% (p<0.001) of the embryos were observed to display enlarged anterior structures (big heads) as anticipated from the published reports (Figure 10).

The mechanism of hDkk-1 modulation of Wnt signaling in the presence of Zmax/LRP5 or HBM was also investigated. Without any hDkk-1 present, it was confirmed that HBM + Wnt5a was a more potent activator of Wnt signaling than Zmax/LRP5 + Wnt5a (p<0.05). Interestingly, in the presence of hDkk-1 (250 pg), Zmax/LRP5-mediated Wnt signaling was repressed (p<0.05) but hDkk-1 was unable to repress HBM-mediated Wnt signaling (p<0.01) (Figure 11). The specificity of this observation can be further addressed by investigating other members of the Dkk family, other Wnt genes, LRP6, additional Zmax/LRP5 mutants, and the peptide aptamers.

Example 7

Effects of exogenous Dkk and LRP5 on Wnt signaling in the TCF-luciferase Assay

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Wnt activity can be antagonized by many proteins including secreted Frizzled related proteins (SFRPs), Cerberus, Wnt Inhibitory Factor-1 and Dkk-1 (Krupnik *et al.*, 1999). The Dkk family of proteins consists of Dkk-1-4 and Soggy, a Dkk-3-like protein. Dkk-1 and Dkk-4 have been shown to antagonize Wnt mediated *Xenopus* embryo development, whereas Dkk-2, Dkk-3, and Soggy do not. Unlike many of these proteins

that antagonize Wnt activity by directly interacting with Wnt proteins, Dkk-1 acts by binding to two recently identified Wnt coreceptors, LRP5 and LRP6. (Mao *et al.*, 2001; Bafico *et al.*, 2001). The details of this interaction have been examined by the present inventors and Mao et al. using deletion constructs of LRP6, which demonstrated that EGF repeats 3 and 4 are important for Dkk-1 interaction. Accordingly, the activity of two Dkk proteins, Dkk-1 and Dkk-2, were investigated with various Wnt members, LRP5, LRP6, and the mutant form of LRP5, designated HBM. The present invention explores whether there is any functional difference between LRP5 and HBM with regard to Dkk action on Wnt mediated signaling. Various reagents were developed, including Dkk-1 peptides, constrained LRP5 peptide aptamers, constrained Dkk-1 peptide aptamers and polyclonal antibodies to Dkk-1 (in Example 5 above) to identify factors that mimic HBM mediated Wnt signaling.

Methods

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Various LRP5 constrained peptides were developed. Specifically, four peptides that interact with the LBD of LRP5 (Figure 4,constructs OST259-262 in Figure 12) and three peptides that interact with the cytoplasmic domain of LRP5 (constructs OST266-OST268 in Figure 12). In addition two Dkk-1 peptides were developed: constructs OST264 and OST265 in Figure 12, corresponding to Dkk-1 amino acids 139-266 and 96-245, containing the smallest region of Dkk-1 that interacts with LRP5 (Figure 6). The cDNA clones encoding the LRP5 LBD interacting peptides and the Dkk-1 peptides were subcloned into pcDNA3.1 with the addition of a Kozak and signal sequence to target the peptide for secretion. The constructs encoding the three peptides interacting with the cytoplasmic domain of LRP5 were also subcloned into pcDNA3.1. However, these latter constructs do not contain a signal sequence.

HOB-03-CE6 osteoblastic cells developed by Wyeth Ayerst (Philadelphia, PA) were seeded into 24-well plates at 150,000 cells per well in 1 ml of the growth media (D-MEM/F12 phenol red-free) containing 10% (v/v) heat-inactivated FBS, 1X penicillin streptomycin, and 1X Glutamax-1, and incubated overnight at 34°C. The following day, the cells were transfected using Lipofectamine 2000® (as described by the

manufacturer, Invitrogen) in OptiMEM (Invitrogen) with 0.35 μ g /well of LRP5, HBM, or control plasmid DNA (empty vector pcDNA3.1) and either Wnt1 or Wnt3a plasmid DNA. Similar experiments were performed with LRP6 plasmid DNA (0.35 μ g/well) or a control pEDdpc4 empty vector. Furthermore, each of these groups were then divided into three groups, those receiving 0.35 μ g/well Dkk-1, Dkk-2, or pcDNA3.1 control DNA. All wells were transfected with 0.025 μ g/well of CMV beta-galactosidase plasmid DNA and 0.35 μ g/well 16X TCF(AS)-luciferase reporter DNA (developed by Ramesh Bhat, Wyeth-Ayerst (Philadelphia, PA)). After 4 hours of incubation, the cells were rinsed and 1 ml of fresh growth media was added to each well. The cells were cultured overnight at 34°C, followed by a wash and a change of media. Cells were cultured for an additional 18-24 hours at 37°C. Cells were then lysed with 50 μ l/well of 1X lysis buffer. The extracts were assayed for beta-galactosidase activity (Galacto Reaction Buffer Diluent & Light Emission Accelerator, Tropix) using 5 μ l extract + 50 μ l beta-galactosidase diluent and luciferase activity (Luciferase Assay Reagent, Promega) using 20 μ l extract.

U2OS human osteosarcoma cells were also utilized. U2OS cells (ATCC) were seeded into 96-well plates at 30,000 cells per well in 200ul of the growth media (McCoy's 5A) containing 10% (v/v) heat-inactivated FBS, 1X penicillin streptomycin, and 1X Glutamax-1, and incubated overnight at 37°C. The following day, the nmedia was replaced with OptiMEM (Invitroge) and cells were transfected using Lipofectamine 2000® (as described by the manufacturer, Invitrogen) with 0.005μg/well of LRP5, HBM, LRP6 or contol plasmid DNA (empty vector pcDNA3.1) and either Wnt1 (.0025ug/well) or Wnt3a (.0025ug/well) plasmid DNA. In addition, the 16x-(AS) TCF-TK-firefly-luciferase (Ramesh Bhat, WHRI, Wyeth) and control TK-renilla luciferase (Promega Corp.) were co-transfected at 0.3ug/well and 0.06ug/well respectively in all experiments. Futhermore, each of these groups was then divided into different groups, those receiving 0.05ug/well Dkk-1, Dkk-2, Dkk3, Dkk1-Alkaline Phosphatase (AP), mutant Dkk-1 (C220A), Soggy or pcDNA3.1 control DNA. In other experiments, cells were co-transfected with 0.005 μg/well of LRP5, 0.0025ug/well of Wnt1 or Wnt3a (using 0.0025 μg/well of a control pcDNA3.1) with LRP5-interacting aptamers (0.05ug/well).

Cells were cultured for an additional 18-20 hours at 37°C. Culture medium was removed. Cells were cultured for an additional 18-20 hours at 37°C. Culture medium was removed. Cells were then lysed with 100 μ l/well of 1X Passive Lysis Buffer (PLB) of Dual Luciferase Reagent kit (DLR-kit-Promega Corp.) 20 μ l of the lysates were combined with LARII reagent of DLR-kit and assayed for TCF-firefly luciferase signal in Top Count (Packard) instrument. After measuring the Firefly readings, 100 μ l of the "Stop and Glo" reagent of DLR kit that contains a quencher and a substrate for renilla luciferase was added into each well. Immediately the renilla luciferase reading was measured using the Top Count (Packard) Instrument. The ratios of the TCF-firefly luciferase to control renilla readings were calculated for each well and the mean ratio of triplicate or more wells was expressed in all data.

Results

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The results of these experiments demonstrate that Dkk-1, in the presence of Wnt1 and LRP5, significantly antagonized TCF-luciferase activity (Figure 14). In marked contrast, Dkk-1 had no effect on HBM/Wnt1 mediated TCF-luciferase activity (Figure 14). In similar experiments, Dkk-1 was also able to antagonize LRP5/Wnt3a but not HBM/Wnt3a mediated TCF-luciferase activity (Figure 15). These results indicate that the HBM mutation renders Dkk-1 inactive as an antagonist of Wnt1 and Wnt3a signaling in HOB03CE6 osteoblastic cells. In other experiments with Wnt1, Dkk-1 had no effect on LRP5 or HBM mediated TCF-luciferase activity (Figure 14). In contrast, with either LRP5 or HBM in the presence of Wnt3a, Dkk-2 was able to antagonize the TCF-luciferase activity (Figure 15). These latter results indicate that the HBM mutation has no effect on Dkk-2 action in the presence of Wnt3a. Experiments were also performed using the closely related LRP6 cDNA in HOB-03-CE6 cells. In these experiments, LRP6/Wnt1 and LRP6/Wnt3a mediated TCF-luciferase were regulated in the same manner as LRP5. Specifically, Dkk-1 antagonized LRP6/Wnt1 mediated TCF-luciferase activity, whereas Dkk-2 had no effect (Figure 14). However, similar to the action of Dkk-2 with LRP5/Wnt3a, Dkk-2 was able to antagonize LRP6/Wnt3a mediated TCF-luciferase activity (Figure 15).

The results in the U2OS cells show a robust effect of the OST262 LRP5 peptide aptamer activation of Wnt signaling in the presence of Wnt3a (Figure 16). These functional results are confirmed by the results shown below in Example 11 using LRP5 peptide aptamers in the Xenopus assay. Such results affirmatively demonstrate that the effects of small molecules on LRP5/LRP6/HBM signaling can be detected using the TCF-luciferase assay.

These data demonstrate that there is a functional difference between LRP5 and HBM regarding the ability of Dkk-1 to antagonize Wnt1 and Wnt3a signaling. These data and previous data showing that Dkk-1 directly interacts with LRP5 suggests that the inability of Dkk-1 to antagonize HBM/Wnt signaling may in part contribute to the HBM phenotype. These experiments further demonstrate the ability to test various molecules (e.g., small molecules, aptamers, peptides, antibodies, LRP5 interacting proteins or Dkk-1 interacting proteins, and the like) for a LRP5 ligand that mimics HBM mediated Wnt signaling or factors that block Dkk-1 interaction with LRP5.

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Example 8

Yeast-2 Hybrid Interaction Trap

Small molecule inhibitors (or partial inhibitors) of the Dkk-LRP interaction may be an excellent osteogenic therapeutic. One way to investigate this important protein-protein interaction is using Y2H techniques substantially as described above and as is well known in the art. Regions of LRP5, such as LRP5 LBD, have been found to functionally interact with Dkk. This interaction is quantitated using a reporter element known in the art, e.g., LacZ or luciferase, which is only activated when bait and prey interact. The Y2H assay is used to screen for compounds which modulate the LRP-Dkk interaction. Such a modulation would be visualized by a reduction in reporter element activation signifying a weaker or disrupted interaction, or by an enhancement of the reporter element activation signifying a stronger interaction. Thus, the Y2H assay can be used as a high-throughput screening technique to identify compounds which disrupt or enhance Dkk interaction with LRP5/LRP6/HBM, which may serve as potential therapeutics.

For example, the Interaction Trap methodology can be used as follows. The LRP5 LBD, for example, was fused with LexA and Dkk-1 was fused with either Gal4-AD or B42. With the LRP5LBD-LexA bait and the Gal4AD-Dkk prey, over a 20-fold activation of a lacZ reporter (under the control of a single LexA operator) was detected over the background. Using a Dkk-1 mutant (C220A) that is unable to bind to LRP, the interaction was reduced in yeast, showing the specificity of this interaction and system (Figure 18). As a result, small molecules may be identified that modulate this interaction between LRP and Dkk.

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Example 9

Cell-Based Functional High-Throughput Assay

To develop a high throughput assay, the TCF-luciferase assay described in Example 7 was modified utilizing low level expression of endogenous LRP5/6 in U2OS and HEK293 cells. However, HOB-03-CE6 cells and any other cells which show a differential response to Dkk depending on whether LRP5, LRP6 or HBM are expressed. Using U2OS (human osteosarcoma) and HEK293 (ATCC) cells, the TCF-luciferase and tk-Renilla reporter element constructs were co-transfected along with Wnt3a/1 and Dkk. Wnt3a alone, by using endogenous LRP5/6, was able to stimulate TCF reporter gene activation. When Dkk, is co-transfected with Wnt3a/Wnt 1 and reporters (TCF-luci and tk-Renilla), Dkk represses reporter element activity. In addition, the TCF-luci signal is activated by Wnt3a/Wnt1 can be repressed by the addition of Dkk-enriched conditioned media to the cells containing Wnt3a/Wnt1 and reporters. The assay is further validated by the lack of TCF-reporter inhibition by a point mutant construct (C220A) of Dkk1.

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The Dkk-mediated repression of the reporter is dependent upon the concentration of transfected Dkk cDNA or on the amount of Dkk-conditioned media added. In addition, the Dkk-mediated reporter suppression can be altered by the cotransfection of LRP5, LRP6, and HBM cDNAs in the U2OS or HEK293 cells. In general, U2OS cells show greater sensitivity to Dkk-mediated reporter suppression than that in HEK-293 cells. In U2OS cells, the transfection of LRP5/LRP6/HBM cDNA leads

to moderate activation of TCF-luci in the absence of Wnt3a/Wnt1 transfection. This activation presumably utilizes the endogenous Wnts present in U2OS cells. Under this condition, Dkk1 can repress TCF-luci and shows a differential signal between LRP5 and HBM. By co-transfecting Wnt3a/Wnt1, there is a generalized increase in the TCF-luci signal in the assay. Further, one can detect Dkk-mediated differential repression of the reporter due to LRP5 and HBM cDNA expression as well as between LRP5 and LRP6 cDNA. The repression is maximal with LRP6, moderate with LRP5, and least with HBM cDNA expression. In addition, the assay can detect the functional impact of the LRP5 interacting peptide aptamers (Figure 4), Dkk1 interacting aptamers and binding domains of Dkk-1 (Figure 6; OST264 and OST265 of Figures 12 and 13).

Using this system with a suppressed Wnt-TCF signal due to the presence of both Dkk and Wnt3a, one can screen for compounds that could alter Dkk modulation of Wnt signaling, by looking for compounds that activate or the TCF-luciferase reporter, and thereby relieve the Dkk-mediated repression of the Wnt pathway. Such compounds identified may potentially serve as HBM-mimetics and be useful, for example, as osteogenic therapeutics. Data generated from this high throughput screen are demonstrated in Figures 19-21. Figure 19 shows that Dkk1 represses Wnt3a-mediated signaling in U2OS bone cells. Figure 20 demonstrates the functional differences between LRP5, LRP6, and HBM. Dkk-1 represses LRP6 and LRP5 but has little or no effect on HBM-generated Wnt1 signaling in U2OS cells. Figure 21 demonstrates the differential effects of various Dkk family members and modified Dkks, including Dkk-1, a mutated Dkk-1 (C220A), Dkk-1-AP (modified with alkaline phosphatase), Dkk-3, and Soggy.

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Example 10

DKK/LRP5/6/HBM ELISA Assay

A further method to investigate Dkk binding to LRP is via ELISA assay. Two possible permutations of this assay are exemplified. LRP5 is immobilized to a solid surface, such as a tissue culture plate well. One skilled in the art will recognize that other supports such as a nylon or nitrocellulose membrane, a silicon chip, a glass slide,

beads, etc. can be utilized. In this example, the form of LRP5 used is actually a fusion protein where the extracellular domain of LRP5 is fused to the Fc portion of human IgG. The LRP5-Fc fusion protein is produced in CHO cell extracts from stable cell lines. The LRP5-Fc fusion protein is immobilized on the solid surface via anti-human Fc antibody or by Protein-A or Protein G-coated plates, for example. The plate is then washed to remove any non-bound protein. Conditioned media containing secreted Dkk protein or secreted Dkk-epitope tagged protein (or purified Dkk or purified Dkk-epitope tagged protein) is incubated in the wells and binding of Dkk to LRP is investigated using antibodies to either Dkk or to an epitope tag. Dkk-V5 epitope tagged protein would be detected using an alkaline phosphatase tagged anti-V5 antibody.

Alternatively, the Dkk protein could be directly fused to a detection marker, such as alkaline phosphatase. Here the detection of the Dkk-LRP interaction can be directly investigated without subsequent antibody-based experiments. The bound Dkk is detected in an alkaline phosphatase assay. If the Dkk-alkaline phosphatase fusion protein is bound to the immobilized LRP5, alkaline phosphatase activity would be detected in a colorimetric readout. As a result, one can assay the ability of small molecule compounds to alter the binding of Dkk to LRP using this system.

Compounds, when added with Dkk (or epitope-tagged Dkk) to each well of the plate, can be scored for their ability to modulate the interaction between Dkk and LRP based on the signal intensity of bound Dkk present in the well after a suitable incubation time and washing. The assay can be calibrated by doing cold competition experiments with unlabeled Dkk or with a second type of epitope-tagged Dkk. Any small molecule that is able to modulate the Dkk-LRP interaction may be a suitable therapeutic candidate, more preferably an osteogenic therapeutic candidate.

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Example 11

Functional Evaluation of Peptide Aptamers in Xenopus

The constrained peptide aptamers constructs OST258-263 (where 258 contains the signal sequence by itself and 263 contains an irrelevant constrained peptide)

(Figures 12 and 13) were used to generate RNA substantially as described in Example

7, except the vector was linearized by restriction endonuclease digestion and RNA was generated using T7 RNA polymerase.

Aptamer RNA was injected at 250 pg per blastomere using the protocol of Example 7. Wnt signaling was activated, as visualized by embryo dorsalization (duplicated body axis) with aptamers 261 and, more strongly, 262. The results of this assay are shown in Figures 22 and 23. These results suggest that aptamers 261 and 262 are able to activate Wnt signaling possibly by binding to the LBD of LRP, thereby preventing the modulation of LRP-mediated signaling by Dkk.

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The aptamers of the present invention can serve as HBM-mimetics. In the Xenopus system they are able to induce Wnt signaling all by themselves. They may also serve as tools for rational drug design by enhancing the understanding of how peptides are able to interact with LRP and modulate Wnt signaling at the specific amino acid level. Thus, one would be able to design small molecules to mimic their effects as therapeutics. In addition, the aptamers identified as positives in this assay may be used as therapeutic molecules themselves.

Example 12

Homogenous Assay

An excellent method to investigate perturbations in protein-protein interactions is via Fluorescence Resonance Energy Transfer (FRET). FRET is a quantum mechanical process where a fluorescent molecule, the donor, transfers energy to an acceptor chromophore molecule which is in close proximity. This system has been successfully used in the literature to characterize the intermolecular interactions between LRP5 and Axin (Mao et al., *Molec. Cell Biol.* 7:801-809). There are many different fluorescent tags available for such studies and there are several ways to fluorescently tag the proteins of interest. For example, CFP (cyan fluorescent protein) and YFP (yellow fluorescent protein) can be used as donor and acceptor, respecively. Fusion proteins, with a donor and an acceptor, can be engineered, expressed, and purified.

For instance, purified LRP protein, or portions or domains thereof, fused to CFP and purified Dkk protein, or portions or domains thereof that interact with Dkk or LRP

respectively, fused to YFP can be generated and purified using standard approaches. If LRP-CFP and Dkk-YFP are in close proximity, the transfer of energy from CFP to YFP will result in a reduction of CFP emission and an increase in YFP emission. Energy is supplied with an excitation wavelength of 450 nm and the energy transfer is recorded at emission wavelengths of 480 nm and 570 nm. The ratio of YFP emission to CFP emission provides a guage for changes in the interaction between LRP and Dkk. This system is amenable for screening small molecule compounds that may alter the Dkk-LRP protein-protein interaction. Compounds that disrupt the interaction would be identified by a decrease in the ratio of YFP emission to CFP emission. Such compounds that modulate the LRP-Dkk interaction would then be considered candidate HBM mimetic molecules. Further characterization of the compounds can be done using the TCF-luciferase or Xenopus embryo assays to elucidate the effects of the compounds on Wnt signaling.

While the above example describes a cell-fee, solution-phase assay using purified components, a similar cell-based assay could also be performed. For example, LRP-CFP fusion protein can be expressed in cells. The Dkk-YFP fusion protein then could be added to the cells either as purified protein or as conditioned media. The interaction of LRP and Dkk is then monitored as described above.

All references cited herein are hereby incorporated by reference in their entirety for all purposes. The following applications are also incorporated by reference in their entirety herein for all purposes: U.S. Application No. 60/290,071, filed May 11, 2001; U.S. Application No. 09/544,398, filed on April 5, 2000; U.S. Application No. 09/543,771, filed April 5, 2000; 09/578,900; U.S. Application No. 09/229,319, filed January 13, 1999; U.S. Provisional Application 60/071,449, filed January 13, 1998; and International Application PCT/US00/16951, filed June 21, 2000; International PCT Application entitled "HBM Variants That Modulate Bone Mass and Lipid Levels," filed May 13, 2002; and International PCT Application entitled "Transgenic Animal Model of Bone Mass Modulation," filed May 13, 2002. Additionally, this application claims priority to U.S. provisional applications 60/291,311, filed May 17, 2001; 60/353,058, filed

February 1, 2002; and 60/361,293, filed March 4, 2002; the texts of which are herein incorporated by reference in their entirety for all purposes.

CLAIMS

We claim:

A method of regulating LRP5, LRP6, or HBM activity in a subject
 comprising administering a composition which modulates a Dkk activity in an amount effective to regulate LRP5, LRP6, or HBM activity.

2. The method of any of Claims 1, 24, 28, 33, 36, 37, 48, 64, 65, 93, 98, 101, 105, 107, 111, or 112, wherein the Dkk is Dkk-1.

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- 3. The method of any of Claims 1, 24, 28, or 33, wherein the Dkk is Dkk-1 and the Dkk activity is inhibited.
- 4. The method of Claims 1 or 24, wherein the Dkk activity modulates bone mass and/or lipid levels.
 - 5. The method of Claim 4, wherein bone mass is increased and/or lipid levels are decreased.

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6. The method of Claim 5, wherein the increase in bone mass is determined via one or more of a decrease in fracture rate, an increase in bone strength, an increase in bone density, an increase in bone mineral density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density, an increase in bone diameter, and an increase in inorganic bone content.

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7. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises one or more compounds selected from the group consisting of Dkk interacting proteins, or a Dkk-binding fragment thereof.

	8.	The method of any of Claims 1, 24, 28, or 33, wherein said			
com	position	comprises an antisense, a siRNA, or shRNA molecule which			
recognizes and binds to a nucleic acid encoding one or more Dkk interacting					
prote	eins.				

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9. The method of any of Claims 1, 24, 28, or 33, and wherein said composition comprises a Dkk peptide aptamer.

10. The method of any of Claims 1, 24, 28, or 33, wherein said 10 composition comprises a mimetic of a Dkk peptide aptamer.

> 11. The method of any of Claims 1, 24, 28, or 33, wherein said composition inhibits Dkk binding to LRP5, LRP6, or HBM.

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- 12. The method of any of Claims 1, 24, 28, or 33, wherein said composition enhances binding of Dkk to LRP5, LRP6, or HBM.
- 13. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises a Dkk interacting protein peptide aptamer.

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The method of any of Claims 1, 24, 28, or 33, wherein said 14. composition comprises a mimetic of a Dkk interacting protein peptide aptamer.

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15. The method of any of Claims 1, 24, 28 or 33, wherein said composition inhibits Dkk interacting protein or Dkk-binding fragment thereof binding to Dkk.

16. The method of any of Claims 1, 24, 28, or 33, wherein said composition enhances binding of Dkk interacting protein or Dkk-binding fragment thereof to Dkk.

17. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a vertebrate or an invertebrate organism.

- 18. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a mammal.
 - 19. The method of any of Claims 1, 24, 28, or 33, wherein said subject is a canine, a feline, an ovine, a primate, an equine, a porcine, a caprine, a camelid, an avian, a bovine, or a rodent.

20. The method of Claim 19, wherein said primate is a human.

21. The method of any of Claims 1, 24, 28, or 33, wherein said composition comprises an LRP5 peptide aptamer.

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- 22. The method of Claim 21, wherein said peptide aptamer is OST262 (SEQ ID NO:208).
- 23. The method of any of Claims 1, 24, 28 or 33, wherein the composition comprises an LRP5 antibody or an immunologically active fragment thereof.
 - 24. A method of regulating Dkk-Wnt pathway activity in a subject comprising administering a composition which modulates Dkk activity in an amount effective to regulate Dkk-Wnt pathway activity.
 - 25. The method of Claims 24, 101, or 107, wherein the Wnt is one or more of Wnt1-Wnt19.

26. The method of Claim 25, wherein the Wnt is Wnt1, Wnt3, Wnt3a, or Wnt10b.

27. The method of Claim 24 wherein said composition which modulates Dkk activity or modulates Dkk interaction with LRP5/LRP6/HBM is administered in an amount effective to modulate Wnt signaling.

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- 28. A method of modulating bone mass in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM in an amount effective to modulate bone mass in the subject.
 - 29. The method of Claim 28, wherein bone mass is increased.
- 30. The method of the previous claim, wherein the increase in bone mass is determined via one or more of a decrease in fracture rate, an increase in bone strength, an increase in bone density, an increase in bone mineral density, an increase in trabecular connectivity, an increase in trabecular density, an increase in cortical density, an increase in bone diameter, and an increase in inorganic bone content.
 - 31. The method of Claims 28 or 36, wherein said subject has a bone mass disorder selected from the group consisting of a bone development disorder, a bone fracture, age-related loss of bone, chrondrodystrophy, druginduced bone disorder, high bone turnover, hypercalcemia, hyperostosis, osteogenesis imperfecta, osteomalacia, osteomyelitis, osteoporosis, Paget's disease, osteoarthritis, and rickets.
 - 32. The method of Claim 28, wherein the composition which modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM is

administered in an amount effective to modulate the amount of trabecular and/or cortical tissue.

- 33. A method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates Dkk activity or Dkk interaction with LRP5, LRP6, or HBM in an amount effective to modulate lipid levels in the subject.
 - 34. The method of Claim 33, wherein lipid levels are decreased.

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35. The method of Claim 33 or 36, wherein the subject has a lipid-modulated disorder and wherein the lipid-modulated disorder is selected from the group consisting of a cardiac condition, atherosclerosis, familial lipoprotein lipase deficiency, familial apoprotein CII deficiency, familial type 3 hyperlipoproteinemia, familial hypercholesterolemia, familial hypertriglyceridemia, multiple lipoprotein-type hyperlipidemia, elevated lipid levels due to dialysis and/or diabetes, and elevated lipid levels of unknown etiology.

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- 36. A method of diagnosing low or high bone mass and/or high or low lipid levels in a subject comprising examining expression of Dkk, LRP5, LRP6, HBM, or and HBM-like variant in the subject and determining whether Dkk, LRP5, LRP6, HBM, or an HBM-like variant is over- or under-expressed to determine whether subject has (a) high or low bone mass and/or (b) has high or low lipid levels.
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- 37. A method of screening for a compound which modulates the interaction of Dkk with LRP5, LRP6, HBM, or a Dkk-binding fragment of LRP5, LRP6, or HBM comprising:

(a) exposing Dkk and a LRP5, LRP6, and/or HBM binding fragment thereof to a compound; and

(b) determining whether said compound modulates Dkk interaction with the LRP5/LRP6/HBM binding fragment.

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38. The method of Claim 37, wherein said modulation is determined by whether said compound binds to Dkk or the LRP5, LRP6, or HBM binding fragment thereof.

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- 39. The method of Claim 37, wherein Dkk or a LRP-binding fragment thereof is attached to a substrate.
- 40. The method of Claim 37, wherein said compound comprises one or more compounds selected from the group consisting of Dkk interacting proteins, or a Dkk-binding fragment thereof.

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41. The method of Claim 37 or 48, wherein said compound comprises a Dkk peptide aptamer.

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- 42. The method of Claim 37 or 48, wherein said compound comprises a mimetic of a Dkk peptide aptamer.
- 43. The method of Claim 37 or 48, wherein said compound comprises a Dkk interacting protein peptide aptamer.

- 44. The method of Claim 37 or 48, wherein the compound comprises an LRP5 peptide aptamer.
- 45. The method of Claim 44, wherein the peptide aptamer is OST262 (SEQ ID NO:208).

46. The method of Claim 37 or 48, wherein the compound comprises an LRP5 antibody.

- 47. The method of Claim 37 or 48, wherein said compound is a mimetic of a Dkk interacting protein peptide aptamer.
 - 48. A method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting protein comprising:
 - (a) exposing a Dkk interacting protein or a Dkk-binding fragment thereof to a compound; and
 - (b) determining whether said compound bound to a Dkk interacting protein or the Dkk-binding fragment thereof; and
 - (c) further determining whether said compound modulates the interaction of Dkk interacting protein and Dkk.
 - 49. The method of Claim 48, wherein the Dkk interacting protein or a Dkk-binding fragment thereof is attached to a substrate.
 - 50. A composition comprising a LRP5, LRP6, or HBM activity-modulating compound and a pharmaceutically acceptable carrier therefor.
 - 51. The composition of Claim 50, wherein said LRP5, LRP6, or HBM activity-modulating compound comprises a compound which binds to Dkk thereby modulating the interaction of Dkk with LRP5, LRP6, or HBM.
 - 52. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises one or more Dkk interacting proteins and Dkk-binding fragments thereof.

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53. The composition of Claim 50, wherein said LRP5, or LRP6, or HBM modulating compound is a monoclonal antibody or an immunologically active fragment thereof which binds to a Dkk interacting protein, or a Dkk-binding fragment thereof.

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- 54. The composition of Claim 53, wherein the monoclonal antibody is human, chimeric, humanized, primatized®, or bispecific.
- 55. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises an antisense, a siRNA, or shRNA molecule which recognizes and binds to a nucleic acid encoding one or more Dkk interacting proteins.
 - 56. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a Dkk peptide aptamer.
 - 57. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a mimetic of a Dkk peptide aptamer.
- 58. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a Dkk interacting protein peptide aptamer.
 - 59. The composition of Claim 50, wherein said LRP5, LRP6, or HBM modulating compound comprises a mimetic of a Dkk interacting protein peptide aptamer.
 - 60. The composition of Claim 50, wherein the compound comprises an LRP5 peptide aptamer.

61. The composition of Claim 60, wherein the peptide aptamer is OST262.

- 62. The composition of Claim 50, wherein the compound comprises an LRP5 antibody.
 - 63. A pharmaceutical composition comprising a compound which modulates Dkk activity and a pharmaceutically acceptable carrier therefor.
- 10 64. A method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:
 - (a) creating an LRP5, LRP6, or HBM fluorescent fusion protein using a first fluorescent tag; and
 - (b) creating a Dkk fusion protein comprising a second fluorescent tag;
 - (c) adding a test compound; and
 - (d) assessing changes in the ratio of fluorescent tag emissions using Fluorescence Resonance Energy Transfer (FRET) or Bioluminescence Resonance Energy Transfer (BRET) to determine whether the compound modulates Dkk and LRP5/LRP6/HBM interactions.

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- 65. A method of identifying binding partners for a Dkk protein comprising the steps of:
- (a) exposing the Dkk protein(s) or a LRP5/LRP6 binding fragment thereof to a potential binding partner; and

- (b) determining if the potential binding partner binds to a Dkk protein or the LRP5/LRP6 binding fragment thereof.
- 66. A nucleic acid encoding a Dkk interacting protein peptide aptamer comprising a nucleic acid encoding a scaffold protein in-frame with the activation

domain of Gal4 or LexA that is in-frame with a nucleic acid that encodes a Dkk interacting protein amino acid sequence.

67. A vector comprising the nucleic acid of Claim 66.

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- 68. The nucleic acid of Claim 66, wherein the scaffold protein is trxA.
- 69. A method of detecting a modulatory activity of a compound on the binding interaction of a first peptide and a second peptide of a peptide binding pair that bind through extracellular interaction in their natural environment, comprising:
 - (i) culturing at least one eukaryotic cell comprising:
 - a nucleotide sequence encoding a first heterologous fusion protein comprising the first peptide or a segment thereof joined to a transcriptional activation protein DNA binding domain;
 - a nucleotide sequence encoding a second heterologous fusion protein comprising the second peptide or a segment thereof joined to a transcriptional activation protein transcriptional activation domain;

wherein binding of the first peptide or segment thereof and the second peptide or segment thereof reconstitutes a transcriptional activation protein; and

- c) a reporter element activated under positive transcriptional control of the reconstituted transcriptional activation protein, wherein expression of the reporter element produces a selected phenotype;
- (ii) incubating the eukaryotic cell in the presence of a compound under conditions suitable to detect the selected phenotype; and

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(iii) detecting the ability of the compound to affect the binding interaction of the peptide binding pair by determining whether the compound affects the expression of the reporter element which produces the selected phenotype;

wherein (1) said first peptide is a Dkk peptide and the second peptide is a peptide selected from LRP5, HBM, LRP6 and the Dkk-binding portion of LRP5/LRP6/HBM or (2) said first peptide is a Dkk interacting protein or the Dkk-binding fragment thereof and said second peptide is a Dkk peptide.

70. The method of Claim 69, wherein the eukaryotic cell is a yeast cell.

71. The method of Claim 70, wherein the yeast cell is Saccharomyces.

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- 72. The method of Claim 71, wherein the *Saccharomyces* cell is *Saccharomyces cerevisiae*.
- 73. The method of Claim 69, wherein the Dkk is Dkk-1 and wherein the compound comprises one or more Dkk interacting proteins or a Dkk-binding fragment thereof.
 - 74. The method of Claim 73, wherein the compound is directly added to assay.

- 75. The method of Claim 73, wherein the compound is recombinantly expressed by said eukaryotic cell in addition to said first and second peptides.
- 76. The method of Claim 69, wherein the compound comprises a Dkk peptide aptamer.

77. The method of Claim 69, wherein the compound comprises a mimetic of a Dkk peptide aptamer.

- 78. The method of Claim 69, wherein the compound comprises a Dkk interacting protein peptide aptamer.
 - 79. The method of Claim 69, wherein the compound comprises a mimetic of a Dkk interacting protein peptide aptamer.
- 80. The method of Claim 69, wherein the eukaryotic cell further comprises at least one endogenous nucleotide sequence selected from the group consisting of a nucleotide sequence encoding the DNA binding domain of a transcriptional activation protein, a nucleotide sequence encoding the transcriptional activation domain of a transcriptional activation protein, and a nucleotide sequence encoding the reporter element, wherein at least one of the endogenous nucleotide sequences is inactivated by mutation or deletion.
 - 81. The method of Claim 69, wherein the peptide binding pair comprises a ligand and a receptor to which the ligand binds.
 - 82. The method of Claim 69, wherein the transcriptional activation protein is Gal4, Gcn4, Hap1, Adr1, Swi5, Ste12, Mcm1, Yap1, Ace1, Ppr1, Arg81, Lac9, Qa1F, VP16, or a mammalian nuclear receptor.
 - 83. The method of Claim 69, wherein at least one of the heterologous fusion proteins is expressed from an autonomously-replicating plasmid.
 - 84. The method of Claim 69, wherein the DNA binding domain is a heterologous DNA-binding domain of a transcriptional activation protein.

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85. The method of Claim 84, wherein the DNA binding protein is selected from the group consisting of a mammalian steroid receptor and bacterial LexA protein.

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- 86. The method of Claim 69, wherein the reporter element is selected from the group consisting of *lacZ*, a polynucleotide encoding luciferase, a polynucleotide encoding green fluorescent protein (GFP), and a polynucleotide encoding chloramphenicol acetyltransferase.

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- 87. The method of Claim 86, wherein the reporter element is LacZ.
- 88. The method of Claim 69, wherein the test sample comprises an LRP5 peptide aptamer.

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- 89. The method of Claim 88, wherein the peptide aptamer is OST262 (SEQ ID NO:208).
- 90. The method of Claim 69, wherein the test sample comprises an LRP5 antibody.

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91. A transgenic animal wherein Dkk-1 is knocked out in a tissuespecific fashion.

- 92. The transgenic animal of Claim 91, wherein the tissue specificity is bone tissue, cancer tissue, or liver tissue.
- 93. A method for identifying potential compounds which modulate Dkk activity comprising:
 - a) measuring the effect on binding of one or more Dkk interacting proteins, or a Dkk-binding fragment thereof, with Dkk or a

fragment thereof in the presence and absence of a compound; and

b) identifying as a potential Dkk modulatory compound a compound which modulates the binding between one or more Dkk interacting proteins or Dkk-binding fragment thereof and Dkk or fragment thereof.

94. A peptide aptamer of Figure 3 (SEQ ID NOs:171-188) or Figure 4 (SEQ ID NOs:189-192).

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- 95. An antibody or antibody fragment which recognizes and binds to one or more peptides of amino acid sequences GNKYQTIDNYQPYPC (SEQ ID NO:118), LDGYSRRTTLSSKMYHTKGQEG (SEQ ID NO:119), RIQKDHHQASNSSRLHTCQRH (SEQ ID NO:120), RGEIEETITESFGND (SEQ ID NO:121), EIFQRCYCGEGLSCRIQKD (SEQ ID NO:122), MYWTDWVETPRIE (SEQ ID NO:123), MYWTDWGETPRIE (SEQ ID NO:124), KRTGGKRKEILSA (SEQ ID NO:125), ERVEKTTGDKRTRIQGR (SEQ ID NO:126), KQQCDSFPDCIDGSDE (SEQ ID NO:127), or a Dkk-1 amino acid sequence selected from the group consisting Asn34-His266 (SEQ ID NO:110), Asn34-Cys245 (SEQ ID NO:111), Asn34-Lys182 (SEQ ID NO:112), Cys97-His266 (SEQ ID NO:113), Val139-His266 (SEQ ID NO:114), Gly183-His266 (SEQ ID NO:115), Cys97-Cys245 (SEQ ID NO:116), or Val139-Cys245 (SEQ ID NO:117).
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- 96. The antibody or antibody fragment of Claim 95, wherein the antibody is a monoclonal antibody.
- 97. The antibody or antibody fragment of Claim 95, wherein the antibody is a polyclonal antibody

98. A method of identifying Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway comprising:

- (a) injecting Dkk and potential Dkk interacting protein mRNA into a *Xenopus* blastomere; and
- (b) assessing axis duplication or analyzing marker gene expression;
 and
- (c) identifying compositions which elicit changes in axis duplication or marker gene expression as Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway.

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99. The method of Claim 98, wherein the mRNA of HBM, LRP5/6, any Wnt, Wnt antagonist, Wnt pathway modulator, or combination of these is coinjected into the *Xenopus* blastomere.

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100. The method of Claim 98, wherein the marker gene analyzed is Siamois, Xnr3, slug, Xbra, HNK-1, endodermin, Xlhbox8, BMP2, BMP4, XLRP6, EF-1, or ODC.

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- 101. A method for identifying Dkk interacting proteins which modulate the interaction of Dkk with the Wnt signaling pathway comprising:
- (a) transfecting cells with constructs containing Dkk and potential Dkk interacting proteins; and
- (b) assessing changes in expression of a reporter gene linked to a Wnt-responsive promoter; and

- (c) identifying as a Dkk interacting protein any protein which alters reporter gene expression compared with cells transfected with a Dkk construct alone.
- 102. The method of Claim 101, wherein the cells are HOB-03-CE6, HEK293, or U2OS cells.

103. The method of Claim 101, wherein the Wnt-responsive promoter is TCF or LEF.

- 104. The method of Claim 101, wherein the cells are co-transfected with CMV β-galactosidase.
 - 105. A method for identifying compounds which modulate Dkk and LRP5/LRP6/HBM interactions comprising:
 - (a) immobilizing LRP5/LRP6/HBM to a solid surface; and
 - (b) treating the solid surface with a secreted Dkk protein or a secreted epitope-tagged Dkk and a test compound; and
 - (c) determining whether the compound regulates binding between Dkk and LRP5/LRP6/HBM using antibodies to Dkk or the epitope tag or by directly measuring activity of an epitope tag.

106. The method of Claim 105, wherein the epitope tag is alkaline phosphatase, histidine, or a V5 tag.

- 107. A method for identifying compounds which modulate the interaction of Dkk with the Wnt signaling pathway comprising:
 - (a) transfecting cells with constructs containing Dkk and Wnt proteins;
- (b) assessing changes in expression of a reporter element linked to a Wnt- responsive promoter; and
- (c) identifying as a Dkk/Wnt interaction modulating compound any compound which alters reporter gene expression compared with cells transfected with a Dkk construct alone.
- 108. The method according to Claim 107, wherein Wnt3a and Wnt1 constructs are co-transfected into the cells.

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109. The method according to Claim 107, wherein the cells are U2-OS, HOB-03-CE6, or HEK293 cells.

- 110. The method according to Claim 107, wherein the reporter element used is TCF-luciferase, tk-Renilla, or a combination thereof.
 - 111. A method of testing compounds that modulate Dkk-mediated activity in a mammal comprising
 - (a) providing a group of transgenic animals having (1) a regulatable one or more Dkk genes, (2) a knock-out of Dkk genes, or (3) a knock-in of one or more Dkk genes;
 - (b) providing a second group of control animals respectively for the group of transgenic animals in step (a); and
 - (c) exposing the transgenic animal group and control animal group to a potential Dkk-modulating compound which modulates bone mass or lipid levels; and
 - (d) comparing the transgenic animals and the control group of animals and determining the effect of the compound on bone mass or lipid levels in the transgenic animals as compared to the control animals.
 - 112. A method of screening for compounds or compositions which modulate the interaction of Dkk and a Dkk interacting protein comprising:
 - (a) exposing a Dkk interacting proteins or a Dkkbinding fragment thereof to a compound; and
 - (b) determining whether said compound binds to a Dkk interacting proteins or the Dkk-binding fragment thereof.

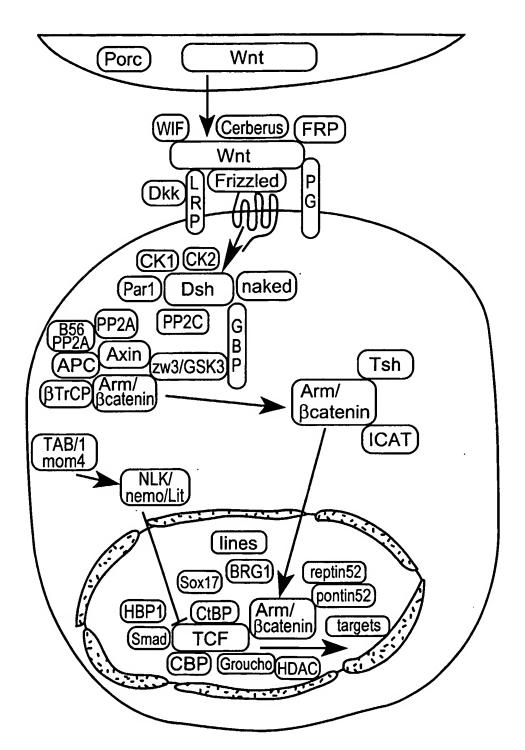
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113. The method of Claim 112, wherein said modulation is determined by whether said compound binds to the Dkk interacting protein or the Dkk-binding fragment thereof.

5 114. An antibody or antibody fragment which recognizes and binds to a sequence depicted in Figure 3 (SEQ ID NOs:171-188) or Figure 4 (SEQ ID NOs:189-192).



Model of Wnt signaling

FIG. 1

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Sequence of baits used in Y2H screens >DKK1 (SEQ ID N0: 168)

AATTCCAACGCTATCAAGAACCTGCCCCCACCGCTGGGCGCGCTG
CGGGGCACCCAGGCTCTGCAGTCAGCGCCGCGCGGGAATCCTGTA
CCCGGGCGGAATAAGTACCAGACCATTGACAACTACCAGCCGTAC
CCGTGCGCAGAGGACGAGGAGTGCGGCACTGATGAGTACTGCGCT
AGTCCCACCCGCGGAGGGGACGCGGGCGTGCAAATCTGTCTCGCCT
GCAGGAAGCGCCGAAAACGCTGCATGCGTCACGCTATGTGCTGCCC
CGGGAATTACTGCAAAAATGGAATATGTGTGTCTTCTGATCAAAAT
CATTTCCGAGGAGAAAATTGAGGAAACCATCACTGAAAGCTTTGGTA
ATGATCATAGCACCTTGGATGGGTATTCCAGAAGAACCACCTTGTC
TTCAAAAATGTATCACACCAAAGGACAAGAAGGTTCTGTTTGTCTC
CGGTCATCAGACTGTGCCTCAGGATTGTGTTTGTCTC
GTCCAAGATCTGTAAACCTGTCCTGAAAGAAGGTCAAGTGTGTACC
AAGCATAGGAGAAAAAGGCTCTCATGGACTAGAAATATTCCAGCGTT
GTTACTGTGGAGAAAAGATCACCA
TCAAGCCAGTAATTCTTCTAGGCTTCACACCTTGTCAGAGACACTAA

FIG. 2A

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>zmax1 LBD1 (SEQ ID NO: 169)

CTCATCCTGCCCTGCATGGACTGAGGAACGTCAAAGCCATCGACTAT GACCCACTGGACAAGTTCATCTACTGGGTGGATGGGCGCCAGAACATC AAGCGAGCCAAGGACGACGGGACCCAGCCCTTTGTTTTGACCTCTCTG AGCCAAGGCCAAAACCCAGACAGGCAGCCCCACGACCTCAGCATCGA CATCTACAGCCGGACACTGTTCTGGACGTGCGAGGCCACCAATACCAT CAACGTCCACAGGCTGAGCGGGGAAGCCATGGGGGTGGTGCTGCGTG GGGACCGCGACAAGCCCAGGGCCATCGTCGTCAACGCGGAGCGAGGG TACCTGTACTTCACCAACATGCAGGACCGGGCAGCCAAGATCGAACGC GCAGCCCTGGACGCACCGAGCGCGAGGTCCTCTTCACCACCGGCCTC ATCCGCCCTGTGGCCCTGGTGGTAGACACACACTGGGCAAGCTGTTC TGGGTGGACGCGGACCTGAAGCGCATTGAGAGCTGTGACCTGTCAGG GGCCAACCGCCTGACCCTGGAGGACGCCAACATCGTGCAGCCTCTGGG CCTGACCATCCTTGGCAAGCATCTCTACTGGATCGACCGCCAGCAGCA GATGATCGAGCGTGTGGAGAAGACCACCGGGGACAAGCGGACTCGCA TCCAGGGCCGTGTCGCCCACCTCACTGGCATCCATGCAGTGGAGGAAG TCAGCCTGGAGGAGTTCTCAGCCCACCCATGTGCCCGTGACAATGGTG GCTGCTCCCACATCTGTATTGCCAAGGGTGATGGGACACCACGGTGCT CATGCCCAGTCCACCTCGTGCTCCTGCAGAACCTGCTGACCTGTGGAG AGCCGCCCACCTGCTCCCCGGACCAGTTTGCATGTGCCACAGGGGAGA TCGACTGTATCCCCGGGGCCTGGCGCTGTGACGGCTTTCCCGAGTGCG ATGACCAGAGCGACGAGGAGGGCTGCCCCGTGTGCTCCGCCGCCCAGT TCCCCTGCGCGCGGGTCAGTGTGTGGACCTGCGCCTGCGCTGCGACG GCGAGGCAGACTGTCAGGACCGCTCAGACGAGGCGGACTGTGACGCC ATCTGCCTGCCCAACCAGTTCCGGTGTGCGAGCGGCCAGTGTGTCCTC ATCAAACAGCAGTGCGACTCCTTCCCCGACTGTATCGACGGCTCCGAC GAGCTCATGTGTGAAATCACCAAGCCGCCC

FIG. 2B

>zmax1 LBD4 (SEQ ID NO: 170)

AGGGCCATCGTCAACGCGGAGCGAGGGTACCTGTACTTCACCAA CATGCAGGACCGGCAGCCAAGATCGAACGCGCAGCCCTGGACGGCA CCGAGCGCGAGGTCCTCTTCACCACCGGCCTCATCCGCCCTGTGGCCC TGGTGGTAGACACACACTGGGCAAGCTGTTCTGGGTGGACGCGGAC CTGAAGCGCATTGAGAGCTGTGACCTGTCAGGGGCCAACCGCCTGAC CCTGGAGGACGCCAACATCGTGCAGCCTCTGGGCCTGACCATCCTTGG CAAGCATCTCTACTGGATCGACCGCCAGCAGCAGATGATCGAGCGTG TGGAGAAGACCACCGGGGACAAGCGGACTCGCATCCAGGGCCGTGTC GCCCACCTCACTGGCATCCATGCAGTGGAGGAAGTCAGCCTGGAGGA GTTCTCAGCCCACCCATGTGCCCGTGACAATGGTGGCTGCTCCCACAT CTGTATTGCCAAGGGTGATGGGACACCACGGTGCTCATGCCCAGTCCA CCTCGTGCTCCTGCAGAACCTGCTGACCTGTGGAGAGCCGCCCACCTG CTCCCGGACCAGTTTGCATGTGCCACAGGGGAGATCGACTGTATCCC CGGGGCCTGGCGCTGTGACGGCTTTCCCGAGTGCGATGACCAGAGCG ACGAGGAGGCTGCCCCGTGTGCTCCGCCGCCCAGTTCCCCTGCGCGC GGGGTCAGTGTGGACCTGCGCCTGCGCTGCGACGGCGAGGCAGAC CAACCAGTTCCGGTGTGCGAGCGGCCAGTGTGTCCTCATCAAACAGC AGTGCGACTCCTTCCCCGACTGTATCGACGGCTCCGACGAGCTCATGT GTGAAATCACCAAGCCGCCCTAAGCGGCCGC

FIG. 2C

Screen of DKK1 X Peptide Library

r epilde Library			SEQ ID
name	motif	# hits	NO:
252-1	SVGCLLCAGLGVWSLS	3	171
252-2	WCCCGLFRGVCVWSCGAD	2	172
	D		
252-3	GWRRCDWCGCVSWCWV	1	173
252-4	MPGSVSHCWGGICEAL	8	174
252-15	SCCAVDVCLRCGGWFR	1	175
252-16	SVLGTCCCGGWILCE	2	176
252-17	VLSVCEVCGGVFVRRC	1	177
252-18	GMWYWSGRDCALCWL	1	178
252-19	CTAVMWGVGSVAYLGE	1	179
252-20	WCWWCGCRGVVWR	1	180
252-21	CVCASFCCCVCGLRLL	1	181
252-23	TYEVCEECGGRVRMWV	6	182
252-25	VVVCASCGQVWHGSGA	2	183
252-26	CCRCCHCWDCEWHMCV	1	184
252-27	FCASCCWCGCDCFGWV	2	185
252-32	CDYCWSCGVWCPSSWL	3	186
252-47	VYLCVWCGAARFGCYG	1	187
252-48	FCVCGCCWCWCAACWC	1	188
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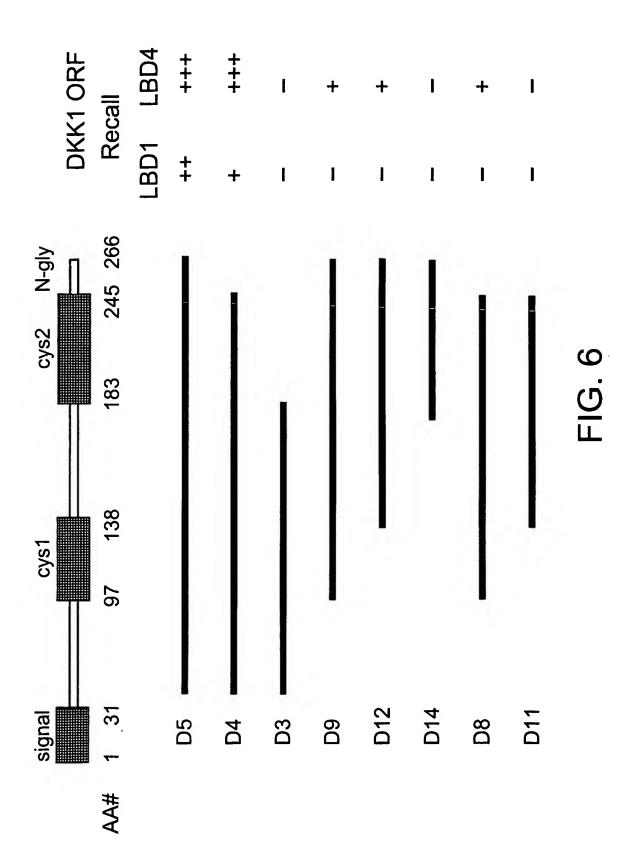
FIG. 3

peptide #	peptide seq	# hits	SEQ ID NO:
9	VVLCSRCGRLWRWSCG	1	189
12	EVRQVTCIRCRRGFLL	1	190
13	GGGGMWEAWSCYACG	1	191
14	GWRWCGRCGALWWRRV	3	192

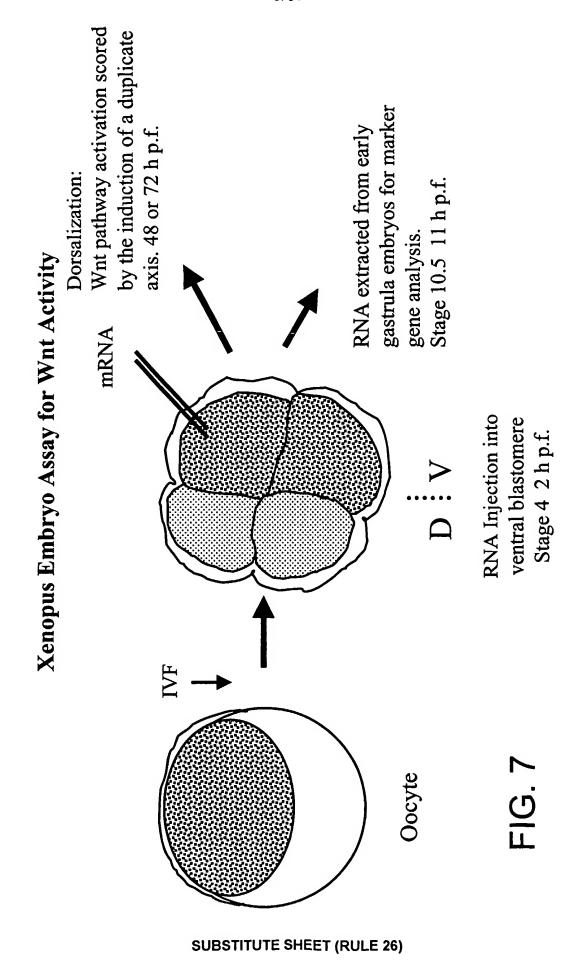
FIG. 4

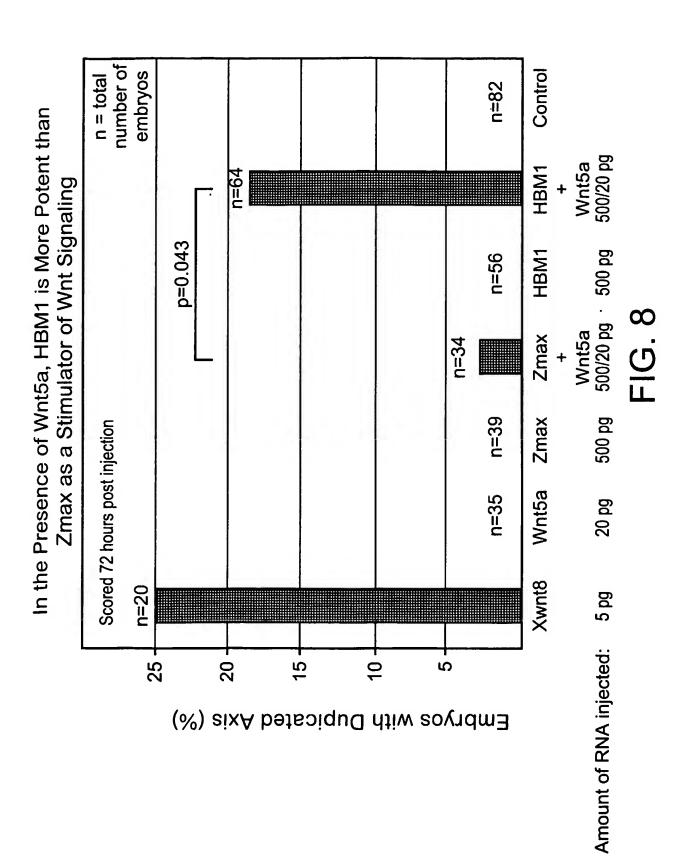
	Genbank	Protein
Gene	Accession #	Accession #
granulin	M75161	AAA58617
similar to cys/His rich protein	BC004544	AAH04544
IGF-BINDING PROTEIN 6	M69054	AAA88070
latent TGFb binding protein 4	AF051344	AAC39879
NOTCH 2	AF315356	AAG37073
fibulin 1	X53743	CAA37772
MDC15 (ADAM15)	U46005	AAC51112
DKFZp761G02121(notch1 Ca++ binding like)	AL137311	CAB70690
chordin	AF076612	AAC69835
fibronectin 1	U42594	AAD00019
MG50(melanoma associated antigen)	AF200348	AAF06354
unknown (notch 4-like)	AX068260	CAC27245
Slit 1	AB017167	BAA35184
tomoregulin (agarin repeat homology)	AB004064	BAA90820
sprouty 1	AF041037	AAC39566
sprouty 2	AF039843	AAC04258
NOV1	X96584	CAA65403
agrin	AF016903	AAC39776
fibrillin 1	L13923	AAB02036
thrombospondin1	X04665	CAA28370
ADAM19	AF134707	AAF22162
Nafl alpha	AJ011895	CAA09855
laminin alpha 5	Z95636	CAB09137
CRIM1	AF167706	AAF34409
nidogen	M30269	AAA59932
fibulin-2	X82494	CAA57876
thrombospondin 2	L12350	AAA03703
KIAA1323	AB037744	BAA92561
fibrillin-2	U03272	AAA18950
MEGF9	AB011542	BAA32470
integrin beta 1	X07979	CAA30790
matrilin-2 precursor	U69263	AAC51260
tenascin	X56160	A32160

FIG. 5



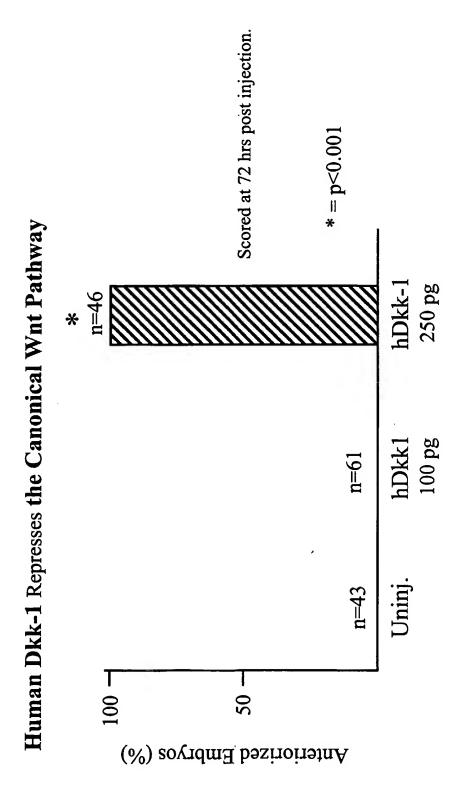
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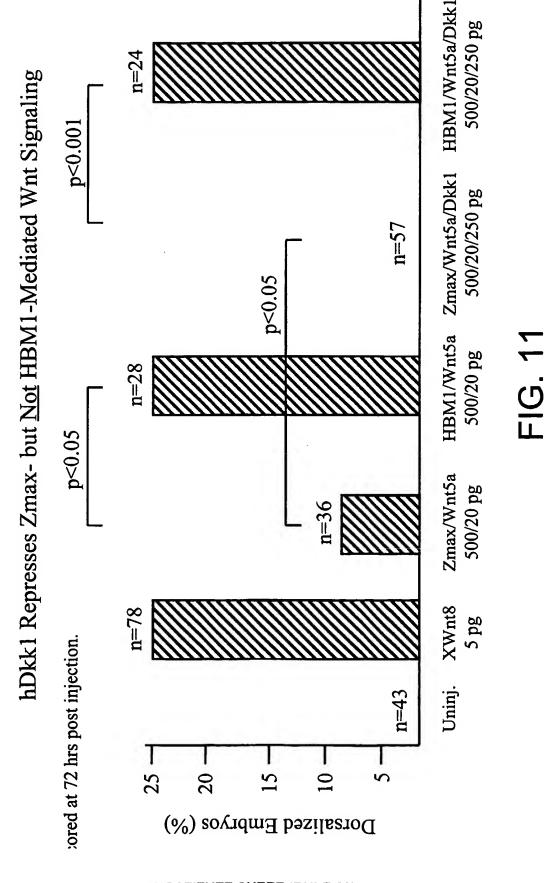
Both Zmax and HBM1, in the presence of Wnt5a, induce secondary axis formation in Xenopus (photos at 48 hrs post-injection) HBM1 + Wnt5a Zmax + Wnt5a XWnt8 Wnt5a **HBM1** Zmax FIG. 9



•Confirms our human construct is active.

•Reproduces reported dose-response.

<u>-1G. 10</u>



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Listed are the pcDNA3.1 construct names followed by the DNA sequence OST258 (control for OST 259-OST262 and OST264,OST265)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCC

OST259 (SEQ ID NO: 193)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACGACTTTTGACACGGATGTACTCAAAGCGGACGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCGTGGTTCTGTGTTCGCGTTGTGGGCGTTTGTGGCGTGG
TCGTGTGGGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAACT
GACCGTTGCAAAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

.OST260 (SEQ ID NO: 194)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTGTTCCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCGGGTGGGGTGTGGTGGGGCTTTGTGGTGG
CGGCGTGTTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAACT
GACCGTTGCAAAACTGAACATCGATCAAAACCCTGGCACTGCCCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST261 (SEQ ID NO: 195)

AAGCTTGCCACCATGGAGACAGACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCMCCCTGCCACAGACTCCGCCAGATTTCCGGGGTTACGTGTATTAGGTGTCGTCGGGGT
TTTCTGTTGACTAGTGGTCCGTGCAAAATGATCGCCCCGGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAACT
GACCGTTGCAAAACTGAACATCGATCAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST262 (SEQ ID NO: 196) ...

AAGCTTGCCACCATGAGACACACACTCCTGCTATGGGTACTGCTGCTGCTGCTGCTGCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAAATTCCGGTGGTGGGGGGGATGATTTGGGAGGCTTGGAGTTGTTAT
GCGTGTGGGACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAACT
GACCGTTGCAAAACTGAACATCGATCAAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

OST263 (SEQ ID NO: 197)

AAGCTTGCCACCATGAGACACACACTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCCATGAGCGATAAAATTATTCACCTGACTGACGACAGTTTTGACACGGATGTACTCAAAGCGGACGGGCGATCC
TCGTCGATTTCTGGGCAGAGTGGTGCGGTCCGAATTCCTTGTGGATTGGGCCGGGTGATCAGGGTCTGTTTCGGCGT
TTTGTTTTTACTAGTGGTCCGTGCAAAATGATCGCCCCGATTCTGGATGAAATCGCTGACGAATATCAGGGCAAAAC
GACCGTTGCAAAACTGAACATCGATCAAAAACCCTGGCACTGCGCCGAAATATGGCATCCGTGGTATCCCGACTCTGC
TGCTGTTCAAAAACCGGTGAAGTGGCGGCAACCAAAGTGGGTGCACTGTCTAAAGGTCAGTTGAAAGAGTTCCTCGAC
GCTAACCTGGCGTAAGCGGCCGC

FIGURE 12A

OST264 (SEQ ID NO: 198)

AAGCTTGCCACCATGGAGACACACACCTCCTGCTATGGGTACTGCTGCTCTGGGTTCCAGGTTCCACTGGTGACGG
ATCCGTGTCTTCTGATCAAAATCATTTCCGAGGAGAAATTGAGGAAACCATCACTGAAAGCTTTGGTAATGATCATA
GCACCTTGGATGGGTATTCCAGAAGAACCACCTTGTCTTCAAAAATGTATCACACCAAAGGACAAGAAGGTTCTGTT
TGTCTCCGGTCATCAGACTGTGCCTCAGGATTGTGTTGTGCTAGACACTTCTGGTCCAAGATCTGTAAACCTGTCCT
GAAAGAAGGTCAAGTGTGTACCAAGCATAGGAGAAAAAGGCTCTCATGGACTAGAAATATTCCAGCGTTGTTACTGTG
GAGAAGGTCTGTCTTGCCGGATACAGAAAGATCACCATCAAGCCAGTAATTCTTCTAGGCTTCACACTTGTCAGAGA
CACTAAGCGGCCGC

OST265 (SEQ ID NO: 199)

OST266 (SEQ ID NO: 200)

OST267 (SEQ ID NO: 201)

OST268 (SEQ ID NO: 202)

FIGURE 12B

Listed below are the amino acid sequences corresponding to the pcDNA3.1 constructs in Appendix 1A OST258

METDTLLLWVLLLWVPGSTGDGS

OST259 (SEQ ID NO: 204)

metdtllwvlllwypgstgdgsmsdkiihltddsfdtdvlkadgailvdfwaewcgpnsvvlcsrcgrlwrwscgt sgpckmiapildeiadeyqgkltvaklnidqnpgtapkygirgiptllfkngevaatkygalskgqlkefldanla

OST260 (SEQ ID NO: 205)

METDTLLLWVLLLWVPGSTGDGSMSDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSGWRWCGRCGALWWRRVT SGPCKMIAPILDEIADEYQGKLTVAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

OST261 (SEQ ID NO: 206)

METDTLLLWVLLLWVPGSTGDGSMSDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSEVRQVTCIRCRRGFLLT SGPCKMIAPILDEIADEYQGKLTVAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

OST262 (SEQ ID NO: 207)

METDTLLLWVLLLWVPGSTGDGSMSDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSGGGGMIWEAWSCYACGT SGPCKMIAPILDEIADEYQGKLTVAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

OST263 (SEQ ID NO: 208)

METDTLLLWVLLLWVPGSTGDGSMSDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSLWIGPGDQGLFRRFVFT SGPCKMIAPILDEIADEYQGKLTVAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

OST264 (SEQ ID NO: 209)

METDTLLLWVLLLWVPGSTGDGSVSSDQNHFRGEIEETLTESFGNDHSTLDGYSRRTTLSSKMYHTKGQEGSVCLRS SDCASGLCCARHFWSKICKPVLKEGQVCTKHRRKGSHGLEIFQRCYCGEGLSCRIQKDHHQASNSSRLHTCQRH

OST265 (SEQ ID NO: 210)

METDTLLLWVLLLWVFGSTGDGSCASPTRGGDAGVQICLACRKRRKRCMRHAMCCFGNYCKNGICVSSDQNHFRGEI EETITESFGNDHSTLDGYSRRTTLSSKMYHTKGQEGSVCLRSSDCASGLCCARHFWSKICKFVLKEGQVCTKHRRKG SHGLEI

FORCYCGEGLSC.

OST256 (SEQ ID NO: 211)

MGDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSYAWLFSCSRCRWWLPWTSGPCKMIAPILDEIADEYQGKLT VAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

OST267 (SEQ ID NO: 212)

MGDKIIHLTDDSFDTDVLKADGAILVDFWAENCGPNSICEVVRLWSRYFWSWVTSGPCKMIAPILDEIADEYQGKLT VAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGOLKEFLDANLA

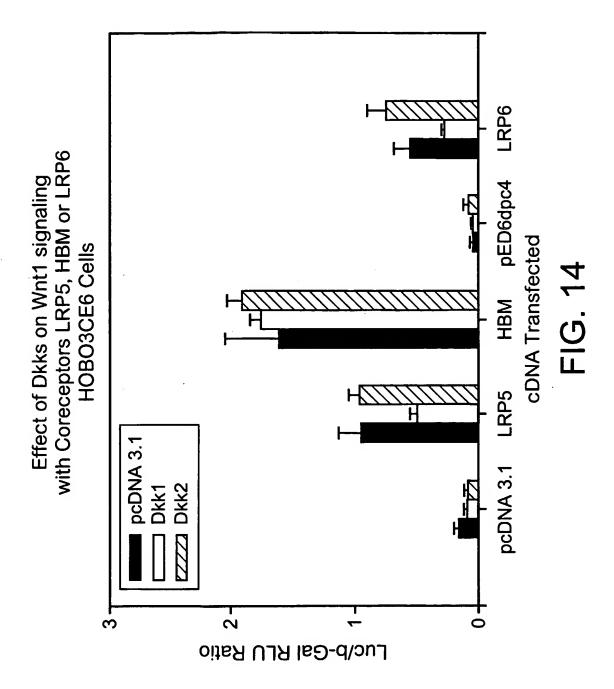
OST268 · (SEQ ID NO: 213)

MGDKIIHLTDDSFDTDVLKADGAILVDFWAEWCGPNSGCTSAVCGAWAEAGRFYCTSGPCKMIAPILDEIADEYQGK LTVAKLNIDQNPGTAPKYGIRGIPTLLLFKNGEVAATKVGALSKGQLKEFLDANLA

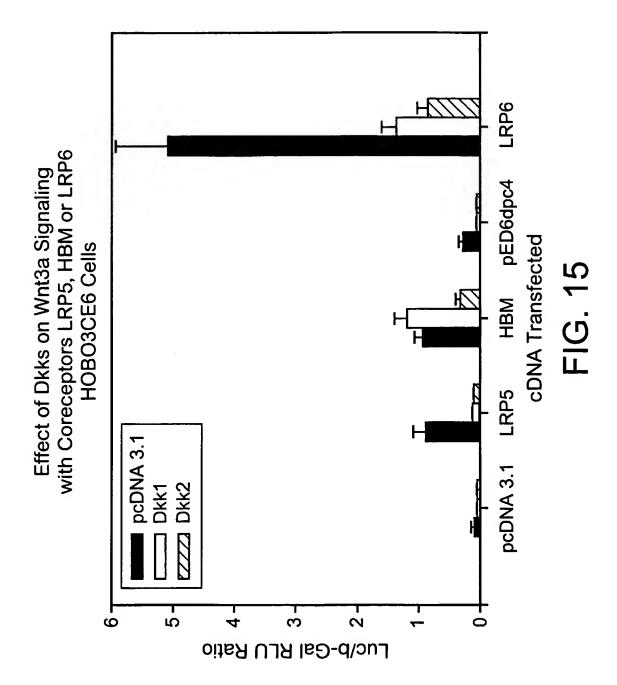
OST269 (SEQ ID NO: 214)

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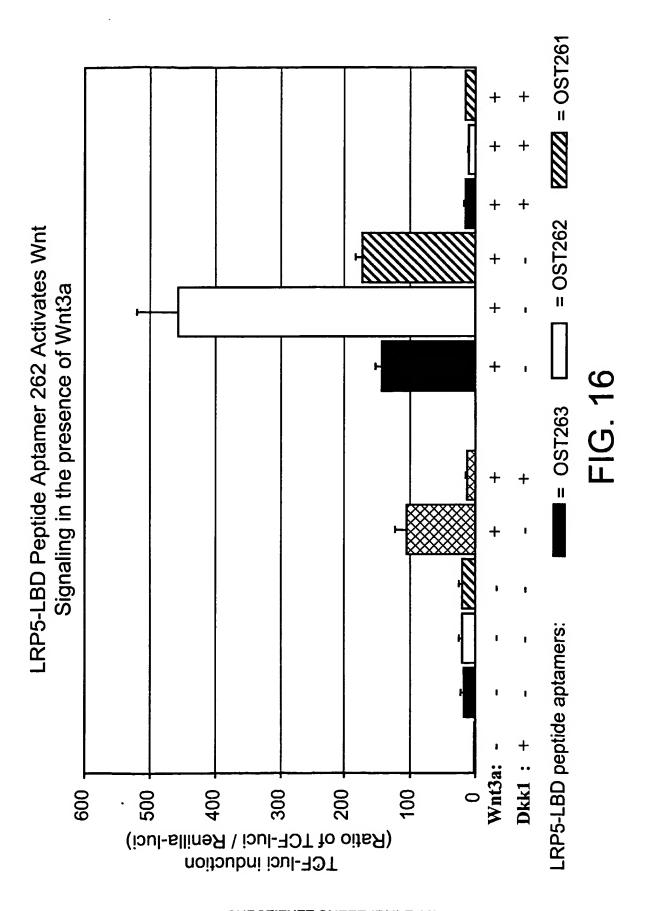
FIGURE 13



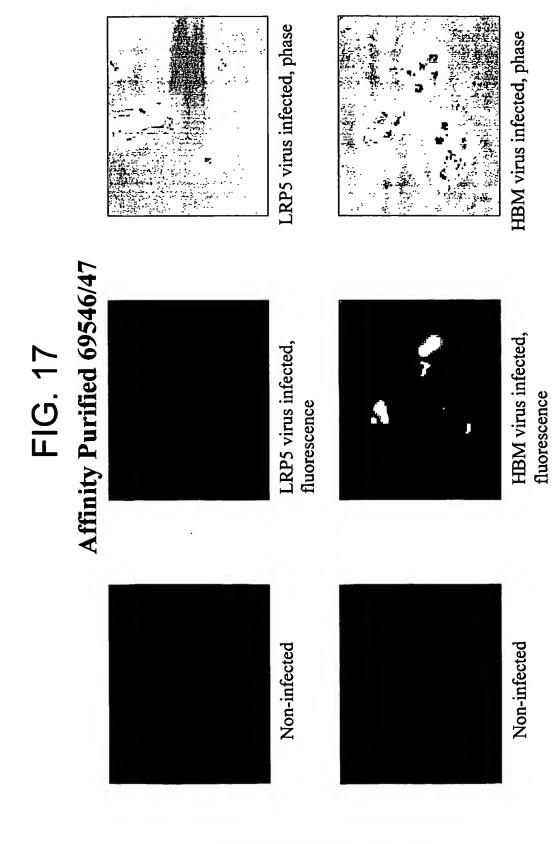
SUBSTITUTE SHEET (RULE 26)



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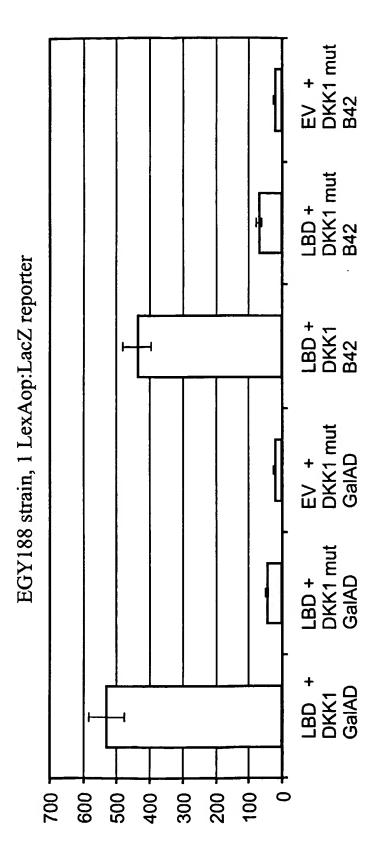


SUBSTITUTE SHEET (RULE 26)



Antibody to: aa 165-177 (Mutation)

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GalAD and B42 and tested for its ability to bind to LBD in Y2H

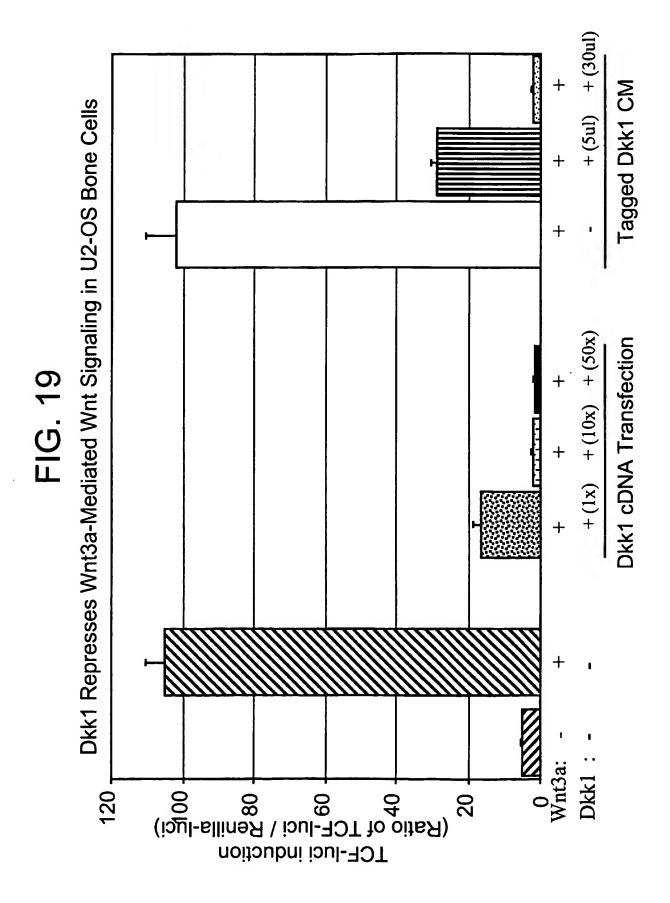
• Interaction LBD-DKK1 20 fold above background

• Interaction LBD-DKK1 C220A 2 to 3 fold above background

• Interaction LBD-DKK1 10 fold above LBD-DKK1 C220A mutant

FIG. 18

· A mutant DKK1, C220A, unable to bind to LRP5, was cloned in



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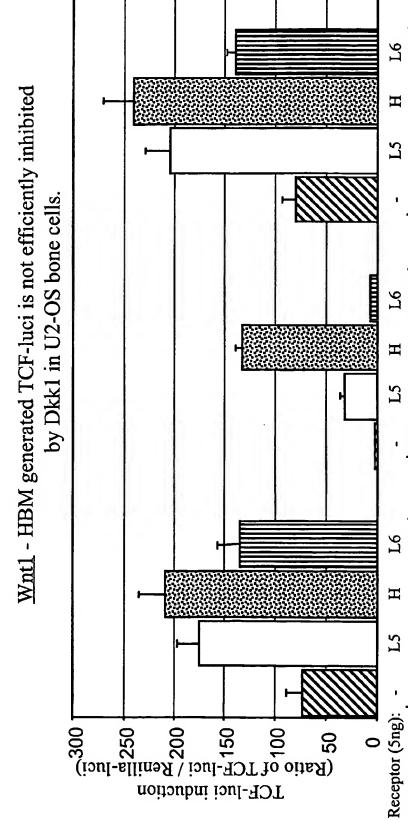
Dkk1-mt (C220A)

Dkk1

pcDNA

(50ng):

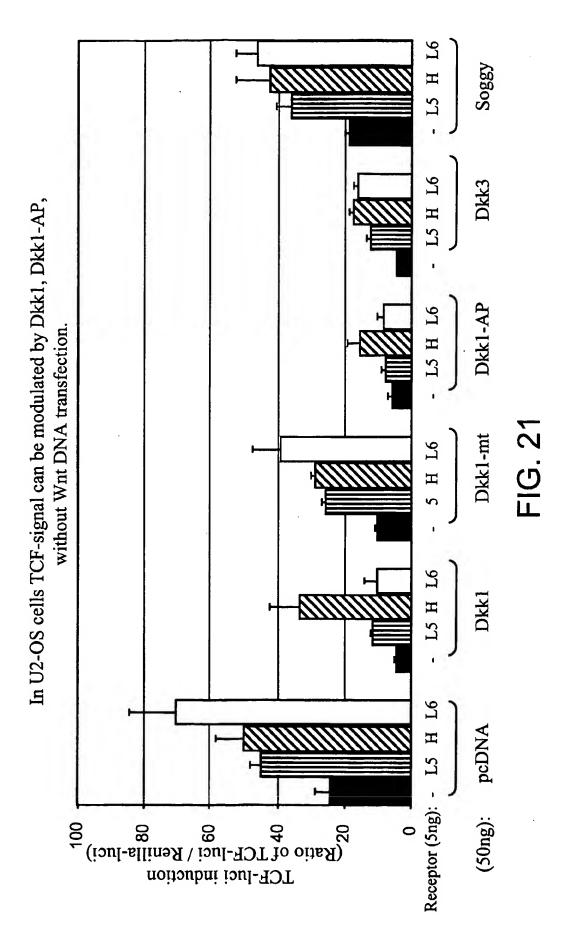
FIG. 20



• With Wnt1 the TCF-signal generated by LRP5 is greater than that of LRP6.

• LRP5/6 -Wnt1 induced TCF- is efficiently blocked byDkk1

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FIG. 22

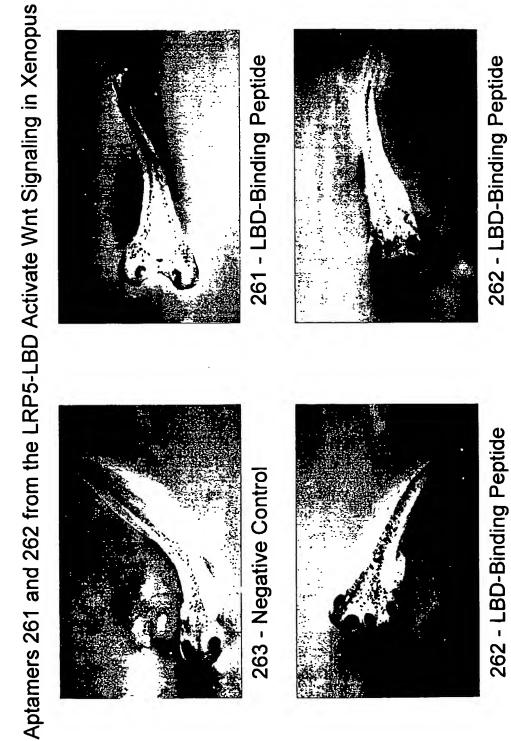
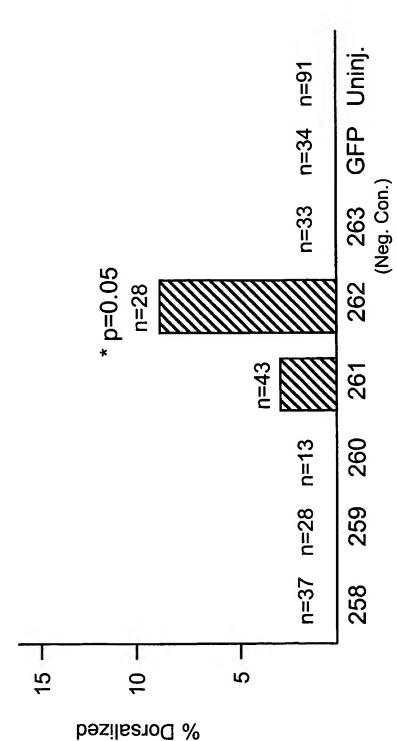
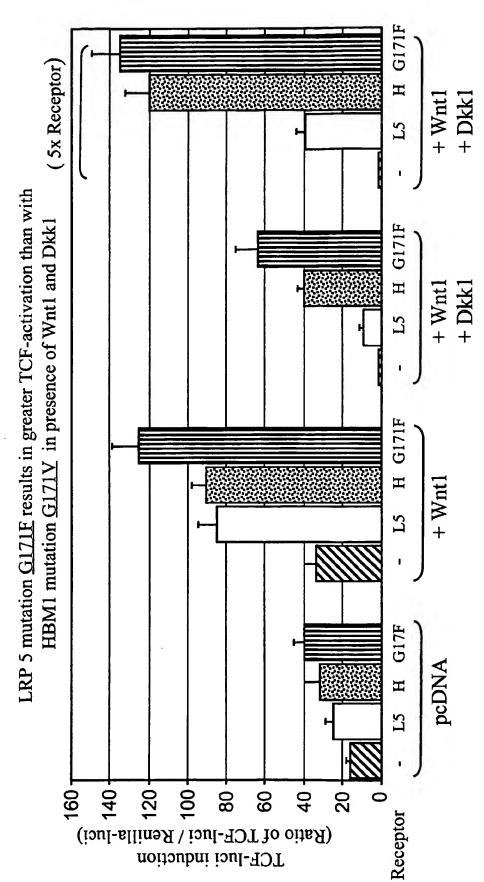


FIG. 23

LRP5 Peptide Aptamers 261 and 262 Induce Wnt Signaling



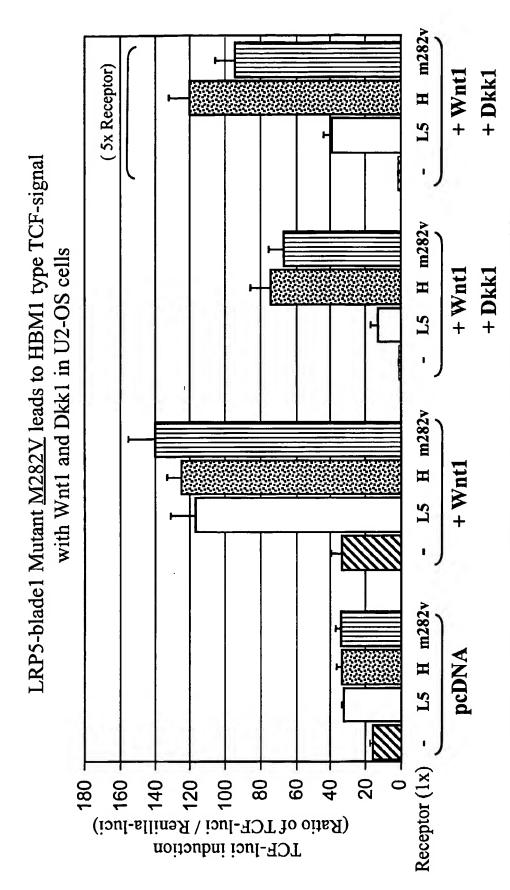
250 pg 250 pg 250 pg 250 pg 250 pg 250 pg RNA: 250 pg



• G171E mutation involves the ringed R group (F) alteration and leads to marginally greater TCF-luci activation than that with HBM1 mutation G171V

FIG. 24

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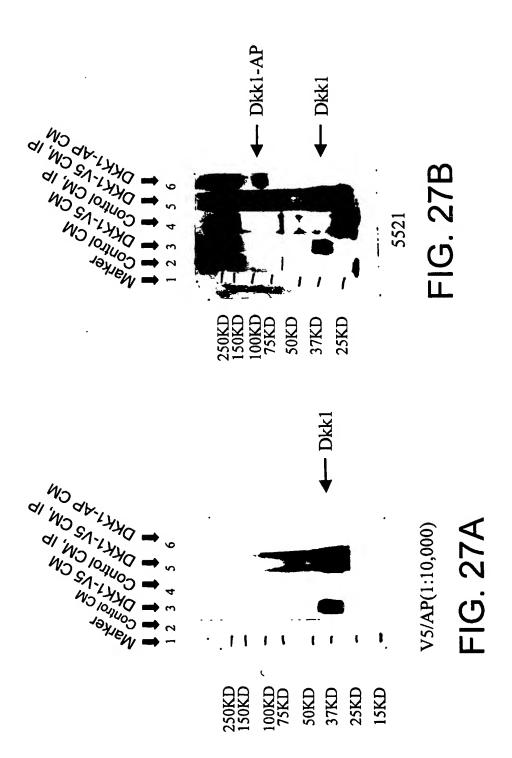
• In blade 1, propeller 1, M282 is at the accessible interior position.

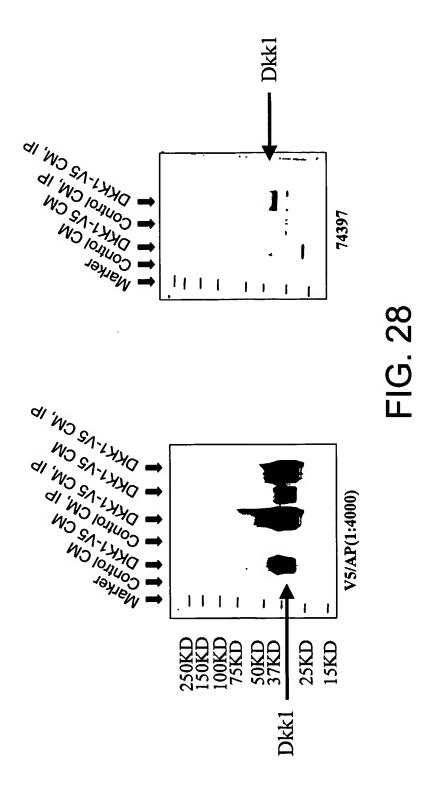
It is conserved in propellers 1-3

FIG. 25

5523/ 5524 266 70624/ 25 245 Cys-2 5521/5522 165-186* 183 DKK1 Protein Polyclonal Antibodies 74396/74397 147-161 138 No titer 71-85* 87-103* 97 5505/5506 * Sigma/Genosys 31 † ResGen Signal Peptides Selected Rabbits Amino Acid

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SEQUENCE LISTING

<110> Allen, Kristina M.
Anisowicz, Anthony
Bhat, Bheem
Damagnez, Veronique
Robinson, John
Yaworsky, Paul

<120> Reagents and Method for Modulating DKK-Mediated Interactions

<130> 032796-132

<150> US 60/291,311

<151> 2001-05-17

<150> US 60/353,058

<151> 2002-02-01

<150> US 60/361,293

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Leu Leu Leu Leu Leu Leu Ala Leu Cys Gly Cys Pro Ala Pro Ala	
15 20 25	
gcg gcc tcg ccg ctc ctg cta ttt gcc aac cgc cgg gac gta cgg ctg	205
Ala Ala Ser Pro Leu Leu Phe Ala Asn Arg Arg Asp Val Arg Leu	
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gtg gac gcc ggc gga gtc aag ctg gag tcc acc atc gtg gtc agc ggc	253
Val Asp Ala Gly Gly Val Lys Leu Glu Ser Thr Ile Val Val Ser Gly	
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ctg gag gat gcg gcc gca gtg gac ttc cag ttt tcc aag gga gcc gtg	301
Leu Glu Asp Ala Ala Ala Val Asp Phe Gln Phe Ser Lys Gly Ala Val	
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Tyr Trp Thr Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn	
80 85 90	
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Gln																
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ccc	gac	ggc	ctc	qcc	tac	gac	taa	ata	aac	aag	aag	cta	tac	taa	acq	445
Pro	Asp	Glv	Leu	Ála	Cvs	Asp	Trp	Val	Glv	Lvs	LVS	Leu	Tyr	Trn	Thr	113
110	•	2			115				-	120	-,0		- 1 -			
	+00	~~~	200												125	
	tca															493
Asp	Ser	Glu	Thr	Asn	Arg	Ile	Glu	Val		Asn	Leu	Asn	Gly	Thr	Ser	
				130					135					140		
cgg	aag	gtg	ctc	ttc	tgg	cag	gac	ctt	gac	cag	ccg	agg	gcc	atc	qcc	541
	Lys															
,			145					150					155		1170	
++~	~~~											.				
	gac															589
ren	Asp		Ата	HIS	GIA	Tyr		Tyr	Trp	Thr	Asp		GTA	Glu	Thr	
		160					165					170				
CCC	cgg	att	gag	cgg	gca	ggg	atg	gat	ggc	agc	acc	cgg	aag	atc	att	637
Pro	Arg	Ile	Glu	Ara	Ala	Glv	Met	Asp	Glv	Ser	Thr	Ara	Lvs	Ile	Ile	
	175			,		180			1		185	9	-,-			
ata		+~~	~~~	2++	+			+	~~~	a+ a						CO.
	gac															685
	Asp	ser	Asp	TTE	_	Trp	Pro	Asn	GTA		Thr	TTE	Asp	Leu	Glu	
190					195					200					205	
gag	cag	aag	ctc	tac	tgg	gct	gac	gcc	aag	ctc	agc	ttc	atc	cac	cqt	733
	Gln															
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acc	aac	cta	~~~		toa	++0	~~~	026		~+~	~+~	~~~	~~~			201
																781
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Thr	His	Pro	Phe	Ala	Leu	Thr	Leu	Ser	Glv	Asp	Thr	Leu	Tvr	Trp	Thr	
		240					245		- 4			250	- 4 -	F		
rac	tgg		200	cac	too	ato		~~~	+~~	220	226		20+	~~~	~~~	077
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ASD	Trp	GIN	Int	Arg	ser		HIS	Ala	Cys	Asn		Arg	Thr	GLA	GTA	
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	agg															925
Lys	Arg	Lys	Glu	Ile	Leu	Ser	Ala	Leu	Tyr	Ser	Pro	Met	Asp	Ile	Gln	
270					275					280			-		285	
ata	cta	agc														
			caq	gag	caa	cad	cct	ttc	ttc		act	cac	tat	gag		973
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		Ser		Glu		cag Gln			Phe	cac				Glu	gag	973
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Asp ttc Phe agg Arg acg Thr 350 gtg	aat Asn tac Tyr acg Thr 335 gac Asp	ggc Gly aca Thr 320 tgt Cys cta Leu	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg	Glu 290 tgc Cys gcc Ala gca Ala agg Arg	tcc Ser tgc Cys gga Gly atc Ile 355 gac	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac	Phe 295 ctg Leu gtg Val gtg Val acg Thr	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc	Cys agc Ser 315 gac Asp gcc Ala acc Thr	Glu 300 gag Glu aac Asn cgg Arg gac Asp	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac	1021 1069 1117 1165
Asp ttc Phe agg Arg acg Thr 350 gtg	aat Asn tac Tyr acg Thr 335 gac Asp	ggc Gly aca Thr 320 tgt Cys cta Leu	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg	Glu 290 tgc Cys gcc Ala gca Ala agg Arg	tcc Ser tgc Cys gga Gly atc Ile 355 gac	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac	Phe 295 ctg Leu gtg Val gtg Val acg Thr	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc	Cys agc Ser 315 gac Asp gcc Ala acc Thr	Glu 300 gag Glu aac Asn cgg Arg gac Asp	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac	1021 1069 1117 1165
Asp ttc Phe agg Arg acg Thr 350 gtg Val	aat Asn tac Tyr acg Thr 335 gac Asp ctg Leu	ggc Gly aca Thr 320 tgt Cys cta Leu cag Gln	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg gtg Val	Glu 290 tgc Cys gcc Ala gca Ala agg Arg	tcc Ser tgc Cys gga Gly atc Ile 355 gac Asp	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc Ile	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg Arg	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac His	Phe 295 ctg Leu gtg Val acg Thr gcc Ala 375	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att Ile	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp gcc Ala	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc Ile	Cys agc Ser 315 gac Asp gcc Ala acc Thr gac Asp	Glu 300 gag Glu aac Asn cgg Arg gac Asp tac Tyr 380	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac Asp	1021 1069 1117 1165
Asp ttc Phe agg Arg acg Thr 350 gtg Val	aat Asn tac Tyr acg Thr 335 gac Asp ctg Leu	ggc Gly aca Thr 320 tgt Cys cta Leu cag Gln	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg gtg Val	Glu 290 tgc Cys gcc Ala gca Ala agg Arg gac Asp 370 tat	tcc Ser tgc Cys gga Gly atc Ile 355 gac Asp	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc Ile tac	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg Arg	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac His	Phe 295 ctg Leu gtg Val acg Thr gcc Ala 375 gat	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att Ile	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp gcc Ala	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc Ile gtg	Cys agc Ser 315 gac Asp gcc Ala acc Thr gac Asp	Glu 300 gag Glu aac Asn cgg Arg gac Asp tac Tyr 380 gcc	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac Asp	1021 1069 1117 1165
Asp ttc Phe agg Arg acg Thr 350 gtg Val	aat Asn tac Tyr acg Thr 335 gac Asp ctg Leu	ggc Gly aca Thr 320 tgt Cys cta Leu cag Gln	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg gtg Val ggc Gly	Glu 290 tgc Cys gcc Ala gca Ala agg Arg gac Asp 370 tat	tcc Ser tgc Cys gga Gly atc Ile 355 gac Asp	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc Ile tac	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg Arg	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac His aca Thr	Phe 295 ctg Leu gtg Val acg Thr gcc Ala 375 gat	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att Ile	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp gcc Ala	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc Ile gtg	Cys agc Ser 315 gac Asp gcc Ala acc Thr gac Asp	Glu 300 gag Glu aac Asn cgg Arg gac Asp tac Tyr 380 gcc	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac Asp	1021 1069 1117 1165
Asp ttc Phe agg Arg acg Thr 350 gtg Val ccg Pro	aat Asn tac Tyr acg Thr 335 gac Asp ctg Leu	ggc Gly aca Thr 320 tgt Cys cta Leu cag Gln gag Glu	Gln ggc Gly 305 tgc Cys aag Lys cgg Arg gtg Val ggc Gly 385	Glu 290 tgc Cys gcc Ala gca Ala agg Arg gac Asp 370 tat Tyr	tcc Ser tgc Cys gga Gly atc Ile 355 gac Asp	Gln cac His ccc Pro gcc Ala 340 tcg Ser atc Ile tac Tyr	Pro ctg Leu acg Thr 325 gag Glu ctg Leu cgg Arg tgg Trp	Phe tgc Cys 310 ggt Gly gag Glu gac Asp cac His aca Thr 390	Phe 295 ctg Leu gtg Val acg Thr Gcc Ala 375 gat Asp	cac His ctg Leu cag Gln ctg Leu ccg Pro 360 att Ile gac Asp	Thr tcc Ser ctg Leu ctg Leu 345 gac Asp gcc Ala gag Glu	Arg cca Pro cag Gln 330 ctg Leu ttc Phe atc Ile gtg Val	Cys agc Ser 315 gac Asp gcc Ala acc Thr gac Asp cgg Arg 395	Glu 300 gag Glu aac Asn cgg Arg gac Asp tac Tyr 380 gcc Ala	gag Glu cct Pro ggc Gly cgg Arg atc Ile 365 gac Asp atc Ile	1021 1069 1117 1165

Arg	Arg	Ala 400	Tyr	Leu	Asp	Gly	Ser 405	Gly	Ala	Gln	Thr	Leu 410	Val	Asn	Thr		
									gtc Val							1	.357
									cgc Arg							1	405
									tcg Ser 455							1	453
									ggc Gly							1	501
									gcc Ala							1	549
									ggg Gly							1	.597
									tgg Trp							1	.645
									acg Thr 535							1	.693
									ttc Phe							1	.741
									agc Ser							1	.789
									gac Asp							1	.837
									gtc Val								1885
									ctg Leu 615							1	1933
									ctg Leu							1	1981
									ttg Leu							2	
									aat Asn							2	2077
	ctc								gcc Ala							2	2125
aac					tgg				agc Ser 695	ctg					cgc	2	2173
gcc									~ ~ ~								

Ala	Phe	Met	Asn 705	Gly	Ser	Ser	Val	Glu 710	His	Val	Val	Glu	Phe	Gly	Leu	
gac	tac	ccc	gag	ggc	atg	gcc	gtt	gac	tgg	atg	ggc	aag	aac	ctc	tac	2269
										Met						
										gtg						2317
_	735	_				740	-			Val	745	_		-	-	
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750		_			755		_	_	_	Leu 760	_			_	765	
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				770					775	Tyr				780	_	
										gac						2461
			785					790		Asp			795	_		0500
										gac Asp						2509
		800		_		_	805			-		810		•	-	٥٥٢٦
										gac Asp						2557
	815					820		_		_	825					2605
										gtg Val						2605
830	002			200	835	01	014	n.y	*41	840	110	ALG	пор	лэр	845	
					-	_	_		_	gat						2653
				850				_	855	Asp	٠.		-	860		
										gac						2701
	_		865					870		Asp	_		875		-	
										ttc						2749
	_	880				_	885			Phe		890	-			0707
										aat Asn						2797
	895					900		_			905	_				2045
										atc						2845
910	GTĂ	GIII	Cys	GIY	915	neu	Cys	Leu	Ala	11e 920	FLO	GIÀ	GIY	птэ	925	
tgc	ggc	tgc	gcc	tca	cac	tac	acc	ctg	gac	ccc	agc	agc	cgc	aac	tgc	2893
Cys	Gly	Cys	Ala	Ser 930	His	Tyr	Thr	Leu	Asp 935	Pro	Ser	Ser	Arg	Asn 940	Cys	
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			945					950		Gln	_		955			
										gat						2989
		960		_	_		965			Asp		970				
										tat						3037
	975					980			_	Tyr	985			_	_	
										atc						3085
	TTE	ryr	Trp	val	995	GIY	Arg	GIN	Asn	Ile 1000	_	Arg	Ala	Lys	-	
990 gac	aaa	acc	cad	ccc		att	tta	acc	tct	ctg		Caa	aac	Caa	1005	3133
940	777		4			900	LLY	400		ucy	uy c	Laa	390	Caa	uac	

Asp	Gly	Thr	Gln	Pro 1010		Val	Leu	Thr	Ser 1015		Ser	Gln	Gly	Gln 1020		
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Pro	Asp	Arg			His	Asp	Leu	Ser	Ile	Asp	Ile	Tyr	Ser	Arg	Thr	
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Leu	Phe	Trp	Thr	Cys	Glu	Ala	Thr	Asn	Thr	Ile	Asn	Val	His	Arg	Leu	
		1040)				1045	5				1050)	_		
agc	ggg	gaa	gcc	atg	ggg	gtg	gtg	ctg	cgt	ggg	gac	cqc	qac	aaq	ccc	3277
											Asp					
	1055				•	1060				1	1065	_		-4-		
agg	qcc	atc	atc	atc	aac	aca	gag	сσа	aaa	tac	ctg	tac	ttc	acc	aac	3325
											Leu					JJ25
1070					1075			9	1	1080		- 1 -	• • • •		1085	
		gac	caa	aca			atc	maa	cac		gcc	cta	asc.	aac		3373
Met	Gln	Asn	Ara	Δla	Ala	Lue	Tla	Glu	Ara	31 s	Ala	Lou	700	Cli	mb-	3373
1100	01	nop	ALG	1090			116	GIU	1095		AIG	Leu	Asp	-		
~~~		~~~	~+~											1100		
											cgc					3421
GIU	Arg	GIU			Phe	Thr	Thr			TTE	Arg	Pro			Leu	
			1105					1110					1115			
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Val	Val			Thr	Leu	Gly	Lys	Leu	Phe	Trp	Val	Asp	Ala	Asp	Leu	
		1120					1125					1130				
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Lys	Arg	Ile	Glu	Ser	Cys	Asp	Leu	Ser	Gly	Ala	Asn	Arq	Leu	Thr	Leu	
	1135				_	1140			_		1145	_				
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											Thr					
1150					1155			,-	3	1160				1	1165	
		tac	taa	atc			сап	сап	cad		atc	nan	cat	ata		3613
											Ile					3013
0		-1-		1170		****	01	01	1175		116	O L iu	arg	1180		
220	200	200	aaa			caa	2.C+	000			ggc	cat	ata			3661
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цуs	1111	1111	1185		гуз	Ary	IIII			GTII	GIA	Arg			nis	
								1190					1195			
											ctg					3709
Leu				HIS	ALA	vai			vaı	Ser	Leu			Phe	Ser	
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											tcc					3757
Ala			Cys	Ala	Arg	Asp	Asn	Gly	Gly	Cys	Ser	His	Ile	Cys	Ile	
	1215	•				1220	)				1225	•				
gcc	aag	ggt	gat	ggg	aca	cca	cgg	tgc	tca	tgc	cca	gtc	cac	ctc	gtg	3805
Ala	Lys	Gly	Asp	Gly	Thr	Pro	Arg	Cys	Ser	Cys	Pro	Val	His	Leu	Val	
1230	)				1235	5				1240	)				1245	
ctc	ctq	caq	aac	cta	cta	acc	tqt	qqa	gag	cca	ccc	acc	tac	tcc	cca	3853
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aac	can	+++	aca			aca	aaa	nan			tgt	atc	ccc	-		3901
											Cys					J J G I
пор	01		1265		ALG	1112	OLY	1270		nsp	Cys	116	1275		VIG	
+~~	000	+ ~+			+++	~~~	<b>~~~</b>			~~~	~~~	266			~~~	3949
											cag					J 74 Y
rrp	Arg			стА	rne	Pro			ASP	ASP	Gln		_	GIU	GIU	
		1280					1285					1290				2025
											tgc					3997
			val	Cys	Ser			Gln	Phe	Pro	Cys		Arg	Gly	Gln	
	1295					1300	)				1305	Ò				
	gtg	gac	ctg	cgc	ctg	cgc	tgc	gac	ggc	gag	gca	gac	tgt	cag	gac	4045

Cys Val Asp Leu 1310	Arg Leu Arg 1315	Cys Asp Gly	Glu Ala Asp Cys Gln 1320	Asp 1325
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		Pro Ala His	agc agt gcc atc ggg Ser Ser Ala Ile Gly 1385	
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gtg tgc cag cgc	gtg gtg tgc		gcg ggg gcc aac ggg Ala Gly Ala Asn Gly	ccc 4333 Pro
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ata gcc ccg ggc	ggt tcc cag	cat ggc ccc	ttc aca ggc atc gca Phe Thr Gly Ile Ala 1450	tgc 4429 Cys
gga aag tcc atg		gtg agc ctg Val Ser Leu	atg ggg ggc cgg ggc Met Gly Gly Arg Gly 1465	
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age gae tae age	gcc agc cgc	tgg aag gcc	agc aag tac tac ctg Ser Lys Tyr Tyr Leu 1560	
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Gln Tyr Leu Ser	gcg gag gac Ala Glu Asp	agc tgc ccg	ccc tcg ccc gcc acc Pro Ser Pro Ala Thr	gag 4861
Arg Ser Tyr Phe	cat ctc ttc	ccg ccc cct Pro Pro Pro	1595 ccg tcc ccc tgc acg Pro Ser Pro Cys Thr	
1600 tca tcc tgacctc	ggc cgggccac	1605 tc tggcttctc	1610 t gtgcccctgt aaatagt	ttt 4965

Ser Ser 1615 5025 taaaaacatg agaaatgtga actgtgatgg ggtgggcagg gctgggagaa ctttgtacaq 5085 tqqaqaaata tttataaact taattttgta aaaca 5120 <210> 2 <211> 5120 <212> DNA <213> Homo sapiens <400> 2 actaaagege egeegeege ceatqgagee egagtgageg eggegegge eegteegge 60 geoggacaac atg gag gea geg eeg eeg eeg eeg tgg eeg etg etg 109 Met Glu Ala Ala Pro Pro Gly Pro Pro Trp Pro Leu Leu ctg ctg ctg ctg ctg ctg ctg gcg ctg tgc ggc tgc ccg gcc ccc gcc 157 Leu Leu Leu Leu Leu Leu Ala Leu Cys Gly Cys Pro Ala Pro Ala 20 geg gec teg eeg ete etg eta ttt gee aac ege egg gae gta egg etg 205 Ala Ala Ser Pro Leu Leu Phe Ala Asn Arg Arg Asp Val Arg Leu gtg gac gcc ggc gga gtc aag ctg gag tcc acc atc gtg gtc agc ggc 253 Val Asp Ala Gly Gly Val Lys Leu Glu Ser Thr Ile Val Val Ser Gly 50 ctg gag gat gcg gcc gca gtg gac ttc cag ttt tcc aag gga gcc gtg 301 Leu Glu Asp Ala Ala Ala Val Asp Phe Gln Phe Ser Lys Gly Ala Val 70 tac tgg aca gac gtg agc gag gcc atc aag cag acc tac ctg aac 349 Tyr Trp Thr Asp Val Ser Glu Glu Ala Ile Lys Gln Thr Tyr Leu Asn 85 cag acg ggg gcc gcc gtg cag aac gtg gtc atc tcc ggc ctg gtc tct 397 Gln Thr Gly Ala Ala Val Gln Asn Val Val Ile Ser Gly Leu Val Ser 100 445 ccc gac ggc ctc gcc tgc gac tgg gtg ggc aag aag ctg tac tgg acg Pro Asp Gly Leu Ala Cys Asp Trp Val Gly Lys Lys Leu Tyr Trp Thr 120 115 gac tca gag acc aac cgc atc gag gtg gcc aac ctc aat ggc aca tcc 493 Asp Ser Glu Thr Asn Arg Ile Glu Val Ala Asn Leu Asn Gly Thr Ser 130 135 cqq aaq qtq ctc ttc tqq caq qac ctt gac cag ccg agg gcc atc gcc 541 Arg Lys Val Leu Phe Trp Gln Asp Leu Asp Gln Pro Arg Ala Ile Ala 150 589 ttg gac ccc gct cac ggg tac atg tac tgg aca gac tgg gtt gag acg Leu Asp Pro Ala His Gly Tyr Met Tyr Trp Thr Asp Trp Val Glu Thr 165 ccc cgg att gag cgg gca ggg atg gat ggc agc acc cgg aag atc att 637 Pro Arg Ile Glu Arg Ala Gly Met Asp Gly Ser Thr Arg Lys Ile Ile 175 180 185 gtg gac tcg gac att tac tgg ccc aat gga ctg acc atc gac ctg gag 685 Val Asp Ser Asp Ile Tyr Trp Pro Asn Gly Leu Thr Ile Asp Leu Glu 195 200 733 gag cag aag ctc tac tgg gct gac gcc aag ctc agc ttc atc cac cgt Glu Gln Lys Leu Tyr Trp Ala Asp Ala Lys Leu Ser Phe Ile His Arg

				210					215					220		
					tcg											781
Ala	Asn	Leu	Asp	Gly	Ser	Phe	Arg	Gln	Lys	Val	Val	Glu	Gly	Ser	Leu	
													005			
			225					230					235			
					ctg											829
Thr	HIS		Pne	Ата	Leu	Thr		Ser	GTA	Asp	Thr		Tyr	Trp	Thr	
		240					245					250				
-		_		-	tcc			-	_		-	_				877
Asp		GIn	Thr	Arg	Ser		His	Ala	Cys	Asn		Arg	Thr	GIY	Gly	
	255					260					265				•	
					ctg											925
_	Arg	гÀ2	GIU	тте	Leu	Ser	Ата	ren	Tyr		Pro	met	Asp	TTE		
270					275					280			A		285	070
					cgg											973
Val	neu	Ser	GIII	290	Arg	GIN	PIO	Pne	295	urs	Inr	Arg	Cys		GIU	
~~~	22+	~~~	~~~		+		-+-			a+ ~	+			300		1001
					tcc Ser											1021
Asp	ASII	GIY	305	Cys	Ser	птз	Leu	310	reu	Leu	Ser	PIO	315	GIU	PIO	
++0	+ 20	202		~~~	+				~+~			~~~				1069
			_	_	tgc		-			_	_	_	_			1009
File	ıyı	320	Cys	MIG	Cys	PIO	325	GIY	Val	GIII	Leu	330	ASP	ASII	GIA	
200	200		224	~~	~~~	~~~		~~~	a+a	c+ a	c+ a		~~~		~~~	1117
					gga Gly											111/
Arg	335	Cys	цуз	VIG	GIY	340	Giu	GIU	VOI	neu	345	пеп	лта	nry	ALG	
200		cta	caa	200	atc		cta	aac.	200	cca		ttc	200	asc	ato	1165
					Ile											1100
350	, 10 b	DÇU	,, <u>r</u> A	my	355	JCI	пец	nop	1111	360	nop		1111	пор	365	
	cta	саσ	ata	gac	gac	atc	caa	cac	acc		acc	atc	gac	tac		1213
					Asp											
		·		370					375					380		
CCG	cta	gag	aac		gtc	tac	taa	aca		gac	gag	ata	caa		atc	1261
					Val											
			385	-] -		- , -		390					395			
cac	agg	aca		cta	gac	aaa	tct		aca	cag	acq	cta		aac	acc	1309
					Asp											
	•	400	-			3	405	4				410	-			
qaq	atc	aac	gac	ccc	gat	qqc	atc	qcq	qtc	gac	tqq	ata	qcc	cqa	aac	1357
					Asp											
	415		•		•	420				•	425					
ctc	tac	tgg	acc	gac	acg	ggc	acg	gac	cgc	atc	gag	gtg	acg	cgc	ctc	1405
					Thr											
430	_	-		_	435			_	_	440					445	
aac	ggc	acc	tcc	cgc	aag	atc	ctg	gtg	tcg	gag	gac	ctg	gac	gag	ccc	1453
Asn	Gly	Thr	Ser	Arg	Lys	Ile	Leu	Val	Ser	Glu	Asp	Leu	Asp	Glu	Pro	
	_			450					455					460		•
cga	gcc	atc	gca	ctg	cac	CCC	gtg	atg	ggc	ctc	atg	tac	tgg	aca	gac	1501
Arg	Ala	Ile	Ala	Leu	His	Pro	Val	Met	Gly	Leu	Met	Tyr	Trp	Thr	Asp	
			465					470					475			
tgg	gga	gag	aac	cct	aaa	atc	gag	tgt	gcc	aac	ttg	gat	ggg	cag	gag	1549
Trp	Gly	Glu	Asn	Pro	Lys	Ile		Cys	Ala	Asn	Leu	Asp	Gly	Gln	Glu	
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					aat											1597
Arg	_	Val	Leu	Val	Asn		Ser	Leu	Gly	Trp		Asn	Gly	Leu	Ala	
	495					500					505					.
ctg	gac	ctg	cag	gag	ggg	aag	ctc	tac	tgg	gga	gac	gcc	aag	aca	gac	1645

510					515					520	Asp				525	
aag Lys	atc Ile	gag Glu	gtg Val	Ile 530	aat Asn	gtt Val	gat Asp	ggg Gly	acg Thr 535	aag Lys	agg Arg	cgg Arg	acc Thr	ctc Leu 540	ctg Leu	1693
											ctg Leu					1741
											gag Glu					1789
											ctg Leu 585	ccc				1837
						gtg					gga Gly					1885
gcg					ggg					tgc	ttc Phe				cac	1933
				ggc					ctg		ctg Leu			gac		1981
			atc					ttc			ttc Phe		agc			2029
		cac					gag				aac Asn 665	gac				2077
	ctc				Lys	gag				Leu	gac Asp				Ser	2125
aac				Tyr					Ser		aag Lys			Ser		2173
								Glu			gtg Val		Phe			2221
_			gag		_	_	Val	_		_	ggc Gly	Lys				2269
	Ala	gac									gcg Ala 745					2317
Gln					Leu	gtg				Leu	gac Asp				Ser	2365
				Pro					Ile		tgg Trp			Trp		2413
			Arg					Phe			ggg Gly		Asn			2461
		Val					Arg				ctc Leu	Thr				2509
gct	gac	800 cag	cgc	ctc	tac	tgg	805 acc	gac	ctg	gac	acc	810 aac	atg	atc	gag	2557

Ala	Asp 815	Gln	Arg	Leu	Tyr	Trp 820	Thr	Asp	Leu	Asp	Thr 825	Asn	Met	Ile	Glu	
tcg	tcc	aac	atg	ctq	ggt	caq	gag	cqq	gtc	gtg	att	qcc	qac	gat	ctc	2605
									Val							-005
830					835		V	5		840					845	
	C20	000	++0	aat				+	200		+-+	2+0				0.550
									agc							2653
Pro	HIS	Pro	Pne		Leu	Thr	GIn	Tyr	Ser	Asp	Tyr	тте	Tyr	Trp	Thr	
				850					855					860		
gac	tgg	aat	ctg	cac	agc	att	gag	cgg	gcc	gac	aag	act	agc	ggc	cgg	2701
Asp	Trp	Asn	Leu	His	Ser	Ile	Glu	Arg	Ala	Asp	Lys	Thr	Ser	Gly	Arg	
	_		865					870		-	-		875	-	• •	
aac	cac	acc	ctc	atc	cag	aac	cac		gac	ttc	ata	atα		atc	cta	2749
									Asp							2147
ASII	ALG		Leu	116	GIII	GIY		Leu	rab	File	val		ASP	TIE	Leu	
		880					885					890				
									ctc							2797
Val	Phe	His	Ser	Ser	Arg	Gln	Asp	Gly	Leu	Asn	Asp	Cys	Met	His	Asn	
	895					900					905					
aac	ggg	cag	tgt	qqq	caq	ctq	tqc	ctt	gcc	atc	ccc	aac	aac	cac	cac	2845
									Āla							
910	•				915		-1-			920		1	1		925	
-	~~~	t-a-a	~~~	+ 0 0		+ > 0	200	a+ a	~~~			.~.				2002
									gac							2893
Cys	GIA	Cys	Ala		HIS	Tyr	Thr	Leu	Asp	Pro	Ser	Ser	Arg		Cys	
				930					935					940		
agc	ccg	CCC	acc	acc	ttc	ttg	ctg	ttc	agc	cag	aaa	tct	gcc	atc	agt	2941
Ser	Pro	Pro	Thr	Thr	Phe	Leu	Leu	Phe	Ser	Gln	Lys	Ser	Ala	Ile	Ser	
			945					950					955			
caa	ato	atc	CCG	gac	gac	car	cac	agc	ccg	cat	ctc	atc		ccc	cta	2989
									Pro							2000
AL G	Hec		FIO	ASP	ASP	GIII		Ser	PIO	Asp	Leu		Leu	PIO	ren	
		960					965					970	_			
									gac							3037
His		Leu	Arg	Asn	Val	Lys	Ala	Ile	Asp	Tyr	Asp	Pro	Leu	Asp	Lys	
	975					980					985					
ttc	atc	tac	tgg	gtg	gat	qqq	cqc	cag	aac	atc	aaq	cqa	qcc	aaq	gac	3085
									Asn							
990		- 2 -			995	2	5			1000	-	9		-,-	1005	
	~~~	300	000			~++	++-		tct							3133
																2133
ASP	GIA	Inr	GIN			var	ren	inr	Ser		Ser	Gin	GIÀ			
				1010					1015					1020		
									atc							3181
Pro	Asp	Arg	Gln	Pro	His	Asp	Leu	Ser	Ile	Asp	Ile	Tyr	Ser	Arg	Thr	
			1025					1030				-	1035			
cta	ttc	taa	aco	tac	gag	acc	acc	aat	acc	atc	aac	atc	cac	aσσ	cta	3229
_			_	_		•			Thr			_			•	
шец	1110	1040		Cys	Oru	nia	1045		1111	110	A311	1050		my	neu	
																2077
									cgt							3277
Ser			Ala	Met	Gly			Leu	Arg	Gly			Asp	Lys	Pro	
	1055	<b>)</b> .				1060	)				1069	5				•
agg	gcc	atc	gtc	gtc	aac	gcq	gaq	cqa	ggg	tac	ctq	tac	ttc	acc	aac	3325
Ara	Ála	Ile	Val	Val	Asn	Ala	Glu	Ara	Ğĺy	Tvr	Leu	Tvr	Phe	Thr	Asn	
1070					1079				1	1080		- 1 -			1085	
		<b>~~</b>	~~~	<b>a</b> a-			ato	~~~	cgc		-	a+-	~~~	~~~		3373
																2212
Mer	GID	ASP	Arg			гλг	тте	GIU	Arg		ΑΙα	ren	Asp	_		
				1090					1099					1100		
									ctc							3421
Glu	Arg	Glu	Val	Leu	Phe	Thr	Thr	Gly	Leu	Ile	Arg	Pro	Val	Ala	Leu	
			1105					1110			_		1115			
gta	ata	qac	aac	aca	cta	qqc	aaq	cta	ttc	taa	ata	gac			cta	3469
	, ,	,			- 3	,,,		- 7		- 22	<del>-</del> -		<b></b>			

Val Val Asp Asn 1120	_	1125	11	30	
aag cgc att gag Lys Arg Ile Glu 1135	Ser Cys Asp 114	Leu Ser Gl O	y Ala Asn Ar. 1145	g Leu Thr Leu	3517
gag gac gcc aac Glu Asp Ala Asn 1150	Ile Val Gln 1155	Pro Leu Gl	y Leu Thr Il. 1160	e Leu Gly Lys 1165	3565
cat ctc tac tgg His Leu Tyr Trp	Ile Asp Arg 1170	Gln Gln Gl 11	n Met Ile Gl .75	a Arg Val Glu 1180	3613
aag acc acc ggg Lys Thr Thr Gly 118	Asp Lys Arg 5	Thr Arg Il 1190	e Gln Gly Ar	y Val Ala His 1195	3661
ctc act ggc atc Leu Thr Gly Ile 1200	His Ala Val	Glu Glu Va 1205	l Ser Leu Gl 12	Glu Phe Ser	3709
gcc cac cca tgt Ala His Pro Cys 1215	Ala Arg Asp 122	Asn Gly Gl O	y Cys Ser Hi. 1225	s Ile Cys Ile	3757
gcc aag ggt gat Ala Lys Gly Asp 1230	Gly Thr Pro 1235	Arg Cys Se	er Cys Pro Va 1240	l His Leu Val 1245	3805
ctc ctg cag aac Leu Leu Gln Asn		Cys Gly Gl			3853
gac cag ttt gca Asp Gln Phe Ala 126	Cys Ala Thr				3901
tgg cgc tgt gac Trp Arg Cys Asp 1280	ggc ttt ccc Gly Phe Pro	gag tgc ga Glu Cys As 1285	it gac cag ag sp Asp Gln Se 12	r Asp Glu Glu	3949
ggc tgc ccc gtg Gly Cys Pro Val 1295		Ala Gln Ph			3997
tgt gtg gac ctg Cys Val Asp Leu 1310					4045
cgc tca gac gag Arg Ser Asp Glu		Asp Ala Il			4093
cgg tgt gcg agc Arg Cys Ala Ser 134	Gly Gln Cys				4141
ttc ccc gac tgt Phe Pro Asp Cys 1360				s Glu Ile Thr	4189
aag ccg ccc tca Lys Pro Pro Ser 1375		Pro Ala Hi			4237
gtc att ggc atc Val Ile Gly Ile 1390	atc ctc tct	ctc ttc gt	c atg ggt gg		4285
gtg tgc cag cgc Val Cys Gln Arg	gtg gtg tgc	Gln Arg Ty	it gcg ggg gc	c aac ggg ccc	4333
ttc ccg cac gag		_	-		4381

Phe	Pro	His	Glu 1425	Tyr	Val	Ser	Gly	Thr 143		His	Val	Pro	Leu 143		Phe	
ata	qcc	cca	aac	ggt	tcc	caq	cat	aac	ccc	ttc	aca	aac	atc	gca	tac	4429
				Gly												3723
		1440	)	_			1445	5				1450	)		•	
gga	aag	tcc	atg	atg	agc	tcc	gtg	agc	ctg	atg	ggg	ggc	cgg	ggc	ggg	4477
Gly	Lys	Ser	Met	Met	Ser	Ser	Val	Ser	Leu	Met	Gly	Gly	Arg	Gly	Gly	
	145	5				1460	)				1465	5				
gtg	CCC	ctc	tac	gac	cgg	aac	cac	gtc	aca	ggg	gcc	tcg	tcc	agc	agc	4525
Val	Pro	Leu	Tyr	Asp	Arq	Asn	His	Val	Thr	Gly	Ala	Ser	Ser	Ser	Ser	
1470			-	-	1475					1480					1485	
tca	tcc	agc	acq	aag	acc	acq	cta	tac	cca	cca	atc	cta	aac	cca	CCa	4573
				Lys												10.0
001	001			1490		****	200	-1-	1499		110	200	11011	1500		
CCC	tee	cca	acc	acg		ccc	too	cta			ata	asc.	ata			4621
				Thr												4021
FIO	Ser	FIU			vsb	FIO	Ser		_	M211	Met	ASP			IÀL	
			1505					1510					1519			
				ccg												4669
Ser	Ser			Pro	Ala	Thr		_	Pro	Tyr	Arg		-	Ile	Ile	
		1520					1525	_				1530				
				CCC												4717
Arg	Gly	Met	Ala	Pro	Pro	Thr	Thr	Pro	Cys	Ser	Thr	Asp	Val	Cys	Asp	
	1539	5				1540	)				1545	5		-	-	
agc	gac	tac	agc	gcc	agc	cgc	tgg	aag	gcc	agc	aag	tac	tac	ctg	gat	4765
Ser	Asp	Tyr	Ser	Ala	Ser	Arg	Trp	Lys	Ala	Ser	Lys	Tyr	Tyr	Leu	Asp	
1550	_	_			1555		-	-		1560		-	•		1565	
tta	aac	tca	gac	tca	gac	ccc	tat	cca	ccc	cca	ccc	acα	ccc	cac	agc	4813
				Ser												
				1570			-1-		1579			****		1580		
сад	tac	cta	tca	gcg		gac	age	tac			tca	CCC	acc		-	4861
				Ala												4001
GIII	ıyı	Leu	1585		GIU	ASP	Ser	1590		FIO	Ser	PIO	159		GIU	
														-		4000
				cat												4909
Arg	Ser	1600		His	Leu	Pne	1609		Pro	Pro	Ser	1610	_	Thr	Asp	
tca	tcc	tgad	ctc	ggc d	cggg	ccact	c to	ggcti	ctct	t gto	gece	ctgt	aaa	tagti	ttt	4965
	Ser	_						_				_		•		
·	161	5														
aaat			aagaa	1222	a ata	atati	tta	trat	tttas	222	aata:	aatai	ר א	taa	gattt	5025
																5085
										-yy (	Jecg	yaya	aa C	cceg	tacag	
tgga	gaaa	La CI	cata	aaacı	t taa	acct	tgta	aaa	ca							5120

<210> 3 <211> 1615 <212> PRT <213> Homo sapiens

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 10
 15

 Leu Leu Leu Leu Leu Leu Ala Leu Cys Gly Cys Pro Ala Pro Ala Ala Ala Ser 20
 25
 30

 Pro Leu Leu Leu Leu Phe Ala Asn Arg Arg Asp Val Arg Leu Val Asp Ala 35
 40
 45

 Gly Gly Val Lys Leu Glu Ser Thr Ile Val Val Ser Gly Leu Glu Asp

	50					55					60				
Ala 65	Ala	Ala	Val	Asp	Phe 70	Gln	Phe	Ser	Lys	Gly 75	Ala	Val	Tyr	Trp	Thr 80
Asp	Val	Ser	Glu	Glu 85	Ala	Ile	Lys	Gln	Thr 90	Tyr	Leu	Asn	Gln	Thr 95	Gly
			100					105					Pro 110	_	_
		115					120					125	Asp		
	130					135					140		Arg	_	
145		_		_	150	_				155			Leu	-	160
				165					170				Pro	175	
			180		_	_		185	_	_			Val 190	•	
		195	-				200			_		205	Glu		-
	210					215					220	_	Ala		
225					230	-				235			Thr		240
				245					250	_			Asp	255	
			260			_		265	-		_	_	Lys 270	_	_
		275				_	280					285	Val Asp		
	290					295					300		Phe		_
305	_				310					315			Arg	-	320
		-		325					330	_		_	Thr	335	_
		_	340					345				_	350 Val		
		355			_		360	_			_	365	Pro		
	370					375					380				Ala
385	_			_	390	_	-			395			Glu	_	400
_				405					410				Leu	415	
•		_	420				-	425					430 Asn		_
	-	435			_		440				_	445	Arg	_	
	450	_				455				_	460		Trp		
465	Den	1113	110	Val	470	GLY	Dea	Mec	ıyı	475	1111	тэр	пр	GLY	480
				485					490				Arg	495	
Leu	Val	Asn	Ala 500	Ser	Leu	Gly	Trp	Pro 505	Asn	Gly	Leu	Ala	Leu 510	Asp	Leu

Gln	Glu	Gly 515	Lys	Leu	Tyr	Trp	Gly 520	Asp	Ala	Lys	Thr	Asp 525	Lys	Ile	Glu
Val	Ile 530	Asn	Val	Asp	Gly	Thr 535	Lys	Arg	Arg	Thr	Leu 540	Leu	Glu	Asp	Lys
Leu 545	Pro	His	Ile	Phe	Gly 550	Phe	Thr	Leu	Leu	Gly 555	Asp	Phe	Ile	Tyr	Trp 560
Thr	Asp	Trp	Gln	Arg 565	Arg	Ser	Ile	Glu	Arg 570	Val	His	Lys	Val	Lys 575	Ala
Ser	Arg	Asp	Val 580	Ile	Tle	Asp	Gln	Leu 585	Pro	Asp	Leu	Met	Gly 590	Leu	Lys
Ala	Val	Asn 595	Val	Ala	Lys	Val	Val 600	Gly	Thr	Asn	Pro	Cys 605	Ala	Asp	Arg
	610	_	_		His	615	-				620				
625		_			Gly 630					635			_		640
				645	Phe				650		_			655	
			660		Thr			665					670		
		675			Ser		680			_		685			
	690				Val	695					700				
705	_				Glu 710					715	_		_	_	720
				725	Asp				730					735	_
			740		Ile			745	_			_	750		_
		755			Arg		760				_	765			
_	770			_	Tyr	775					780	_	_	_	
785					Phe 790			_		795	_				800
	_		-	805	Ala		_		810			_		815	
_		•	820		Asp		_	825					830		
		835			_		840			_	•	845			Pro
	850					855		_		_	860				Asn
865					870			_		875	_	_			Thr 880
				885					890	_				895	His
		_	900		_			905	_				910	_	Gln
	-	915			Leu		920					925	_	_	-
	930				Leu	935					940	_			
945					950			-		955			=		11e 960
rro	Asp	ASP	GID	uls	ser	PTO	ASP	ьеи	тте	Leu	Pro	Leu	HIS	GTÀ	Leu

	965			970		975	
Arg Asn Val	980		985	5		990	
Trp Val Asp 995		Gln Asn		Arg Ala	Lys Asp 100		Thr
Gln Pro Phe 1010		101	5	_	1020		•
Gln Pro His 1025		1030		103	5		1040
Thr Cys Glu	1045	5		1050		1055	5
Ala Met Gly	1060		106	55		1070	
Val Val Asn 107	5		1080	_	108	5	•
Arg Ala Ala 1090		109	5	_	1100	•	
Val Leu Phe 1105		1110		111	5		1120
Asn Thr Leu	1125	5	-	1130	-	1135	5
Glu Ser Cys	1140	_	114	5		1150	
Asn Ile Val 115	5		1160		116	5	
Trp Ile Asp 1170		117	5	_	1180	_	
Gly Asp Lys 1185		1190		119	5		1200
Ile His Ala	1205	5		1210		1215	5
Cys Ala Arg	1220		122	25		1230	_
Asp Gly Thr	5.		1240		124	5	
Asn Leu Leu 1250		125	5		1260		
Ala Cys Ala 1265		1270		127	5	_	1280
Asp Gly Phe	1285	5		1290		1295	5
Val Cys Ser	1300		130	)5	_	1310	_
Leu Arg Leu 131	5	_	1320		132	5	_
Glu Val Asp 1330		133	5		1340		
Ser Gly Gln 1345		1350		135	5		1360
Cys Ile Asp	1365	5		1370		1375	5
Ser Asp Asp	1380		138	15	-	1390	_
Ile Ile Leu 139	5		1400		140	5	
Arg Val Val		* · · · · · · · · · · · · · · · · · · ·	31 - 01	. 71- 7	Clar Dan	Dh - D	

### 032796-132.ST25

Glu Tyr Val Ser Gly Thr Pro His Val Pro Leu Asn Phe Ile Ala Pro 1430 1435 Gly Gly Ser Gln His Gly Pro Phe Thr Gly Ile Ala Cys Gly Lys Ser 1445 1450 Met Met Ser Ser Val Ser Leu Met Gly Gly Arg Gly Val Pro Leu 1465 Tyr Asp Arg Asn His Val Thr Gly Ala Ser Ser Ser Ser Ser Ser Ser 1475 1480 1485 Thr Lys Ala Thr Leu Tyr Pro Pro Ile Leu Asn Pro Pro Pro Ser Pro 1490 1495 1500 Ala Thr Asp Pro Ser Leu Tyr Asn Met Asp Met Phe Tyr Ser Ser Asn 1510 1515 Ile Pro Ala Thr Ala Arg Pro Tyr Arg Pro Tyr Ile Ile Arg Gly Met 1530 Ala Pro Pro Thr Thr Pro Cys Ser Thr Asp Val Cys Asp Ser Asp Tyr 1545 1550 Ser Ala Ser Arg Trp Lys Ala Ser Lys Tyr Tyr Leu Asp Leu Asn Ser 1560 1565 Asp Ser Asp Pro Tyr Pro Pro Pro Pro Thr Pro His Ser Gln Tyr Leu 1575 1580 Ser Ala Glu Asp Ser Cys Pro Pro Ser Pro Ala Thr Glu Arg Ser Tyr 1590 1595 Phe His Leu Phe Pro Pro Pro Pro Ser Pro Cys Thr Asp Ser Ser 1610

<210> 4 <211> 1615 <212> PRT <213> Homo sapiens

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Asp	Ile	Tyr 195	Trp	Pro	Asn	Gly	Leu 200	Thr	Ile	Asp	Leu	Glu 205	Glu	Gln	Lys
Leu	Tyr 210	Trp	Ala	Asp	Ala	Lys 215	Leu	Ser	Phe	Ile	His 220	Arg	Ala	Asn	Leu
Asp 225	Gly	Ser	Phe	Arg	Gln 230	Lys	Val	Val	Glu	Gly 235	Ser	Leu	Thr	His	Pro 240
Phe	Ala	Leu	Thr	Leu 245	Ser	Gly	Asp	Thr	Leu 250	Tyr	Trp	Thr	Asp	Trp 255	Gln
Thr	Arg	Ser	Ile 260	His	Ala	Cys	Asn	Lys 265	Arg	Thr	Gly	Gly	Lys 270	Arg	Lys
Glu	Ile	Leu 275	Ser	Ala	Leu	Tyr	Ser 280	Pro	Met	Asp	Ile	Gln 285	Val	Leu	Ser
Gln	Glu 290	Arg	Gln	Pro	Phe	Phe 295	His	Thr	Arg	Cys	Glu 300	Glu	Asp	Asn	Gly
305					Cys 310					<b>315</b>				_	320
				325	Gly				330			_	_	335	_
Lys	Ala	Gly	Ala 340	Glu	Glu	Val	Leu	Leu 345	Leu	Ala	Arg	Arg	Thr 350	Asp	Leu
_		355			Asp		360	_			_	365			
	370	_		_	His	375				-	380				
385			_	_	Thr 390	_	_			395			_	_	400
Tyr	Leu	Asp	Gly	Ser 405	Gly	Ala	Gln	Thr	Leu 410	Val	Asn	Thr	Glu	Ile 415	Asn
Asp	Pro	Asp	Gly 420	Ile	Ala	Val	Asp	Trp 425	Val	Ala	Arg	Asn	Leu 430	Tyr	Trp
		435	_		Asp	_	440				_	445			
	450	_			Val	455				-	460		_		
465					Met 470					475				_	480
		_		485	Cys				490	_			_	495	
			500		Leu	_		505					510	_	
		515			Tyr	_	520	_				525	-		
	530				Gly	535					540			_	_
545					Gly 550					555					560
		_		565	Arg				570			_		575	
			580		Ile			585					590		_
		595			Lys		600					605			
	610				His	615	_				620				
625					Gly 630					635	_		_		640
Ile	Val	Pro	Glu	Ala	Phe	Leu	Val	Phe	Thr	Ser	Arg	Ala	Ala	Ile	His

				645					650					655	
Arg	Ile	Ser	Leu 660	Glu	Thr	Asn	Asn	Asn 665	Asp	Val	Ala	Ile	Pro 670	Leu	Thr
Gly	Val	Lys 675	Glu	Ala	Ser	Ala	Leu 680	Asp	Phe	Asp	Val	Ser 685	Asn	Asn	His
Ile	Tyr 690	Trp	Thr	Asp	Val	Ser 695	Leu	Lys	Asn	Ile	Ser 700	Arg	Ala	Phe	Met
705				Val	710					715	_		_	_	720
				Val 725	_	•		_	730			-	•	735	-
			740	Arg				745					750		-
		755		Trp			760					765			
	770			Gly	_	775	_	_			780		_	_	
785				Ala	790					795					800
				Arg 805					810			_		815	
_		_	820	Thr	_		_	825					830		
		835		Glu			840			_	_	845			
	850			Gln		855					860				
865				Glu	870			_		875	_			_	880
				His 885		_			890					895	
			900	Asp	_			905	_				910	_	
		915		Cys			920					925		_	
	930		_	Thr		935				_	940	_			
945				Leu	950			_		955			_		960
	_	_		His 965					970					975	
			980	Ala		_	_	985			_	_	990		-
_		995	_	Arg			1000	ב ב	_		_	100	5		
	1010	)		Leu		1015	5				1020	כ		_	_
1025	5			Leu	103	)				103	5				1040
	_			Thr 1045	5				1050	)	_			105	ō
			1060					1069	5				1070	)	
		1075	5	Glu			1080	)				1089	5		
Arg	Ala 1090		rys	Ile	GLu	Arg 1095		Ala	Leu	Asp	Gly 1100		Glu	Arg	Glu

Val Le	ı Phe	Thr	Thr	Gly 111	Leu 0	Ile	Arg	Pro	Val 111:		Leu	Val	Val	Asp 1120
Asn Th	r Leu	Gly	Lys 112		Phe	Trp	Val	Asp 113		Asp	Leu	Lys	Arg	Ile
Glu Se	c Cys	Asp 114		Ser	Gly	Ala	Asn 1145		Leu	Thr	Leu	Glu 1150	Asp	
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Trp Ile	70				1175	5				1180	)	-		
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1425				1430	)				1435	5				1440
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420

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<210> 87

<211> 892

<212> PRT

<213> Homo sapiens

<400> 87

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Thr 305	Met	His	Ala	Met	Gln 310	Gln	Lys	Leu	Glu	Asp 315	Phe	Arg	Asp	Tyr	Arg 320
			Lys	325					330		_			335	
			Thr 340					345	_				350		
		355	Ser				360					365			_
	370		Glu			375					380				
385			Arg		390					395			-		400
			Ser	405					410			-		415	
			Lys 420					425					430	_	
		435	Lys				440			_		445			
	450		Glu			455					460				
465			Asp		470					475	_		-		480
			Asp	485					490			_	_	495	
			Thr 500					505			_		510	_	
		515	Lys	<u> </u>			520				_	525			
	530		Leu			535					540				
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			Glu	565					570					575	
			Gln 580					585			_		590		-
		595	Thr		•		600		_	_	•	605			
	610		Pro	_	_	615					620				_
625			Asn		630				•	635					640
			Pro	645					650				_	655	
			Met 660					665					670		_
		675	Lys				680	_				685	_		
	690		His			695					700		_		_
705			Tyr		710					715		_			720
Leu	Thr	Thr	Ile	Ala 725	Arg	Thr	Ile	Asn	Glu 730	Val	Glu	Asn	Gln	Ile 735	Leu
Thr	Arg	Asp	Ala	Lys	Gly	Ile	Ser	Gln	Glu	Gln	Met	Asn	Glu	Phe	Arg

745 Ala Ser Phe Asn His Phe Asp Arg Asp His Ser Gly Thr Leu Gly Pro 760 Glu Glu Phe Lys Ala Cys Leu Ile Ser Leu Gly Tyr Asp Ile Gly Asn · 780 775 Asp Pro Gln Gly Glu Ala Glu Phe Ala Arg Ile Met Ser Ile Val Asp 790 795 Pro Asn Arg Leu Gly Val Val Thr Phe Gln Ala Phe Ile Asp Phe Met 805 810 Ser Arg Glu Thr Ala Asp Thr Asp Thr Ala Asp Gln Val Met Ala Ser 825 Phe Lys Ile Leu Ala Gly Asp Lys Asn Tyr Ile Thr Met Asp Glu Leu 840 Arg Arg Glu Leu Pro Pro Asp Gln Ala Glu Tyr Cys Ile Ala Arg Met . 860 855 Ala Pro Tyr Thr Gly Pro Asp Ser Val Pro Gly Ala Leu Asp Tyr Met 870 875 Ser Phe Ser Thr Ala Leu Tyr Gly Glu Ser Asp Leu 885 <210> 88 <211> 197 <212> PRT

<213> Homo sapiens

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Gly Glu Lys Ser Asp 195

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<211> 739

<212> PRT

<213> Homo sapiens

PCT/US02/15982

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	Thr 50					55					60				-
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	Gly			85					90				_	95	
	Pro		100					105					110		
	Ser	115					120					125			
	Asn 130					135					140				
145	Thr				150					155					160
	Ala			165					170			_		175	-
	Val		180					185					190		
	Glu	195					200					205			_
	11e 210					215			_		220		_		
225	Thr				230					235					240
	Gln			245					250			_		255	-
	Asn		260					265			_		270	_	
	Gly	275					280					285			_
	Val 290					295					300		•		-
305	Arg				310					315				_	320
	Met			325					330				•	335	
	Arg		340					345					350		
	Asn	355					360		_	_		365		_	
	Tyr 370					375					380				
385	Ile				390					395					400
	Met			405					410					415	
	Pro		420					425					430		-
۲ne	Phe	Leu	Leu	ser	His	Glu	Val	Leu	Asn	Pro	Met	Tyr	Cys	Leu	Phe

435 440 445 Glu Tyr Ala Gly Lys Asp Asn Tyr Cys Leu Gln Ile Asn Pro Ala Ser 455 460 Tyr Ile Asn Pro Asp His Leu Lys Tyr Phe Arg. Phe Ile Gly Arg Phe 470 475 Ile Ala Met Ala Leu Phe His Gly Lys Phe Ile Asp Thr Gly Phe Ser 485 490 Leu Pro Phe Tyr Lys Arg Ile Leu Asn Lys Pro Val Gly Leu Lys Asp 500 505 Leu Glu Ser Ile Asp Pro Glu Phe Tyr Asn Ser Leu Ile Trp Val Lys 520 Glu Asn Asn Ile Glu Glu Cys Asp Leu Glu Met Tyr Phe Ser Val Asp 535 540 Lys Glu Ile Leu Gly Glu Ile Lys Ser His Asp Leu Lys Pro Asn Gly 550 555 Gly Asn Ile Leu Val Thr Glu Glu Asn Lys Glu Glu Tyr Ile Arg Met 565 570 Val Ala Glu Trp Arg Leu Ser Arg Gly Val Glu Glu Gln Thr Gln Ala 585 Phe Phe Glu Gly Phe Asn Glu Ile Leu Pro Gln Gln Tyr Leu Gln Tyr 600 Phe Asp Ala Lys Glu Leu Glu Val Leu Leu Cys Gly Met Gln Glu Ile 615 620 Asp Leu Asn Asp Trp Gln Arg His Ala Ile Tyr Arg His Tyr Ala Arg 630 635 Thr Ser Lys Gln Ile Met Trp Phe Trp Gln Phe Val Lys Glu Ile Asp 645 650 Asn Glu Lys Arg Met Arg Leu Leu Gln Phe Val Thr Gly Thr Cys Arg 665 Leu Pro Val Gly Gly Phe Ala Asp Leu Met Gly Ser Asn Gly Pro Gln 680 Lys Phe Cys Ile Glu Lys Val Gly Lys Glu Asn Trp Leu Pro Arg Ser 695 His Thr Cys Phe Asn Arg Leu Asp Leu Pro Pro Tyr Lys Ser Tyr Glu 710 715 Gln Leu Lys Glu Lys Leu Leu Phe Ala Ile Glu Glu Thr Glu Gly Phe Gly Gln Glu

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<212> PRT

<213> Homo sapiens

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                                            140
Gly Ser Pro Gly Ala Leu Ala Gly Ala Arg Val Gly Ala Ala Gly Pro
Leu Glu Arg Arg Gly Ala Gln Pro Gly Arg His Ser Val Thr Gly Tyr
                165
                                    170
Gly Asp Cys Ala Val Gly Ala Arg Tyr Gln Asp Glu Leu Thr Ala Leu
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Leu Arg Leu Thr Val Gly Thr Gly Gly Arg Glu Ala Gly Ala Arg Gly
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Glu Pro Ser Gly Ile Glu Pro Ser Gly Leu Glu Glu Pro Pro Gly Pro
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                                            220
Phe Val Pro Glu Ala Ala Arg Ala Arg Met Arg Glu Pro Glu Ala Arg
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Glu Asp Tyr Phe Gly Thr Cys Ile Lys Cys Asn Lys Gly Ile Tyr Gly
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Gln Ser Asn Ala Cys Gln Ala Leu Asp Ser Leu Tyr His Thr Gln Cys
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Phe Val Cys Cys Ser Cys Gly Arg Thr Leu Arg Cys Lys Ala Phe Tyr
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                                                285
Ser Val Asn Gly Ser Val Tyr Cys Glu Glu Asp Tyr Leu Phe Ser Gly
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                                            300
Phe Gln Glu Ala Ala Glu Lys Cys Cys Val Cys Gly His Leu Ile Leu
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                                        315
Glu Lys Ile Leu Gln Ala Met Gly Lys Ser Tyr His Pro Gly Cys Phe
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                                    330
Arg Cys Ile Val Cys Asn Lys Cys Leu Asp Gly Ile Pro Phe Thr Val
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Asp Phe Ser Asn Gln Val Tyr Cys Val Thr Asp Tyr His Lys Asn Tyr
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Ala Pro Lys Cys Ala Ala Cys Gly Gln Pro Ile Leu Pro Ser Glu Gly
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Cys Glu Asp Ile Val Arg Val Ile Ser Met Asp Arg Asp Tyr His Phe
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<212> PRT

<213> Homo sapiens

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Glu Glu Glu Glu Leu Val Ser Thr Asp Pro Arg Pro Ala Ser Tyr Ser		50					55					60				
Phe Cys         Ser Gly         Lys         Gly         Val         Gly         Ile         Lys         Gly         Glu         Thr         Ser         Thr         Ala         85         Ser         Asp         Leu         Gly         Tyr         Glu         Pro         Glu         Gly         Ser         Thr         Ala         Ser         Thr         Ala         Ser         Leu         Ala         Gly         Ile         Lys         Trp         Ala         Glu         Ser         Leu         Hean         Thr         Phe         Leu         Leu         His         Ser           Gln         Glu         Cys         Ala         Asp         Leu         Leu         Asp         Phe         Trp         Phe         Ala         Cys         Thr         Ile         Leu         Lys         Ile         Leu         Lys         Ile         Leu         Lys         Ile         Leu         Lys         Tyr         Ile         Leu         Lys         Tyr         Ile         Lu         Lys         Tyr         Ile         Lu         Lys         Ile         Lys         Tyr         Ile         Lys         Lys         Ile         Lys         Ile         Lys	Glu		Gly	Glu	Leu	Val		Thr	Asp	Pro	Arg		Ala	Ser	Tyr	Ser
The Pro Arg Arg Ser Asp Leu Asp Leu Lys Trp Ala Glu Ser Iteu Kis Ser 100	65		_			70			_		75				_	80
100	Phe	Cys	Ser	Gly		Gly	Val	Gly	Ile		Gly	Glu	Thr	Ser		Ala
115	Thr	Pro	Arg	_	Ser	Asp	Leu	Asp		Gly	Tyr	Glu	Pro		Gly	Ser
130	Ala	Ser		Thr	Pro	Pro	Tyr		Lys	Trp	Ala	Glu		Leu	His	Ser
145	Leu		Asp	Asp	Gln	Asp		Ile	Ser	Leu	Phe		Thr	Phe	Leu	Lys
Leu Ala Arg Ala Ile Tyr Arg Lys Tyr Ile Leu Asp Asn Asn Gly Ile		Glu	Gly	Суѕ	Ala		Leu	Leu	Asp	Phe		Phe	Ala	Cys	Thr	-
Nail   Ser	Phe	Arg	Lys	Leu		Pro	Cys	Asp	Ser		Glu	Glu	Lys	Arg		Lys
195	Leu	Ala	Arg		Ile	Tyr	Arg	Lys		Ile	Leu	Asp	Asn		Gly	Ile
210	Val	Ser		Gln	Thr	Lys	Pro		Thr	Lys	Ser	Phe		Lys	Gly	Cys
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Val Cys   Ser   Asp   Gln   Ser   Ser   Gly   Ser   Gly   Thr   Gly   Lys   Gly   Ile   Ser   265   265   270   275   275   285   285   270   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285   285	_	Ile	Gln	Ala	Thr		Glu	Glu	Asn	Thr	_	Pro	Ser	Phe	Leu	_
Gly Tyr Leu Pro Thr Leu Asn Glu Asp Glu Glu Trp Lys Cys Asp Gln 275	Ser	Asp	Ile	Tyr		Glu	Tyr	Thr	Arg		Gly	Ser	Glu	Ser		Lys
Asp   Met   Asp   Glu   Asp   Asp   Gly   Arg   Asp   Ala   Ala   Pro   Pro   Gly   Arg   Leu   295   Ser   Ser   300	Val	Cys	Ser		Gln	Ser	Ser	Gly		Gly	Thr	Gly	Lys		Ile	Ser
Pro   Gln   Lys   Leu   Leu   Leu   Glu   Thr   Ala   Ala   Pro   Arg   Val   Ser   Ser   Ser   305   310   310   315   320     Arg   Arg   Tyr   Ser   Glu   Gly   Arg   Glu   Phe   Arg   Tyr   Gly   Ser   Trp   Arg   Glu   325   330     Pro   Val   Asn   Pro   Tyr   Tyr   Val   Asn   Ala   Gly   Tyr   Ala   Leu   Ala   Pro   Ala   345   350     Thr   Ser   Ala   Asn   Asp   Ser   Glu   Gln   Gln   Ser   Leu   Ser   Ser   Asp   Ala   Asp   365     Thr   Leu   Ser   Leu   Thr   Asp   Ser   Ser   Val   Asp   Gly   Ile   Pro   Pro   Tyr   Arg   370   375   380     Ile   Arg   Lys   Gln   His   Arg   Arg   Glu   Met   Gln   Glu   Ser   Ala   Gln   Val   Asn   385   390   365     Glu   Arg   Val   Pro   Leu   Pro   His   Ile   Pro   Arg   Thr   Tyr   Arg   Val   Pro   Lys   Ala   Gln   Val   Asn   Ala   Glu   Asn   Asp   Ala   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu   Glu   Ala   Glu   Glu	Gly	Tyr		Pro	Thr	Leu	Asn		Asp	Glu	Glu	Trp	_	Cys	Asp	Gln
305	Asp		Asp	Glu	Asp	Asp		Arg	Asp	Ala	Ala		Pro	Gly	Arg	Leu
Pro Val Asn   Pro   Tyr   Tyr   Val Asn   Ala   Gly   Tyr   Ala   Leu   Ala   Pro   Ala   340   Ala   345   Ala   345   Ala   350   Ala   350   Ala   355   Ala   Asn   Asn   Asn   360   Asn   365   Asn   355   Asn   360   Ala   365   Ala   365   Ala   Asn   375   Ala   Asn   365   Ala   Asn   370   Ala   375   Ala   380   Ala   Asn   385   Ala   Asn   385   Ala   Asn   390   Ala   395   Ala   Asn   385   Ala   Ala		Gln	Lys	Leu	Leu		Glu	Thr	Ala	Ala		Arg	Val	Ser	Ser	
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Thr Leu Ser Leu Thr Asp Ser Ser Val Asp Gly Ile Pro Pro Tyr Arg 370				340	_	_			345	_	-			350		
370			355					360					365			
385		370					375					380				
Glu Val Arg Val Glu Pro Gln Lys Phe Ala Glu Glu Leu Ile His Arg 425	385					390					395					400
Leu Glu Ala Val Gln Arg Thr Arg Glu Ala Glu Glu Lys Leu Glu Glu 435  Arg Leu Lys Arg Val Arg Met Glu Glu Glu Glu Glu Asp Gly Asp Pro 450  Ser Ser Gly Pro Pro Gly Pro Cys His Lys Leu Pro Pro Ala Pro Ala 465  Trp His His Phe Pro Pro Arg Leu Cys Trp Thr Trp Ala Cys Ala Gly 485  Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His	_				405					410					415	
Arg Leu Lys Arg Val Arg Met Glu Glu Glu Glu Glu Asp Gly Asp Pro 450  Ser Ser Gly Pro Pro Gly Pro Cys His Lys Leu Pro Pro Ala Pro Ala 465  Trp His His Phe Pro Pro Arg Leu Cys Trp Thr Trp Ala Cys Ala Gly 485  Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His				420				_	425					430		_
450 455 460  Ser Ser Gly Pro Pro Gly Pro Cys His Lys Leu Pro Pro Ala Pro Ala 465 470 475 70 485  Trp His His Phe Pro Pro Arg Leu Cys Trp Thr Trp Ala Cys Ala Gly 485 490 70 495  Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His			435			_		440					445			
465 470 475 480  Trp His His Phe Pro Pro Arg Leu Cys Trp Thr Trp Ala Cys Ala Gly 485 490 . 495  Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His	_	450	-				455					460			_	
Trp His His Phe Pro Pro Arg Leu Cys Trp Thr Trp Ala Cys Ala Gly 485 490 . 495  Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His		Ser	Gly	Pro	Pro		Pro	Cys	His	Lys		Pro	Pro	Ala	Pro	
Leu Arg Asp Ala His Glu Glu Asn Pro Glu Ser Ile Leu Asp Glu His		His	His	Phe			Arg	Leu	Cys			Trp	Ala	Cys		
	Leu	Arg	Asp			Glu	Glu	Asn			Ser	Ile	Leu	-		His

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His Arg Ser Pro Asp Ser Gly His Val Ala Lys Met Pro Val Ala Leu
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Gly Gly Ala Ala Ser Gly His Gly Lys His Val Pro Lys Ser Gly Ala
                    550
Lys Leu Asp Ala Ala Gly Leu His His His Arg His Val His His His
                                    570
Val His His Ser Thr Ala Arg Pro Lys Glu Gln Val Glu Ala Glu Ala
                                585
Thr Arg Arg Ala Gln Ser Ser Phe Ala Trp Gly Leu Glu Pro His Ser
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His Gly Ala Arg Ser Arg Gly Tyr Ser Glu Ser Val Gly Ala Ala Pro
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                                            620
Asn Ala Ser Asp Gly Leu Ala His Ser Gly Lys Val Gly Val Ala Cys
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Lys Arg Asn Ala Lys Lys Ala Glu Ser Gly Lys Ser Ala Ser Thr Glu
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Val Pro Gly Ala Ser Glu Asp Ala Glu Lys Asn Gln Lys Ile Met Gln
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Trp Ile Ile Glu Gly Glu Lys Glu Ile Ser Arg His Arg Arg Thr Gly
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His Gly Ser Ser Gly Thr Arg Lys Pro Gln Pro His Glu Asn Ser Arg
                        695
Pro Leu Ser Leu Glu His Pro Trp Ala Gly Pro Gln Leu Arg Thr Ser
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                    710
Val Gln Pro Ser His Leu Phe Ile Gln Asp Pro Thr Met Pro Pro His
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                                    730
Pro Ala Pro Asn Pro Leu Thr Gln Leu Glu Glu Ala Arg Arg Arg Leu
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Glu Glu Glu Lys Arg Ala Ser Arg Ala Pro Ser Lys Gln Arg Tyr
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Val Gln Glu Val Met Arg Arg Gly Arg Ala Cys Val Arg Pro Ala Cys
                        775
                                            780
Ala Pro Val Leu His Val Val Pro Ala Val Ser Asp Met Glu Leu Ser
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Glu Thr Glu Thr Arg Ser Gln Arg Lys Val Gly Gly Ser Ala Gln
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Pro Tyr Arg Thr Leu Val Arg Gly Arg Ala Val Thr Leu Gly Gln Phe
                            840
Lys Glu Leu Leu Thr Lys Lys Gly Ser Tyr Arg Tyr Tyr Phe Lys Lys
                        855
                                            860
Val Ser Asp Glu Phe Asp Cys Gly Val Val Phe Glu Glu Val Arg Glu
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Glu Lys Val Asp
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<213> Homo sapiens
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			100		Gly			105				-	110		•
		115			Leu		120					125	_		
	130				Pro	135					140				
145					Val 150					155				_	160
				165	Leu				170					175	
			180		Ala			185					190		
		195			Gly		200					205			
	210				Lys	215					220		_		_
225					His 230					235					240
				245	Gln				250					255	
			260		Gln			265					270	_	-
		275			Ser		280					285			
	290				Met -	295		٠			300		_		_
305					Leu 310					315			_		320
				325	Glu			_	330		_		-	335	
			340		Glu			345					350		
		355			Tyr		360		_			365	_		
	370				Asn	375					380		_	_	
385					Lys 390					395					400
				405	Leu -				410				_	415	-
			420		Leu			425					430		
		435			Glu		440					445			_
Cys	Tyr	Trp	Arg	Ala	Tyr	Ala	Val	Gly	Asp	Val	Glu	Lys	Met.	Ala	Leu

#### 032796-132.ST25

455 Val Lys Leu Ala Lys Leu His Glu Gln Leu Thr Glu Ser Glu Gln Ala 470 475 Ala Gln Cys Tyr Ile Lys Tyr Ile Gln Asp Ile Tyr Ser Cys Gly Glu 485 490 Ile Val Glu His Leu Glu Glu Ser Thr Ala Phe Arg Tyr Leu Ala Gln 505 Tyr Tyr Phe Lys Cys Lys Leu Trp Asp Glu Ala Ser Thr Cys Ala Gln 520 Lys Cys Cys Ala Phe Asn Asp Thr Arg Glu Glu Gly Lys Ala Leu Leu 535 Arg Gln Ile Leu Gln Leu Arg Asn Gln Gly Glu Thr Pro Thr Thr Glu 550 Val Pro Ala Pro Phe Phe Leu Pro Ala Ser Leu Ser Ala Asn Asn Thr 570 Pro Thr Arg Arg Val Ser Pro Leu Asn Leu Ser Ser Val Thr Pro 585 <210> 93 <211> 914 <212> PRT <213> Homó sapiens <400> 93 Val Tyr Gln Val Leu Leu Val Gly Ser Thr Leu Leu Lys Glu Val Pro Ser Gly Leu Gln Leu Glu Gln Leu Pro Ser Gln Ser Leu Leu Thr His Ile Pro Thr Ala Gly Leu Pro Thr Ser Leu Gly Gly Gly Leu Pro Tyr 40 Cys His Gln Ala Trp Leu Asp Phe Arg Arg Arg Leu Glu Ala Leu Leu Gln Asn Cys Gln Ala Ala Cys Ala Leu Leu Gln Gly Ala Ile Glu Ser Val Lys Ala Val Pro Gln Pro Met Glu Pro Gly Glu Val Gly Gln Leu 90 Leu Gln Gln Thr Glu Val Leu Met Gln Gln Val Leu Asp Ser Pro Trp 100 105 Leu Ala Trp Leu Gln Cys Gln Gly Gly Arg Glu Leu Thr Trp Leu Lys 120 Gln Glu Val Pro Glu Val Thr Leu Ser Pro Asp Tyr Arg Thr Ala Met 135 140 Asp Lys Ala Asp Glu Leu Tyr Asp Arg Val Asp Gly Leu Leu His Gln 150 Leu Thr Leu Gln Ser Asn Gln Arg Ile Gln Ala Leu Glu Leu Val Gln 165 170 Thr Leu Glu Ala Arg Glu Ser Gly Leu His Gln Ile Glu Val Trp Leu 185 Gln Gln Val Gly Trp Pro Ala Leu Glu Glu Ala Gly Glu Pro Ser Leu 200 Asp Met Leu Leu Gln Ala Gln Gly Ser Phe Gln Glu Leu Tyr Gln Val 215 220 Ala Gln Glu Gln Val Arg Gln Gly Glu Lys Phe Leu Gln Pro Leu Thr 230 235 Gly Trp Glu Ala Ala Glu Leu Asp Pro Pro Gly Ala Arg Phe Leu Ala 245 250 Leu Arg Ala Gln Leu Thr Glu Phe Ser Arg Ala Leu Ala Gln Arg Cys

			260					0.55							
C1 =	7	Lou	260	7	<b>71</b> -	C1	<b>7</b>	265	D	C1-	T	nh.	270	C1	
GIII	Arg	275	пта	Asp	Ата	Giu	280	Leu	rne	GIN	Leu	285	Arg	GIU	ATA
Leu	Thr 290		Ala	Glu	Glu	Gly 295		Arg	Val	Leu	Ala 300		Leu	Glu	Gln
Glu 305	Arg	Pro	Gly	Val	Val 310	Leu	Gln	Gln	Leu	Gln 315		His	Trp	Thr	Arg 320
	Pro	Asp	Leu	Pro 325	Pro	Ala	His	Phe	Arg 330		Met	Trp	Ala	Leu 335	
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		355		Asp			360					365			
Ser	Leu 370	Lys	Leu	Pro	Pro	Val 375	Gly	Ser	Thr	Ala	Ser 380	Leu	Cys	Val	Ser
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				Leu 405					410				_	415	_
			420	Thr				425					430	_	_
		435		Gln			440					445	_		
	450			Leu		455					460				
465				Glu	470					475					480
				Leu 485					490					495	_
			500	Leu				505				_	510		
		515		Leu			520					525			
	530			Ala		535	_		_		540				_
545				Lys	550					555					560
			•	Phe 565		_	_	_	570				_	575	
			580	Ser				585					590	_	_
_		595		Leu			600		_		_	605	_		
	610			Ala		615					620				
625	Arg	HIS	GIĀ	Asn	630	Leu	ren	ALG	met	635	ATa	TIE	GIN	GIA	Cys 640
•	Val	Asn	Leu	Lys 645		Gln	Gly	Gln	Leu 650		Arg	Gln	Asp	Glu 655	
Val	Val	Arg	Thr 660	Gly	Arg	His	Lys	Ser 665	Val	Arg	Arg	Ile	Phe 670	Leu	Phe
		675		Leu			680		_		_	685		_	
	690			Tyr		695					700				
Thr 705	Glu	Cys	Cys	Gly	Asn 710	Ser	Asn	Leu	Arg	Phe 715	Glu	Ile	Trp	Phe	Arg 720

Arg Arg Lys Ala Arg Asp Thr Phe Val Leu Gln Ala Ser Ser Leu Ala 730 Ile Lys Gln Ala Trp Thr Ala Asp Ile Ser His Leu Leu Trp Arg Gln 740 745 Ala Val His Asn Lys Glu Val Arg Met Ala Glu Met Val Ser Met Gly 760 Val Gly Asn Lys Ala Phe Arg Asp Ile Ala Pro Ser Glu Glu Ala Ile 775 Asn Asp Arg Thr Val Asn Tyr Val Leu Lys Cys Arg Glu Val Arg Ser 790 795 Arg Ala Ser Ile Ala Val Ala Pro Phe Asp His Asp Ser Leu Tyr Leu 810 Gly Ala Ser Asn Ser Leu Pro Gly Asp Pro Ala Ser Cys Ser Val Leu 825 Gly Ser Leu Asn Leu His Leu Tyr Arg Asp Pro Ala Leu Leu Gly Leu 840 Arg Cys Pro Leu Tyr Pro Ser Phe Leu Glu Glu Ala Ala Leu Glu Ala 855 860 Glu Ala Glu Leu Gly Gly Gln Pro Ser Leu Thr Ala Glu Asp Ser Glu 870 875 Ile Ser Ser Gln Cys Pro Ser Ala Ser Gly Ser Ser Gly Ser Asp Ser 885 890 Ser Cys Val Ser Gly Gln Ala Leu Gly Arg Gly Leu Glu Asp Leu Pro 905 Cys Val

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<212> PRT

<213> Homo sapiens

#### <400> 94

Leu Asn Tyr Leu Leu Glu Ser Arg Leu Glu Ala Ala Ala His Cys Ala 10 Leu Lys Gln Gly Ile Ala Thr Ala Ser Leu Leu Pro Ala Gln Leu Gln 25 Pro Ala Val Leu Thr Val Val Thr Cys His Val Val Val Ser Val His 40 Gly His His Thr Asp Gly Cys Leu Ala Ala Leu Cys Arg Glu Asp Arg 55 Thr Gly Thr Gly Gly Ala Phe Trp Cys Lys Asn Arg Val Ile Val Ser 70 His Ala Val Asp Val Val Leu His Val His Gly Glu Gly Asn Pro Val Gln Ala Leu Ile Ala His Gly Ala Pro Glu Ala Ala Trp Val Val Gly 105 Leu Ala Gln Gly Leu Gln Asp His Phe His Asp Glu Met Ser Thr His 120 Ala Ala Phe Val Gly Arg Leu Leu Glu Pro Gly Val Gln Glu Val Leu 135 140 Leu Ala Val His Phe Leu Thr His Val Val Glu Arg Leu Pro Thr Glu 150 155 Ser Ser Pro Thr Arg Val Ala Gly Glu Ala Val Ser Val Ile Lys Thr 165 170 Pro His Cys Leu Ala Arg Leu Leu Gly Ser Val Asp Ala Lys Pro Thr 180 185

032796-132.ST25 Leu Asp Ala Asn Ala Glu Val Val Pro Arg Arg Ala Arg Leu Glu Arg 195 200 Pro Leu Gln Leu Pro Gly Glu Arg Leu Gln Pro Pro Leu Gly Arg Ala 215 Trp Ala Ala Leu Pro Ala Arg Gly Gln Arg Glu Cys Arg Gln Arg Glu 230 Gly Gly Arg Pro Arg Arg Leu Arg Gly Ala Ser Gly Arg Gly Ala Gly 250 Ala Gly Arg Glu Glu Val Ser Val Gly Phe Ser Ala Gln Trp Glu Phe 265 Gly Ser Gly Arg His 275 <210> 95 <211> 1120 <212> PRT <213> Homo sapiens <400> 95 Met Trp Arg Val Lys Lys Leu Ser Leu Ser Leu Ser Pro Ser Pro Gln Thr Gly Lys Pro Ser Met Arg Thr Pro Leu Arg Glu Leu Thr Leu Gln 25 Pro Gly Ala Leu Thr Thr Ser Gly Lys Arg Ser Pro Ala Cys Ser Ser Leu Thr Pro Ser Leu Cys Lys Leu Gly Leu Gln Glu Gly Ser Asn Asn 55 Ser Ser Pro Val Asp Phe Val Asn Asn Lys Arg Thr Asp Leu Ser Ser 70 Glu His Phe Ser His Ser Ser Lys Trp Leu Glu Thr Cys Gln His Glu 90 Ser Asp Glu Gln Pro Leu Asp Pro Ile Pro Gln Ile Ser Ser Thr Pro 105 Lys Thr Ser Glu Glu Ala Val Asp Pro Leu Gly Asn Tyr Met Val Lys 120 Thr Ile Val Leu Val Pro Ser Pro Leu Gly Gln Gln Asp Met Ile 135 140 Phe Glu Ala Arg Leu Asp Thr Met Ala Glu Thr Asn Ser Ile Ser Leu 150 155 Asn Gly Pro Leu Arg Thr Asp Asp Leu Val Arg Glu Glu Val Ala Pro 170 Cys Met Gly Asp Arg Phe Ser Glu Val Ala Ala Val Ser Glu Lys Pro 185

 Ile Phe Gln Glu Ser Pro 195
 Ser His Leu Leu Glu Glu Glu Ser Pro 205
 Pro Pro Asn 205

 Pro Cys Ser Glu Gln Leu His Cys Ser Lys Glu Ser Leu Ser Ser Arg 210
 215
 220

 Thr Glu Ala Val Arg Glu Asp Leu Val Pro Ser Glu Ser Asn Ala Phe 225
 230
 235

 Leu Pro Ser Ser Val Leu Trp Leu Ser Pro Ser Thr Ala Leu Ala Ala 245
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 Asp Phe Arg Val Asn His Val Asp Pro Glu Glu Glu Glu Glu His 260
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 Gly Ala Met Glu Glu Glu Arg Glu Met Arg Phe Pro Thr His Pro Lys Glu 285

Ser Glu Thr Glu Asp Gln Ala Leu Val Ser Ser Val Glu Asp Ile Leu 290 295 300

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Pro	Gly	Pro	Ala	Val 325	Glu	Asp	Val	Gly	Arg 330	Ile	Leu	Gly	Ser	Asp 335	Thr
Glu	Ser	Trp	Met 340	Ser	Pro	Leu	Ala	Trp 345	Leu	Glu	Lys	Gly	Val 350	Asn	Thr
Ser	Val	Met 355	Leu	Glu	Asn	Leu	Arg 360	Gln	Ser	Leu	Ser	Leu 365	Pro	Ser	Met
Leu	Arg 370		Ala	Ala	Ile	Gly 375	Thr	Thr	Pro	Phe	Ser 380	Thr	Cys	Ser	Val
Gly 385	Thr	Trp	Phe	Thr	Pro 390	Ser	Ala	Pro	Gln	Glu 395	Lys	Ser	Thr	Asn	Thr 400
Ser	Gln	Thr	Gly	Leu 405	Val	Gly	Thr	Lys	His 410	Ser	Thr	Ser	Glu	Thr 415	
Gln	Leu	Leu	Cys 420	Gly	Arg	Pro	Pro	Asp 425	Leu	Thr	Ala	Leu	Ser 430	Arg	His
Asp	Leu	Glu 435	Asp	Asn	Leu	Leu	Ser 440	Ser	Leu	Val	Ile	Val 445	Glu	Phe	Leu
	450					455					460	Val			
465					470					475		His		_	480
				485					490			Met	_	495	
			500					505				Leu	510		
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	530					535					540	Thr			_
545					550					555	_	Leu			560
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	690					695					700	Leu			-
705					710					715		Arg			720
				725					730			Asp		735	
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Lys	Asp	Glu	Leu	Leu	Cys	Gln	Leu	Thr	Gln	Ser	Asn	Glu	Glu	Gln	Ala

765

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760

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Ala Gln Cys Val Lys Glu Glu Met Ala Leu Lys His Met Gln Ala Glu
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Leu Gln Gln Gln Ala Val Leu Ala Lys Glu Val Arg Asp Leu Lys
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                                         795
                                                             800
Glu Thr Leu Glu Phe Ala Asp Gln Glu Asn Gln Val Ala His Leu Glu
                                    810
Leu Gly Gln Val Glu Cys Gln Leu Lys Thr Thr Leu Glu Val Leu Arg
                                825
Glu Arg Ser Leu Gln Cys Glu Asn Leu Lys Asp Thr Val Glu Asn Leu
                            840
Thr Ala Lys Leu Ala Ser Thr Ile Ala Asp Asn Gln Glu Gln Asp Leu
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Glu Lys Thr Arg Gln Tyr Ser Gln Lys Leu Gly Leu Leu Thr Glu Gln
                    870
                                        875
Leu Gln Ser Leu Thr Leu Phe Leu Gln Thr Lys Leu Lys Glu Lys Thr
                885
                                    890
Glu Gln Glu Thr Leu Leu Leu Ser Thr Ala Cys Pro Pro Thr Gln Glu
            900
                                905
His Pro Leu Pro Asn Asp Arg Thr Phe Leu Gly Ser Ile Leu Thr Ala
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Val Ala Asp Glu Glu Pro Glu Ser Thr Pro Val Pro Leu Leu Gly Ser
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                                            940
Asp Lys Ser Ala Phe Thr Arg Val Ala Ser Met Val Ser Leu Gln Pro
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                                        955
Ala Glu Thr Pro Gly Met Glu Glu Ser Leu Ala Glu Met Ser Ile Met
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                                    970
Thr Thr Glu Leu Gln Ser Leu Cys Ser Leu Leu Gln Glu Ser Lys Glu
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Glu Ala Ile Arg Thr Leu Gln Arg Lys Ile Cys Glu Leu Gln Ala Arg
                            1000
Leu Gln Ala Gln Glu Gln His Gln Glu Val Gln Lys Ala Lys Glu
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                                            1020
Ala Asp Ile Glu Lys Leu Asn Gln Ala Leu Cys Leu Arg Tyr Lys Asn
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Glu Lys Glu Leu Gln Glu Val Ile Gln Gln Asn Glu Lys Ile Leu Glu
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                                   1050
Gln Ile Asp Lys Ser Gly Glu Leu Ile Ser Leu Arg Glu Glu Val Thr
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                                1065
His Leu Thr Arg Ser Leu Arg Arg Ala Glu Thr Glu Thr Lys Val Leu
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                                                1085
Gln Glu Ala Trp Gln Ala Ser Trp Thr Pro Thr Ala Ser Leu Trp Pro
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Leu Arg Pro Glu His Phe Gln Glu Val Gly Tyr Ala Ala Pro Pro Ser
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	50					55					Pro 60				
His 65	Gly	Pro	Pro	Phe	Glu 70	Gly	Gln	Ser	Gln	Val 75	Gln	Pro	Pro	Pro	Ser 80
Gln	Glu	Ala	Thr	Pro 85	Leu	Gln	Gln	Glu	Lys 90	Leu	Leu	Pro	Ala	Gln 95	Leu
Pro	Ala	Glu	Lys 100	Glu	Val	Gly	Pro	Pro 105	Leu	Pro	Gln	Glu	Ala 110	Val	Pro
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	130	•				135					Pro 140				
145					150					155	Ser			_	160
				165					170		Pro			175	
			180					185			Val		190	-	
		195					200				Thr	205		_	
	210					215					Cys 220	_		-	-
225					230					235	Val	-			240
				245					250		Lys		_	255	
			260					265			Ser		270		
		275					280				Cys	285		•	
	290					295					Pro 300		_		
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				325					330		Gln	_		335	
			340					345			Cys	_	350		_
		355					360				Asp	365			
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385					390					395	Phe				400
				405					410		Ile			415	_
			420					425			Asn		430		
		435					440				Asn	445			Ī
	450					455					Cys 460				
465					470					475	Arg				480
Arg	Asp	Pro	ALA	Leu 485	Cys	Cys	Tyr	Leu	Ser 490	Pro	Gly	Asp	Glu	Gln 495	Val

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 Ile
 Ser
 Ala
 Gly
 Tyr
 Ala
 Pro
 Val
 Leu
 Asp
 Cys
 His
 Thr
 Ala
 His
 Ile
 365
 Ala
 His
 Ile
 365
 Ala
 His
 Ile
 365
 Ala
 His
 Ile
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 Ala
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 Ile
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 Arg
 Ala
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<210> 98 <211> 2328 <212> PRT

<213> Homo sapiens

<400> 98

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Thr	Gln	Thr	Tyr	Gly 325	Gly	Asn	Leu	Asn	Gly 330	Glu	Pro	Cys	Val	Leu 335	Pro
Phe	Thr	Tyr	Asn 340	Gly	Arg	Thr	Phe	Tyr 345	Ser	Cys	Thr	Thr	Glu 350	Gly	Arg
Gln	Asp	Gly 355	His	Leu	Trp	Cys	Ser 360	Thr	Thr	Ser	Asn	Tyr 365	Glu	Gln	Asp
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				405	Asp				410				-	415	
			420		Thr			425					430		_
		435			Ala		440					445			_
	450				Gly	455	•				460		_		_
465					Thr 470					475		_		-	480
				485	Gln				490					495	
			500		Asp			505					510	_	
		515			Cys		520					525	_	_	
	530				Cys	535					540			•	
545					Glu 550					555		_	_		560
				565	Gly				570					575	
			580		Ser			585					590		
		595			Ser		600					605			
	610				Tyr	615					620				
625					Ala 630					635				_	640
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		675			Pro		680					685			
	690				Leu	695					700				
705					Val 710					715					720
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Gln	Tyr	Leu	Asp	Leu	Pro	Ser	Thr	Ala	Thr	Ser	Val	Asn	Ile	Pro	Asp

				740					745					750		
τ.	.011	ī.en	Pro		Dr.	T	π∽	Tla	745	700	Val	Ψ~	Gln	750	C	G1
			755					760					765			
		770					775					780	Thr			
7	85					790					795		Asp			800
					805					810			Gly	_	815	
				820					825				Leu	830		
			835					840					Pro 845	_		
		850					855					860	Glu			
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					885					890			Asp		895	
				900					905				Gly	910	_	
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		930					935					940	Ser			
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					965					970			Pro		975	
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		1010	)				1015	5				1020				-
1	025	5				1030	)				1035	5	Val			1040
					1045	5				1050	)		Gly		1055	5
				1060	)				1065	5			Thr	1070	)	
			1075	5				1080	)				Arg 1089	5	-	
		1090	)				1095	5		_		1100				
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				•	1125	5				1130	0		Gln		1135	5
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			1155	5				116	)				Thr 1169	5		_
G	lu	Arg 1170		Thr	Thr	Pro	Asp 1179		Thr	Gly	Tyr	Arg 1180	Ile		Thr	Thr
	ro 185		Asn	Gly	Gln	Gln 1190		Asn	Ser	Leu	Glu 1199		Val	Val	His	Ala 1200

Asp Gln Ser Ser Cys Thr Phe Asp Asn Leu Ser Pro Gly Leu Glu Tyr Asn Val Ser Val Tyr Thr Val Lys Asp Asp Lys Glu Ser Val Pro Ile Ser Asp Thr Ile Ile Pro Ala Val Pro Pro Pro Thr Asp Leu Arg Phe Thr Asn Ile Gly Pro Asp Thr Met Arg Val Thr Trp Ala Pro Pro Pro 1250 1255 Ser Ile Asp Leu Thr Asn Phe Leu Val Arg Tyr Ser Pro Val Lys Asn Glu Glu Asp Val Ala Glu Leu Ser Ile Ser Pro Ser Asp Asn Ala Val Val Leu Thr Asn Leu Leu Pro Gly Thr Glu Tyr Val Val Ser Val Ser Ser Val Tyr Glu Gln His Glu Ser Thr Pro Leu Arg Gly Arg Gln Lys Thr Gly Leu Asp Ser Pro Thr Gly Ile Asp Phe Ser Asp Ile Thr Ala Asn Ser Phe Thr Val His Trp Ile Ala Pro Arg Ala Thr Ile Thr Gly Tyr Arg Ile Arg His His Pro Glu His Phe Ser Gly Arg Pro Arg Glu Asp Arg Val Pro His Ser Arg Asn Ser Ile Thr Leu Thr Asn Leu Thr Pro Gly Thr Glu Tyr Val Val Ser Ile Val Ala Leu Asn Gly Arg Glu Glu Ser Pro Leu Leu Ile Gly Gln Gln Ser Thr Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr Tyr Arg Ile Thr Tyr Gly 1455 · Glu Thr Gly Gly Asn Ser Pro Val Gln Glu Phe Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys Pro Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly Asp Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu Ile Asp Lys Pro Ser Gln Met Gln Val Thr Asp Val Gln Asp Asn Ser Ile Ser Val Lys Trp Leu Pro Ser Ser Pro Val Thr Gly Tyr Arg Val Thr Thr Pro Lys Asn Gly Pro Gly Pro Thr Lys Thr Lys Thr Ala Gly Pro Asp Gln Thr Glu Met Thr Ile Glu Gly Leu Gln Pro Thr Val Glu Tyr Val Val Ser Val Tyr Ala Gln Asn Pro Ser Gly Glu Ser Gln Pro Leu Val Gln Thr Ala Val Thr Asn Ile Asp Arg Pro Lys Gly Leu Ala Phe Thr Asp Val Asp Val Asp Ser Ile Lys Ile Ala Trp Glu Ser Pro Gln Gly Gln Val Ser Arg Tyr Arg Val Thr Tyr Ser Ser Pro Glu Asp Gly Ile His Glu Leu Phe Pro Ala Pro Asp Gly Glu Glu Asp Thr Ala Glu Leu Gln Gly

	165	-				165					166	0			
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				1685	5				1690					169	5
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		171	5				1720	)		Thr		1725	5		_
	173	0				1735	5 .			Lys	1740	)			
174	5				1750	)				Leu 1755	•				1760
				1765	5				1770					1775	5
			1780	)				1785	5	Val		•	1790	) _	_
		1799	5				1800	)		Ile		1805	<b>,</b>	-	-
	1810	)				1815	5			Val	1820	)			
1825	5				1830	)				Lys 1835	i			_	1840
				1845	•				1850					1855	<b>;</b>
			1860	)				1865	<b>,</b>	Pro			1870	)	
		1875	5				1880	)		Arg		1885	1		
	1890					1895	•			Pro	1900		Arg	Ile	Thr
	Tvr	Ile													
1905	5				1910	)				1915	,	Pro	_		1920
Val	Pro	Arg	Pro	Arg 1925	1910 Pro	Gly	Val	Thr	Glu 1930	1915 Ala )	Thr	Ile	Thr	Gly 1935	1920 Leu
Val Glu	Pro Pro	Arg Gly	Pro Thr 1940	Arg 1925 Glu	1910 Pro Tyr	Gly Thr	Val Ile	Thr Tyr 1945	Glu 1930 Val	1915 Ala ) Ile	Thr Ala	Ile Leu	Thr Lys 1950	Gly 1935 Asn	1920 Leu Asn
Val Glu Gln	Pro Pro Lys	Arg Gly Ser 1955	Pro Thr 1940 Glu	Arg 1925 Glu Pro	1910 Pro Tyr Leu	Gly Thr	Val Ile Gly 1960	Thr Tyr 1945 Arg	Glu 1930 Val Lys	1915 Ala ) Ile Lys	Thr Ala Thr	Ile Leu Asp 1965	Thr Lys 1950 Glu	Gly 1935 Asn Leu	1920 Leu Asn Pro
Val Glu Gln Gln	Pro Pro Lys Leu 1970	Arg Gly Ser 1955 Val	Pro Thr 1940 Glu i	Arg 1925 Glu Pro Leu	1910 Pro Tyr Leu	Gly Thr Ile His 1975	Val Ile Gly 1960 Pro	Thr Tyr 1945 Arg Asn	Glu 1930 Val Lys Leu	1915 Ala Ile Lys His	Thr Ala Thr Gly 1980	Ile Leu Asp 1965 Pro	Thr Lys 1950 Glu Glu	Gly 1935 Asn Leu Ile	1920 Leu Asn Pro
Val Glu Gln Gln Asp 1985	Pro Pro Lys Leu 1970 Val	Gly Ser 1955 Val	Pro Thr 1940 Glu Thr Ser	Arg 1925 Glu Pro Leu	1910 Pro Tyr Leu Pro Val	Gly Thr Ile His 1975	Val Ile Gly 1960 Pro Lys	Thr Tyr 1945 Arg Asn Thr	Glu 1930 Val Lys Leu Pro	1915 Ala Ile Lys His Phe 1995	Thr Ala Thr Gly 1980 Val	Ile Leu Asp 1965 Pro	Thr Lys 1950 Glu Glu His	Gly 1935 Asn Leu Ile	1920 Leu Asn Pro Leu Gly 2000
Val Glu Gln Gln Asp 1985 Tyr	Pro Pro Lys Leu 1970 Val	Gly Ser 1955 Val Pro	Thr 1940 Glu Thr Ser	Arg 1925 Glu Pro Leu Thr Asn 2005	Tyr Leu Pro Val 1990 Gly	Gly Thr Ile His 1975 Gln Ile	Val Ile Gly 1960 Pro Lys Gln	Thr Tyr 1945 Arg Asn Thr	Glu 1930 Val Lys Leu Pro 2010	1915 Ala Ile Lys His Phe 1995 Gly	Thr Ala Thr Gly 1980 Val	Ile Leu Asp 1965 Pro Thr	Thr Lys 1950 Glu Glu His	Gly 1935 Asn Leu Ile Pro Gln 2015	1920 Leu Asn Pro Leu Gly 2000 Gln
Val Glu Gln Gln Asp 1985 Tyr	Pro Pro Lys Leu 1970 Val Asp	Gly Ser 1955 Val Pro Thr	Thr 1940 Glu Thr Ser Gly Gly 2020	Arg 1925 Glu Pro Leu Thr Asn 2005 Gln	Tyr Leu Pro Val 1990 Gly	Gly Thr Ile His 1975 Gln Ile Met	Val Ile Gly 1960 Pro Lys Gln Ile	Thr 1945 Arg Asn Thr Leu Phe 2025	Glu 1930 Val Lys Leu Pro 2010 Glu	1915 Ala Ile Lys His Phe 1995 Gly	Thr Ala Thr Gly 1980 Val Thr	Ile Leu Asp 1965 Pro Thr Ser	Thr Lys 1950 Glu Glu His Gly Phe 2030	Gly 1935 Asn Leu Ile Pro Gln 2015 Arg	1920 Leu Asn Pro Leu Gly 2000 Gln Arg
Val Glu Gln Gln Asp 1985 Tyr Pro	Pro Pro Lys Leu 1970 Val Asp Ser	Arg Gly Ser 1955 Val Pro Thr Val Pro 2035	Thr 1940 Glu Thr Ser Gly Gly 2020 Pro	Arg 1925 Glu Pro Leu Thr Asn 2005 Gln	Tyr Leu Pro Val 1990 Gly Gln	Gly Thr Ile His 1975 Gln Ile Met Ala	Val Ile Gly 1960 Pro Lys Gln Ile Thr 2040	Thr 1945 Arg Asn Thr Leu Phe 2025 Pro	Glu 1930 Val Lys Leu Pro 2010 Glu	1915 Ala Ile Lys His Phe 1995 Gly Glu Arg	Thr Ala Thr Gly 1980 Val Thr His	Ile Leu Asp 1965 Pro Thr Ser Gly Arg 2045	Thr Lys 1950 Glu Glu His Gly Phe 2030 Pro	Gly 1935 Asn Leu Ile Pro Gln 2015 Arg	Asn Pro Leu Gly 2000 Gln Arg
Val Glu Gln Gln Asp 1985 Tyr Pro Thr	Pro Pro Lys Leu 1970 Val Asp Ser Thr	Arg Gly Ser 1955 Val Pro Thr Val Pro 2035 Pro	Thr 1940 Glu Thr Ser Gly 2020 Pro	Arg 1925 Glu Pro Leu Thr Asn 2005 Gln Thr	Tyr Leu Pro Val 1990 Gly Gln Thr	Gly Thr Ile His 1975 Gln Ile Met Ala Gln 2055	Val Ile Gly 1960 Pro Lys Gln Ile Thr 2040 Glu	Thr 1945 Arg Asn Thr Leu Phe 2025 Pro	Glu 1930 Val Lys Leu Pro 2010 Glu Ile	IP15 Ala Ile Lys His Phe 1995 Gly Glu Arg	Thr Ala Thr Gly 1980 Val Thr His Gln 2060	Ile Leu Asp 1965 Pro Thr Ser Gly Arg 2045 Thr	Thr Lys 1950 Glu Glu His Gly Phe 2030 Pro	Gly 1935 Asn Leu Ile Pro Gln 2015 Arg	1920 Leu Asn Pro Leu Gly 2000 Gln Arg Pro
Val Glu Gln Gln Asp 1985 Tyr Pro Thr Tyr Trp 2065	Pro Pro Lys Leu 1970 Val Asp Ser Thr Pro 2050 Ala	Ser 1955 Val Pro Thr Val Pro 2035 Pro	Thr 1940 Glu Thr Ser Gly 2020 Pro Asn	Arg 1925 Glu Pro Leu Thr 2005 Gln Thr Val	Tyr Leu Pro Val 1990 Gly Gln Thr Gly Asp	Gly Thr Ile His 1975 Gln Ile Met Ala Gln 2055	Val Ile Gly 1960 Pro Lys Gln Ile Thr 2040 Glu Ser	Thr 1945 Arg Asn Thr Leu Phe 2025 Pro	Glu 1930 Val Lys Leu Pro 2010 Glu Ile Leu Tyr	IP15 Ala Ile Lys His Phe 1995 Gly Glu Arg Ser Ile 2075	Thr Ala Thr Gly 1980 Val Thr His Gln 2060 Ile	Ile Leu Asp 1965 Pro Thr Ser Gly Arg 2045 Thr	Thr Lys 1950 Glu Glu His Gly Phe 2030 Pro Thr	Gly 1935 Asn Leu Ile Pro Gln 2015 Arg Arg	1920 Leu Asn Pro Leu Gly 2000 Gln Arg Pro Ser Pro 2080
Glu Gln Gln Asp 1985 Tyr Pro Thr Tyr Trp 2065 Val	Pro Pro Lys Leu 1970 Val Asp Ser Thr Pro 2050 Ala	Arg Gly Ser 1955 Val Pro Thr Val Pro 2035 Pro Pro	Thr 1940 Glu Thr Ser Gly 2020 Pro Asn Phe	Arg 1925 Glu Pro Leu Thr Asn 2005 Gln Val Gln Glu 2085	Tyr Leu Pro Val 1990 Gly Gln Thr Gly Asp 2070 Glu	Gly Thr Ile His 1975 Gln Ile Met Ala Gln 2055 Thr	Val Ile Gly 1960 Pro Lys Gln Ile Thr 2040 Glu Ser Leu	Thr Tyr 1945 Arg Asn Thr Leu Phe 2025 Pro Ala Glu Gln	Glu 1930 Val Lys Leu Pro 2010 Glu Ile Leu Tyr Phe 2090	IP15 Ala Ile Lys His Phe 1995 Gly Glu Arg Ser Ile 2075 Arg	Thr Ala Thr Gly 1980 Val Thr His Gln 2060 Ile	Ile Leu Asp 1965 Pro Thr Ser Gly Arg 2045 Thr Ser	Thr Lys 1950 Glu Glu His Gly Phe 2030 Pro Thr Cys Gly	Gly 1935 Asn Leu Ile Pro Gln 2015 Arg Ile His	1920 Leu Asn Pro Leu Gly 2000 Gln Arg Pro Ser Pro 2080 Ser

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Val Val Thr Val Gly Asn Ser Val Asn Glu Gly Leu Asn Gln Pro Thr
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Asp Asp Ser Cys Phe Asp Pro Tyr Thr Val Ser His Tyr Ala Val Gly
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                                      2155
Asp Glu Trp Glu Arg Met Ser Glu Ser Gly Phe Lys Leu Leu Cys Gln
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                                  2170
Cys Leu Gly Phe Gly Ser Gly His Phe Arg Cys Asp Ser Ser Arg Trp
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                              2185
Cys His Asp Asn Gly Val Asn Tyr Lys Ile Gly Glu Lys Trp Asp Arg
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Gln Gly Glu Asn Gly Gln Met Met Ser Cys Thr Cys Leu Gly Asn Gly
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Lys Gly Glu Phe Lys Cys Asp Pro His Glu Ala Thr Cys Tyr Asp Asp
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Gly Lys Thr Tyr His Val Gly Glu Gln Trp Gln Lys Glu Tyr Leu Gly
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Ala Ile Cys Ser Cys Thr Cys Phe Gly Gly Gln Arg Gly Trp Arg Cys
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                              2265
Asp Asn Cys Arg Arg Pro Gly Gly Glu Pro Ser Pro Glu Gly Thr Thr
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                                              2285
Gly Gln Ser Tyr Asn Gln Tyr Ser Gln Arg Tyr His Gln Arg Thr Asn
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<213> Homo sapiens

165

<400> 99

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Ala Pro Leu Asp Leu Pro Gly Ser Ala Glu Pro Pro Lys Gln Cys His
Pro Cys Pro Gly Val Pro Gln Gly Thr Ser Pro Ala Pro Val Pro Tyr
Gly Tyr Phe Gly Gly Gly Tyr Tyr Ser Cys Arg Val Ser Arg Ser Ser
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Leu Lys Pro Cys Ala Gln Ala Ala Thr Leu Ala Ala Tyr Pro Ala Glu
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Thr Pro Thr Ala Gly Glu Glu Tyr Pro Ser Arg Pro Thr Glu Phe Ala
                         120
                                               125
Phe Tyr Pro Gly Tyr Pro Gly Thr Tyr His Ala Met Ala Ser Tyr Leu
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                                            140
Asp Val Ser Val Val Gln Thr Leu Gly Ala Pro Gly Glu Pro Arg His
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Asp Ser Leu Leu Pro Val Asp Ser Tyr Gln Ser Trp Ala Leu Ala Gly
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Gly Trp Asn Ser Gln Met Cys Cys Gln Gly Glu Gln Asn Pro Pro Gly
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Pro Phe Trp Lys Ala Ala Phe Ala Asp Ser Ser Gly Gln His Pro Pro
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Asp Ala Cys Ala Phe Arg Arg Gly Arg Lys Lys Arg Ile Pro Tyr Ser
                        215
Lys Gly Gln Leu Arg Glu Leu Glu Arg Glu Tyr Ala Ala Asn Lys Phe
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                                        235
Ile Thr Lys Asp Lys Arg Arg Lys Ile Ser Ala Ala Thr Ser Leu Ser
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Glu Arg Gln Ile Thr Ile Trp Phe Gln Asn Arg Arg Val Lys Glu Lys
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Lys Gly Ala Gly Asn Asn Pro Glu Phe Glu Glu Thr Arg Arg Val Phe
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			Leu	85					90					95	_
			Asp 100					105					110		_
		115					120					125		_	
	130		Gln			135				•	140		•		-
145			Cys		150					155					160
			Gln	165					170					175	
			Tyr 180					185					190	•	•
		195	Val				200					205			_
	210		Pro			215					220			_	_
His 225	Pro	Leu	Asn	His	Arg 230	Gln	Leu	Ser	Leu	Ser 235	Ser	Ser	Arg	Ser	Ser 240
Glu	Gly	Ser	Leu	Gly 245	Gly	Gln	Asn	Ser	Gly 250		Gly	Gly	Arg	Ser 255	Ser
Glu	Lys	Pro	Thr 260	Gly	Leu	Trp	Ser	Thr 265	Ala	Ser	Ser	Gln	Arg 270	Val	Ser
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	290		Pro			295					300				
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			Gly	325					330					335	_
			Gly 340					345					350		
		355	Gly				360					365			
	370		Pro			375					380				
385			Leu		390					395					400
			Gly -	405					410					415	
			Asp 420					425					430	-	
		435	Pro				440					445			
Leu	Glu 450	ATA	Leu	Thr	Gln	Arg 455	Leu	Glu	Arg	Glu	Met 460	Asp	Ala	His	Pro
465			Tyr		470					475	Ser				480
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Cys Phe Thr Cys Ala Ala Cys Ser Arg Lys Leu Arg Gly Lys Ala Phe 505 Tyr Phe Val Asn Gly Lys Val Phe Cys Glu Glu Asp Phe Leu Tyr Ser 520 Gly Phe Gln Gln Ser Ala Asp Arg Cys Phe Leu Cys Gly His Leu Ile 535 Met Asp Met Ile Leu Gln Ala Leu Gly Lys Ser Tyr His Pro Gly Cys 555 550 Phe Arg Cys Val Ile Cys Asn Glu Cys Leu Asp Gly Val Pro Phe Thr 570 Val Asp Ser Glu Asn Lys Ile Tyr Cys Val Arg Asp Tyr His Lys Val 585 Leu Ala Pro Lys Cys Ala Ala Cys Gly Leu Pro Ile Leu Pro Pro Glu 600 Gly Ser Asp Glu Thr Ile Arg Val Val Ser Met Asp Arg Asp Tyr His 615 620 Val Glu Cys Tyr His Cys Glu Asp Cys Gly Leu Glu Leu Asn Asp Glu 630 635 Asp Gly His Arg Cys Tyr Pro Leu Glu Asp His Leu Phe Cys His Ser 645 650 Cys His Val Lys Arg Leu Glu Lys Arg Pro Ser Ser Thr Ala Leu His Gln His His Phe 675 <210> 102 <211> 296 <212> PRT <213> Homo sapiens <400> 102 Ser Thr Gly Ser Glu Phe Pro Leu Cys Thr Lys Ala Ser Pro Cys Ser Ala Ala Arg Ala Gly Gly Arg Ala Leu Gly Trp Arg Leu Gln Gln 25 Arg Glu Thr Arg Gly Asn Pro Gly Asn Pro Gly Leu Gly Val Ala Ala 40 Thr Met Thr Gly Ser Asn Met Ser Asp Ala Leu Ala Asn Ala Val Cys Gln Arg Cys Gln Ala Arg Phe Ser Pro Ala Glu Arg Ile Val Asn Ser Asn Gly Glu Leu Tyr His Glu His Cys Phe Val Cys Ala Gln Cys Phe 90 Arg Pro Phe Pro Glu Gly Leu Phe Tyr Glu Phe Glu Gly Arg Lys Tyr 105 Cys Glu His Asp Phe Gln Met Leu Phe Ala Pro Cys Cys Gly Ser Cys 125 120 Gly Glu Phe Ile Ile Gly Arg Val Ile Lys Ala Met Asn Asn Asn Trp

130

135

140

His Pro Gly Cys Phe Arg Cys Glu Leu Cys Asp Val Glu Leu Ala Asp
145

150

155

160

Leu Gly Phe Val Lys Asn Ala Gly Arg His Leu Cys Arg Pro Cys His
165

170

175

Asn Arg Glu Lys Ala Lys Gly Leu Gly Lys Tyr Ile Cys Gln Arg Cys

Ash Arg Giu Lys Ala Lys Giy Leu Giy Lys Tyr Tie Cys Gin Arg Cys
180 185 190

His Leu Val Ile Asp Glu Gln Pro Leu Met Phe Arg Ser Asp Ala Tyr
195 200 205

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His Pro Asp His Phe Asn Cys Thr His Cys Gly Lys Glu Leu Thr Ala 215 Glu Ala Arg Glu Leu Lys Gly Glu Leu Tyr Cys Leu Pro Cys His Asp 230 235 Lys Met Gly Val Pro Ile Cys Gly Ala Cys Arg Arg Pro Ile Glu Gly 250 Arg Val Val Asn Ala Leu Gly Lys Gln Trp His Val Glu His Phe Val 265 Cys Ala Lys Cys Glu Lys Pro Phe Leu Gly His Arg His Tyr Glu Lys 280 Lys Gly Leu Ala Tyr Cys Glu Leu <210> 103 <211> 500 <212> PRT <213> Homo sapiens <400> 103 Met Gly Ile Gly Leu Ser Ala Gln Gly Val Asn Met Asn Arg Leu Pro Gly Trp Asp Lys His Ser Tyr Gly Tyr His Gly Asp Asp Gly His Ser Phe Cys Ser Ser Gly Thr Gly Gln Pro Tyr Gly Pro Thr Phe Thr Thr Gly Asp Val Ile Gly Cys Cys Val Asn Leu Ile Asn Asn Thr Cys Phe 55 Tyr Thr Lys Asn Gly His Ser Leu Gly Ile Ala Phe Thr Asp Leu Pro 70 Pro Asn Leu Tyr Pro Thr Val Gly Leu Gln Thr Pro Gly Glu Val Val 90 Asp Ala Asn Phe Gly Gln His Pro Phe Val Phe Asp Ile Glu Asp Tyr 105 Met Arg Glu Trp Arg Thr Lys Ile Gln Ala Gln Ile Asp Arg Phe Pro 120 Ile Gly Asp Arg Glu Gly Glu Trp Gln Thr Met Ile Gln Lys Met Val 135 Ser Ser Tyr Leu Val His His Gly Tyr Cys Ala Thr Ala Glu Ala Phe 150 Ala Arg Ser Thr Asp Gln Thr Val Leu Glu Glu Leu Ala Ser Ile Lys 170 Asn Arg Gln Arg Ile Gln Lys Leu Val Leu Ala Gly Arg Met Gly Glu 180 185 Ala Ile Glu Thr Thr Gln Gln Leu Tyr Pro Ser Leu Leu Glu Arg Asn 200 Pro Asn Leu Leu Phe Thr Leu Lys Val Arg Gln Phe Ile Glu Met Val 215 220 Asn Gly Thr Asp Ser Glu Val Arg Cys Leu Gly Gly Arg Ser Pro Lys 230 235 Ser Gln Asp Ser Tyr Pro Val Ser Pro Arg Pro Phe Ser Ser Pro Ser 245 250 Met Ser Pro Ser His Gly Met Asn Ile His Asn Leu Ala Ser Gly Lys 265 Gly Ser Thr Ala His Phe Ser Gly Phe Glu Ser Cys Ser Asn Gly Val 280 Ile Ser Asn Lys Ala His Gln Ser Tyr Cys His Ser Asn Lys His Gln 290 295

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Ser Ser Asn Leu Asn Val Pro Glu Leu Asn Ser Ile Asn Met Ser Arg 315 310 Ser Gln Gln Val Asn Asn Phe Thr Ser Asn Asp Val Asp Met Glu Thr 330 Asp His Tyr Ser Asn Gly Val Gly Glu Thr Ser Ser Asn Gly Phe Leu 345 Asn Gly Ser Ser Lys His Asp His Glu Met Glu Asp Cys Asp Thr Glu 360 Met Glu Val Asp Ser Ser Gln Leu Arg Arg Gln Leu Cys Gly Gly Ser 375 Gln Ala Ala Ile Glu Arg Met Ile His Phe Gly Arg Glu Leu Gln Ala 395 390 Met Ser Glu Gln Leu Arg Arg Asp Cys Gly Lys Asn Thr Ala Asn Lys 410 Lys Met Leu Lys Asp Ala Phe Ser Leu Leu Ala Tyr Ser Asp Pro Trp 425 430 420 Asn Ser Pro Val Gly Asn Gln Leu Asp Pro Ile Gln Arg Glu Pro Val 440 Cys Ser Ala Leu Asn Ser Ala Ile Leu Glu Thr His Asn Leu Pro Lys 455 Gln Pro Pro Leu Ala Leu Ala Met Gly Gln Ala Thr Gln Cys Leu Gly 470. 475 Leu Met Ala Arg Ser Gly Ile Gly Ser Cys Ala Phe Ala Thr Val Glu 490 Asp Tyr Leu His 500 <210> 104 <211> 387 <212> PRT <213> Homo sapiens <400> 104 Met Ala Thr Ser Gly Val Leu Pro Gly Gly Gly Phe Val Ala Ser Ala 10 Ala Ala Val Ala Gly Pro Glu Met Gln Thr Gly Arg Asn Asn Phe Val Ile Arg Arg Asn Pro Ala Asp Pro Gln Arg Ile Pro Ser Asn Pro Ser His Arg Ile Gln Cys Ala Ala Gly Tyr Glu Gln Ser Glu His Asn Val Cys Gln Asp Ile Asp Glu Cys Thr Ala Gly Thr His Asn Cys Arg Ala 75 70 Asp Gln Val Cys Ile Asn Leu Arg Gly Ser Phe Ala Cys Gln Cys Pro 90 Pro Gly Tyr Gln Lys Arg Gly Glu Gln Cys Val Asp Ile Asp Glu Cys

85 90 95

Pro Gly Tyr Gln Lys Arg Gly Glu Gln Cys Val Asp Ile Asp Glu Cys 100 105 110

Thr Ile Pro Pro Tyr Cys His Gln Arg Cys Val Asn Thr Pro Gly Ser 115 120 125

Phe Tyr Cys Gln Cys Ser Pro Gly Phe Gln Leu Ala Ala Asn Asn Tyr 130 135 140

Thr Cys Val Asp Ile Asn Glu Cys Asp Ala Ser Asn Gln Cys Ala Gln 145 150 155 160

Gln Cys Tyr Asn Ile Leu Gly Ser Phe Ile Cys Gln Cys Asn Gln Gly

Tyr Glu Leu Ser Ser Asp Arg Leu Asn Cys Glu Asp Ile Asp Glu Cys
180 185 190

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Arg Thr Ser Ser Tyr Leu Cys Gln Tyr Gln Cys Val Asn Glu Pro Gly 200 Lys Phe Ser Cys Met Cys Pro Gln Gly Tyr Gln Val Val Arg Ser Arg 215 Thr Cys Gln Asp Ile Asn Glu Cys Glu Thr Thr Asn Glu Cys Arg Glu 230 235 Asp Glu Met Cys Trp Asn Tyr His Gly Gly Phe Arg Cys Tyr Pro Arg 245 250 Asn Pro Cys Gln Asp Pro Tyr Ile Leu Thr Pro Glu Asn Arg Cys Val 265 Cys Pro Val Ser Asn Ala Met Cys Arg Glu Leu Pro Gln Ser Ile Val 280 Tyr Lys Tyr Met Ser Ile Arg Ser Asp Arg Ser Val Pro Ser Asp Ile 295 Phe Gln Ile Gln Ala Thr Thr Ile Tyr Ala Asn Thr Ile Asn Thr Phe 310 315 Arg Ile Lys Ser Gly Asn Glu Asn Gly Glu Phe Tyr Leu Arg Gln Thr 325 330 Ser Pro Val Ser Ala Met Leu Val Leu Val Lys Ser Leu Ser Gly Pro 345 Arg Glu His Ile Val Asp Leu Glu Met Leu Thr Val Ser Ser Ile Gly 360 Thr Phe Arg Thr Ser Ser Val Leu Arg Leu Thr Ile Ile Val Gly Pro Phe Ser Phe 385 <210> 105 <211> 531 <212> PRT <213> Homo sapiens <400> 105 Met Ser Lys Pro His Ser Glu Ala Gly Thr Ala Phe Ile Gln Thr Gln 10 Gln Leu His Ala Ala Met Ala Asp Thr Phe Leu Glu His Met Cys Arg Leu Asp Ile Asp Ser Pro Pro Ile Thr Ala Arg Asn Thr Gly Ile Ile Cys Thr Ile Gly Pro Ala Ser Arg Ser Val Glu Thr Leu Lys Glu Met 55 Ile Lys Ser Gly Met Asn Val Ala Arg Leu Asn Phe Ser His Gly Thr 70 75 His Glu Tyr His Ala Glu Thr Ile Lys Asn Val Arg Thr Ala Thr Glu Ser Phe Ala Ser Asp Pro Tyr Leu Tyr Arg Pro Val Ala Val Ala Leu 105 Asp Thr Lys Gly Pro Glu Ile Arg Thr Gly Leu Ile Lys Gly Ser Gly 120 Thr Ala Glu Leu Glu Leu Lys Lys Gly Ala Thr Leu Lys Ile Thr Leu 135 140 Asp Asn Ala Tyr Met Glu Lys Cys Asp Glu Asn Ile Leu Trp Leu Asp 150 Tyr Lys Asn Ile Cys Lys Val Val Glu Val Gly Ser Lys Ile Tyr Val 170 165 Asp Asp Gly Leu Ile Ser Leu Gln Val Lys Gln Lys Gly Ala Asp Phe 180

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Phe Ala Ser Phe Ile Arg Lys Ala Ser Asp Val His Glu Val Arg Lys
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Val Leu Gly Glu Lys Gly Lys Asn Ile Lys Ile Ile Ser Lys Ile Glu
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Lys Val Phe Leu Ala Gln Lys Met Met Ile Gly Arg Cys Asn Arg Ala
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Asp Tyr Pro Leu Glu Ala Val Arg Met Gln His Leu Ile Ala Arg Glu
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                        375
Ala Glu Ala Ala Ile Tyr His Leu Gln Leu Phe Glu Glu Leu Arg Arg
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Leu Ala Pro Ile Thr Ser Asp Pro Thr Glu Ala Thr Ala Val Gly Ala
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                                    410
Val Glu Ala Ser Phe Lys Cys Cys Ser Gly Ala Ile Ile Val Leu Thr
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Lys Ser Gly Arg Ser Ala His Gln Val Ala Arg Tyr Arg Pro Arg Ala
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Pro Ile Ile Ala Val Thr Arg Asn Pro Gln Thr Ala Arg Gln Ala His
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Leu Tyr Arg Gly Ile Phe Pro Val Leu Cys Lys Asp Pro Val Gln Glu
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Leu 145	Ser	Asp	Glu	Val	Lys 150	Arg	Lys	Gln	Tyr	Asp 155	Ala	Tyr	Gly	Ser	Ala 160
Gly	Phe	Asp	Pro	Gly 165	Ala	Ser	Gly	Ser	Gln 170	His	Ser	Tyr	Trp	Lys 175	Gly
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		195	Ser			_	200					205	_		
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	290		Cys	_	_	295					300				
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			Lys	325					330					335	
			Arg 340					345					350		
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•		435					440					445			
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420 425 430 Cys Glu Gly Cys Tyr Thr Asp Thr Leu Glu Lys Cys Asn Thr Cys Gly 440 Glu Pro Ile Thr Asp Arg Met Leu Arg Ala Thr Gly Lys Ala Tyr His 455 460 Pro His Cys Phe Thr Cys Val Val Cys Ala Arg Pro Leu Glu Gly Thr 470 475 Ser Phe Ile Val Asp Gln Ala Asn Arg Pro His Cys Val Pro Asp Tyr 490 485 His Lys Gln Tyr Ala Pro Arg Cys Ser Val Cys Ser Glu Pro Ile Met 500 505 Pro Glu Pro Gly Arg Asp Glu Thr Val Arg Val Val Ala Leu Asp Lys 520 Asn Phe His Met Lys Cys Tyr Lys Cys Glu Asp Cys Gly Lys Pro Leu 535 Ser Ile Glu Ala Asp Asp Asn Gly Cys Phe Pro Leu Asp Gly His Val 550 555 Leu Cys Arg Lys Cys His Thr Ala Arg Ala Gln Thr 565 <210> 108 <211> 2861 <212> PRT <213> Homo sapiens <400> 108 Met Lys Ala Met Asp Val Leu Pro Ile Leu Lys Glu Lys Val Ala Tyr Leu Ser Gly Gly Arg Asp Lys Arg Gly Gly Pro Ile Leu Thr Phe Pro 25 Ala Arg Ser Asn His Asp Arg Ile Arg Gln Glu Asp Leu Arg Arg Leu 40 Ile Ser Tyr Leu Ala Cys Ile Pro Ser Glu Glu Val Cys Lys Arg Gly Phe Thr Val Ile Val Asp Met Arg Gly Ser Lys Trp Asp Ser Ile Lys 70 75 Pro Leu Leu Lys Ile Leu Gln Glu Ser Phe Pro Cys Cys Ile His Val Ala Leu Ile Ile Lys Pro Asp Asn Phe Trp Gln Lys Gln Arg Thr Asn 100 105 110 Phe Gly Ser Ser Lys Phe Glu Phe Glu Thr Asn Met Val Ser Leu Glu 120 Gly Leu Thr Lys Val Val Asp Pro Ser Gln Leu Thr Pro Glu Phe Asp 140

135 Gly Cys Leu Glu Tyr Asn His Glu Glu Trp Ile Glu Ile Arg Val Ala 150 155 Phe Glu Asp Tyr Ile Ser Asn Ala Thr His Met Leu Ser Arg Leu Glu 165 170 Glu Leu Gln Asp Ile Leu Ala Lys Lys Glu Leu Pro Gln Asp Leu Glu 180 185 190

Gly Ala Arg Asn Met Ile Glu Glu His Ser Gln Leu Lys Lys Lys Val 195 200 205

Ile Lys Ala Pro Ile Glu Asp Leu Asp Leu Glu Gly Gln Lys Leu Leu 215 220

Gln Arg Ile Gln Ser Ser Glu Ser Phe Pro Lys Lys Asn Ser Gly Ser 230 235 Gly Asn Ala Asp Leu Gln Asn Leu Leu Pro Lys Val Ser Thr Met Leu

				245					250					255	
			260					His 265					270		
	_	275	Ī		_		280	Phe				285			
_	290					295		Ile			300		_		
305			-		310			Thr		315					320
				325				Ala	330		-			335	_
			340	_				Val 345					350		
_		355					360	Arg				365			
	370	_	_			375		Ala			380	_			
385					390			Gln	_	395		-	-		400
		-		405	_			Cys	410					415	
			420					11e 425					430		_
		435				-	440	Glu				445	_	_	
	450					455		Leu			460				
465					470	_	ŧ	Lys		475					480
				485				Gln	490					495	
		_	500		_			Gln 505					510		
		515					520	Asp	V			525		_	
	530					535		Val			540				
545				-	550			Asp		555					560
				565		•		Leu	570					575	
			580					585					590		Gln
		595				_	600	Val	_			605			
	610					615					620				Leu
625		_			630			_		635					Tyr 640
				645					650					655	Gln
			660					665					670		Gly
		675					680					685			Lys
Thr	Pro 690	HIS	ASN	ser	ser	695	Asn	His	ш	Glu	700	val	Leu	GIn	Gln

Leu 705	Asp	Glu	Ala	Gln	Ser 710	Gln	Met	Glu	Glu	Leu 715	Phe	Gln	Glu	Arg	Lys 720
Ile	Lys	Leu	Glu	Leu 725	Phe	Leu	His	Val	Arg 730	Ile	Phe	Glu	Arg	Asp 735	Ala
Ile	Asp	Ile	Ile 740	Ser	Asp	Leu	Glu	Ser 745	Trp	Asn	Asp	Glu	Leu 750	Ser	Gln
Gln	Met	Asn 755	Asp	Phe	Asp	Thr	Glu 760	Asp	Leu	Thr	Ile	Ala 765	Glu	Gln	Arg
	770				V	775					Asn 780				
785					790					795	Tyr				800
			_	805					810		Asp			815	
	_		820	_				825			Glu		830		
	•	835					840				Leu	845			
	850	_				855					Val 860				
865		_			870					875	Ile				880
				885					890		Glu			895	
			900					905			Val		910		
		915					920				Asp	925			
_	930		_			935					Leu 940				
945	-				950					955	Ala				960
				965					970		Glu			975	
_			980					985			Leu		990		
		995					100	0			His	100	5		
	101	0				101	5				Arg 102	0			
Phe 102		Lys	Tyr	Leu	H1S		Asn	Ser	var	103	Met 5	Pro	GIÀ	met	1040
		Ile	Lys	Ala 104	Pro		Gln	Gln	Val 105	Lys	Asn	Ile	Leu	Asn 105	
Leu	Phe	Gln	Arg	Glu		Arg	Val	Leu 106		Tyr	Trp	Thr	Met 107		Lys
Arg	Arg	Leu 107	Asp		Cys	Gln	Gln 108	Tyr		Val	Phe	Glu 108		Ser	Ala
Lys	Gln 109	Ala		Glu	Trp	Ile 109		Asp	Asn	Gly	Glu 110		Tyr	Leu	Ser
Thr 110	His		Ser	Thr	Gly 111	Ser		Ile	Gln	His 111		Gln	Glu	Leu	Leu 1120
		His	Glu	Glu 112		Gln	Ile	Thr	Ala 113		Gln	Thr	Lys	Glu 113	Arg 5
Val	Lys	Leu	Leu 114		Gln	Leu	Ala	Asp 114	Gly		Cys	Glu	Lys 115		His
Ala	His	Ala	Ala	Glu	Ile	Lys	Lys	Cys	Val	Thr	Ala	Val	Asp	Lys	Arg

1155		1160		1165	
Tyr Arg Asp Phe 1170	117.	5	1180	)	
Lys Ala Leu Gly 1185	1190		1195		1200
Gln Leu Asp Ile	1205	1	1210		1215
Arg Asp Ala Ala 122	0	1225		1230	
Arg Lys Glu Phe 1235		1240		1245	
Val Arg Asp Leu 1250	Arg Glu Cys 125		Thr Tyr Leu 1260	_	Met Thr
Ser Gly Val Glu 1265	1270	_	1275		1280
Ile Phe Gly Asn	1285	1	L290		1295
Leu Lys Glu Leu 130	0 .	1305		1310	
Cys Phe Val Thr 1315		1320		1325	
Lys Asn Lys Pro 1330	133	5	1340	)	
Tyr Phe Asp Glu 1345		Arg His G	Gly Leu Ala 1355	Asn Ser	Ile Ser 1360
Ser Tyr Leu Ile		Gln Arg I			
Leu Lys Glu Leu 138	_	Cys Glu G 1385	Glu Gly Lys	Gly Glu 1390	
Asp Gly Leu Glu 1395	Val Met Leu	Ser Val E	Pro Lys Arg	Ala Asn 1405	Asp Ala
Met His Leu Ser 1410	Met Leu Glu 141		Asp Glu Asn 1420		Ser Gln
Gly Glu Leu Ile 1425	1430		1435	•	1440
Leu Ile Arg Lys	Gly Arg Glu 1445		Leu Phe Leu 1450	Phe Glu	Met Ser 1455
Leu Val Phe Ser 146	0 ·	1465		1470	)
Leu Tyr Lys Ser 1475	Lys Leu Phe	Thr Ser (	Glu Leu Gly	Val Thr 1485	Glu His
Val Glu Gly Asp 1490	Pro Cys Lys 149		Leu Trp Val 150		Thr Pro
Thr Ser Asp Asn 1505	1510		1515		1520
Gln Asp Trp Ile	Lys His Ile 1525		Val Ile Gln 1530	Glu Arg	Thr Ile 1535
His Leu Lys Gly 154		Glu Pro 1 1545		Pro Lys 1550	
Pro Ala Thr Arg 1555	Gln Lys Gly	Arg Arg 1 1560	Asp Gly Glu	Asp Leu 1565	Asp Ser
Gln Gly Asp Gly 1570	157	5	158	0	
Thr Ser Gln Asn 1585	1590		1595		1600
Leu Thr Val Val	Ile His Asp 1605		Ala Cys Asn 1610	Ser Asn	Glu Leu 1615

1620 16	<del>-</del> -
Lys Pro Asp Trp Cys Leu Val Arg Th	r Thr Asp Arg Ser Pro Ala Ala 1645
Glu Gly Leu Val Pro Cys Gly Ser Let 1650 1655	u Cys Ile Ala His Ser Arg Ser 1660
Ser Met Glu Met Glu Gly Ile Phe Ass 1665 1670	1675 1680
Ser Ser Asn Asp Ala Ser Pro Pro Ala 1685	a Ser Val Ala Ser Leu Gln Pro 1690 1695
His Met Ile Gly Ala Gln Ser Ser Pro 1700 17	
Thr Leu Arg Lys Trp Leu Thr Ser Pro 1715 1720	1725
Lys Ala Asp Gly His Val Lys Lys Let 1730 1735	1740
Arg Glu Val Arg Lys Ser Ala Asp Ala 1745 1750	1755 1760
Asp Ser Ala Ala Thr Pro Gln Asp Gl 1765	1770 1775
Asn Glu Gly Leu Ser Ser Gly Thr Let 1780 17	85 1790
Met Gln Ser Cys Gly Glu Glu Glu Gl 1795 1800	1805
Pro Leu Pro Pro Pro Met Ala Ile Gli 1810 1815	. 1820
Asp Ser Gln Asp Asp Lys Ala Ser Se 1825 1830	1835 1840
Ser Ser Glu Thr Pro Ser Ala Ala Gl 1845	1850 1855
	65 1870
Val Aca Cla Cly Aca Car Car Car Dr.	
1875 1880	o Ser Phe Asn Pro Ser Asp Asn 1885
1875 1880 Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895	1885 e Asp Glu Met Glu Glu Arg Lys 1900
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910	1885 e Asp Glu Met Glu Glu Arg Lys 1900 r Val Leu Gln Glu Leu Val Glu 1915 1920
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925	1885 e Asp Glu Met Glu Glu Arg Lys 1900 r Val Leu Gln Glu Leu Val Glu 1915 u Gly Tyr Val Val Glu Gly Tyr 1930 1935
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19	1885 e Asp Glu Met Glu Glu Arg Lys 1900 r Val Leu Gln Glu Leu Val Glu 1915 1920 ru Gly Tyr Val Val Glu Gly Tyr 1930 1 Pro Asp Asp Met Lys Gly Lys 1950
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960	1885  e Asp Glu Met Glu Glu Arg Lys 1900  r Val Leu Gln Glu Leu Val Glu 1915  u Gly Tyr Val Val Glu Gly Tyr 1930  1 Pro Asp Asp Met Lys Gly Lys 45  s Gln Ile Tyr Asp Trp His Arg 1965
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975	e Asp Glu Met Glu Glu Arg Lys 1900 r Val Leu Gln Glu Leu Val Glu 1915 1920 u Gly Tyr Val Val Glu Gly Tyr 1930 1 Pro Asp Asp Met Lys Gly Lys 45 1950 s Gln Ile Tyr Asp Trp His Arg 1965 rs Cys Leu Glu Asp Pro Glu Lys 1980
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975  Leu Gly Ser Leu Phe Val Lys His Gl 1985	1885  e Asp Glu Met Glu Glu Arg Lys 1900  r Val Leu Gln Glu Leu Val Glu 1915  u Gly Tyr Val Val Glu Gly Tyr 1930  l Pro Asp Asp Met Lys Gly Lys 45  s Gln Ile Tyr Asp Trp His Arg 1965  c Cys Leu Glu Asp Pro Glu Lys 1980  u Arg Arg Leu His Met Tyr Ile 1995
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975  Leu Gly Ser Leu Phe Val Lys His Gl 1985 1990  Ala Tyr Cys Gln Asn Lys Pro Lys Se 2005	1885  e Asp Glu Met Glu Glu Arg Lys 1900  r Val Leu Gln Glu Leu Val Glu 1915  u Gly Tyr Val Val Glu Gly Tyr 1930  l Pro Asp Asp Met Lys Gly Lys 1950  s Gln Ile Tyr Asp Trp His Arg 1965  s Cys Leu Glu Asp Pro Glu Lys 1980  u Arg Arg Leu His Met Tyr Ile 1995  c Glu His Ile Val Ser Glu Tyr 2010
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975  Leu Gly Ser Leu Phe Val Lys His Gl 1985 1990  Ala Tyr Cys Gln Asn Lys Pro Lys Se 2005  Ile Asp Thr Phe Phe Glu Asp Leu Ly 2020 20	## 1885  ## Asp Glu Met Glu Glu Arg Lys
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975  Leu Gly Ser Leu Phe Val Lys His Gl 1985 1990  Ala Tyr Cys Gln Asn Lys Pro Lys Se 2005  Ile Asp Thr Phe Phe Glu Asp Leu Ly 2020 20  Gln Leu Thr Asp Leu Leu Ile Lys Pr	## 1885  ## Asp Glu Met Glu Glu Arg Lys
1875 1880  Ser Leu Leu Ser Ser Ser Ser Pro II 1890 1895  Ser Ser Ser Leu Lys Arg Arg His Ty 1905 1910  Thr Glu Arg Asp Tyr Val Arg Asp Le 1925  Met Ala Leu Met Lys Glu Asp Gly Va 1940 19  Asp Lys Ile Val Phe Gly Asn Ile Hi 1955 1960  Asp Phe Phe Leu Gly Glu Leu Glu Ly 1970 1975  Leu Gly Ser Leu Phe Val Lys His Gl 1985 1990  Ala Tyr Cys Gln Asn Lys Pro Lys Se 2005  Ile Asp Thr Phe Phe Glu Asp Leu Ly 2020 20  Gln Leu Thr Asp Leu Leu Ile Lys Pr	## 1885  ## Asp Glu Met Glu Glu Arg Lys

2065	2070	2075		2080
Arg Arg Cys Asn Asp 208	5	2090	209	95
Gly Lys Ile Val Ala 2100		2105	2110	
Val Thr Asp Gln Asp 2115	212	0	2125	
Ile Phe Leu Phe Glu 2130	2135	21	40	
Lys Lys Gly Phe Ser 2145	2150	2155		2160
Val Ser Cys Leu Cys 216	5	2170	217	75
Phe Ala Leu Thr Ser 2180	-	2185	2190	
His Ser Ser Ser Pro	220	0	2205	
Gln Ile Leu Glu Asn 2210 Ile Glu Tyr Gln Arg	2215	22	20	
2225 Gly Ala Ala Ala Gly	2230	2235		2240
224 Ala Ala Ala Ala Thr	5	2250	225	55
2260 Ala Arg Ala Gly Ala		2265	2270	•
2275 Thr Pro Pro Cys Trp	228	0	2285	
2290	2295	23	300 -	
Thr Arg Cys Gln Ser 2305	2310	2315		2320
Leu Val Thr His Asp	25	2330	23:	35
Tyr Gln Gly Glu Val 2340		2345	2350	. V
Phe Leu Val Phe Arg 2355 Trp Ile Pro Gly Phe	236	0	2365	
2370 Asn Pro Asp Gly Thi	2375	2:	380	
2385 Arg Leu Arg Lys Lys	2390	2395		2400
240 Gly Lys Leu Glu Ass	)5	2410	24	15
2420 Lys Val Ser Val Lys		2425	2430	
2435	244	0	2445	
Pro Glu Phe Val Ile 2450	2455	2	460	
Thr Val Val Leu Arg	2470	2475		2480
Thr Trp Lys Gly Pro	35	2490	24	95
Ser Ile Ser Tyr Ser 2500	_	2505	2510	
Val Thr Thr Glu Asp 2515	Asp Gly Ile 252		2525	n Asp

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Met Gly Ser Ala Ser Ser Ser Ala Ser Leu Arg Val Leu Gly Pro Gly 2535 2530 Met Asp Gly Ile Met Val Thr Trp Lys Asp Asn Phe Asp Ser Phe Tyr 2550 2555 2560 Ser Glu Val Ala Glu Leu Gly Arg Gly Arg Phe Ser Val Val Lys Lys 2570 2575 2565 Cys Asp Gln Lys Gly Thr Lys Arg Ala Val Ala Thr Lys Phe Val Asn 2585 2580 Lys Lys Leu Met Lys Arg Asp Gln Val Thr His Glu Leu Gly Ile Leu 2600 Gln Ser Leu Gln His Pro Leu Leu Val Gly Leu Leu Asp Thr Phe Glu 2615 2620 Thr Pro Thr Ser Tyr Ile Leu Val Leu Glu Met Ala Asp Gln Gly Arg 2630 2635 Leu Leu Asp Cys Val Val Arg Trp Gly Ser Leu Thr Glu Gly Lys Ile 2650 2645 Arg Ala His Leu Gly Glu Val Leu Glu Ala Val Arg Tyr Leu His Asn 2660 2665 Cys Arg Ile Ala His Leu Asp Leu Lys Pro Glu Asn Ile Leu Val Asp 2680 2685 Glu Ser Leu Ala Lys Pro Thr Ile Lys Leu Ala Asp Phe Gly Asp Ala 2695 2700 Val Gln Leu Asn Thr Thr Tyr Tyr Ile His Gln Leu Leu Gly Asn Pro 2715 2710 Glu Phe Ala Ala Pro Glu Ile Ile Leu Gly Asn Pro Val Ser Leu Thr 2725 2730 Ser Asp Thr Trp Ser Val Gly Val Leu Thr Tyr Val Leu Leu Ser Gly 2740 2745 Val Ser Pro Phe Leu Asp Asp Ser Val Glu Glu Thr Cys Leu Asn Ile 2760 2765 Cys Arg Leu Asp Phe Ser Phe Pro Asp Asp Tyr Phe Lys Gly Val Ser 2775 2780 Gln Lys Ala Lys Glu Phe Val Cys Phe Leu Leu Gln Glu Asp Pro Ala 2795 2790 Lys Arg Pro Ser Ala Ala Leu Ala Leu Gln Glu Gln Trp Leu Gln Ala 2810 2805 Gly Asn Gly Arg Ser Thr Gly Val Leu Asp Thr Ser Arg Leu Thr Ser 2825 2820 Phe Ile Glu Arg Arg Lys His Gln Asn Asp Val Arg Pro Ile Arg Ser 2840 Ile Lys Asn Phe Leu Gln Ser Arg Leu Leu Pro Arg Val 2850 2855 <210> 109 <211> 271 <212> PRT <213> Homo sapiens <400> 109 Met Val Leu Ile Lys Glu Phe Arg Val Val Leu Pro Cys Ser Val Gln

 Met Val Leu Ile Lys Glu Phe Arg Val Val Leu Pro Cys Ser Val Gln

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 5
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 15

 Glu Tyr Gln Val Gly Gln Leu Tyr Ser Val Ala Glu Ala Ser Lys Asn
 20
 25
 30

 Glu Thr Gly Gly Gly Glu Gly Ile Glu Val Leu Lys Asn Glu Pro Tyr
 35
 40
 45

 Glu Lys Asp Gly Glu Lys Gly Gln Tyr Thr His Lys Ile Tyr His Leu
 50
 60

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Lys Ser Lys Val Pro Ala Phe Val Arg Met Ile Ala Pro Glu Gly Ser Leu Val Phe His Glu Lys Ala Trp Asn Ala Tyr Pro Tyr Cys Arg Thr Ile Val Thr Asn Glu Tyr Met Lys Asp Asp Phe Phe Ile Lys Ile Glu 105 . Thr Trp His Lys Pro Asp Leu Gly Thr Leu Glu Asn Val His Gly Leu 120 Asp Pro Asn Thr Trp Lys Thr Val Glu Ile Val His Ile Asp Ile Ala 135 Asp Arg Ser Gln Val Glu Pro Ala Asp Tyr Lys Ala Asp Glu Asp Pro 155 150 Ala Leu Phe Gln Ser Val Lys Thr Lys Arg Gly Pro Leu Gly Pro Asn 170 165 Trp Lys Lys Glu Leu Ala Asn Ser Pro Asp Cys Pro Gln Met Cys Ala 185 190 180 Tyr Lys Leu Val Thr Ile Lys Phe Lys Trp Trp Gly Leu Gln Ser Lys 200 Val Glu Asn Phe Ile Gln Lys Gln Glu Lys Arg Ile Phe Thr Asn Phe 215 His Arg Gln Leu Phe Cys Trp Ile Asp Lys Trp Ile Asp Leu Thr Met 235 230 Glu Asp Ile Arg Arg Met Glu Asp Glu Thr Gln Lys Glu Leu Glu Thr 250 Met Arg Lys Arg Gly Ser Val Arg Gly Thr Ser Ala Ala Asp Val 265

<210> 110

<211> 233

<212> PRT

<213> Homo sapiens

<400> 110

Asn Ser Val Leu Asn Ser Asn Ala Ile Lys Asn Leu Pro Pro Pro Leu 10 Gly Gly Ala Ala Gly His Pro Gly Ser Ala Val Ser Ala Ala Pro Gly Ile Leu Tyr Pro Gly Gly Asn Lys Tyr Gln Thr Ile Asp Asn Tyr Gln Pro Tyr Pro Cys Ala Glu Asp Glu Glu Cys Gly Thr Asp Glu Tyr Cys Ala Ser Pro Thr Arg Gly Gly Asp Ala Gly Val Gln Ile Cys Leu Ala 75 70 Cys Arg Lys Arg Lys Arg Cys Met Arg His Ala Met Cys Cys Pro Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn His 105 Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp 120 His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys 140 135 Met Tyr His Thr Lys Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser 150

Asp Cys Ala Ser Gly Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile

Cys Lys Pro Val Leu Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg 180 185 190

170

Lys Gly Ser His Gly Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu
195 200 205

Gly Leu Ser Cys Arg Ile Gln Lys Asp His His Gln Ala Ser Asn Ser
210 215 220

Ser Arg Leu His Thr Cys Gln Arg His
225 230

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<400> 112

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Cys Arg Lys Arg Arg Lys Arg Cys Met Arg His Ala Met Cys Cys Pro 85
Gly Asn Tyr Cys Lys Asn Gly Ile Cys Val Ser Ser Asp Gln Asn His 100
Phe Arg Gly Glu Ile Glu Glu Thr Ile Thr Glu Ser Phe Gly Asn Asp 115
His Ser Thr Leu Asp Gly Tyr Ser Arg Arg Thr Thr Leu Ser Ser Lys 130
Met Tyr His Thr Lys
145
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 Ser Val Cys
 Leu Arg
 Ser Ser Asp
 Cys Ala
 Ser Gly Leu Cys Cys Ala
 60

 Arg His Phe Trp
 Ser Lys Ile Cys Lys Pro Val Leu Lys Glu Gly Gln
 75
 80

 Val Cys Thr Lys His Arg Arg Lys Gly Ser His Gly Leu Glu Ile Phe
 85
 90
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 Gln Arg Cys Tyr Cys Gly Glu Glu Gly Leu Ser Cys Arg Ile Gln Lys Asp
 100
 105
 110

 His His Gln Ala Ser Asn Ser Ser Arg Leu His Thr Cys Gln Arg His
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<400> 115

Gly Gln Glu Gly Ser Val Cys Leu Arg Ser Ser Asp Cys Ala Ser Gly 10 15

Leu Cys Cys Ala Arg His Phe Trp Ser Lys Ile Cys Lys Pro Val Leu 20 25 30

Lys Glu Gly Gln Val Cys Thr Lys His Arg Arg Lys Gly Ser His Gly 35

Leu Glu Ile Phe Gln Arg Cys Tyr Cys Gly Glu Gly Leu Ser Cys Arg 50 55 60

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



15 A

# (54) Title: REAGENTS AND METHODS FOR MODULATING DKK-MEDIATED INTERACTIONS

(57) Abstract: The present invention provides reagents, compounds, compositions, and methods relating to novel interactions of the extracellular domain of LRP5, HBM (a variant of LRP5), and/or LRP6 with Dkk, including Dkk-1. The various nucleic acids, polypeptides, antibodies, assay methods, diagnostic methods, and methods of treatment of the present invention are related to and impact on Dkk, LRP5, LRP6, HBM, and Wnt signaling. Dkk, LRP5, LRP6, HBM, and Wnt are implicated in bone and lipid cellular signaling. Thus, the present invention provides reagents and methods for modulating lipid levels and/or bone mass and is useful in the treatment and diagnosis of abnormal lipid levels and bone mass disorders, such as osteoporosis.

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/15982

A. CLASSIFICATION OF SUBJECT MATTER					
IPC(7) : A61K 39/395, 00, 38 US CL : 424/130.1, 184.1					
According to International Patent Classification (IPC) or to both	national classification and IPC				
B. FIELDS SEARCHED					
Minimum documentation searched (classification system followed by classification symbols) U.S.: 424/130.1, 184.1					
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched					
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Continuation Sheet					
C. DOCUMENTS CONSIDERED TO BE RELEVANT					
Category * Citation of document, with indication, where	appropriate, of the relevant passages	Relevant to claim No.			
A GONG, Y et al. LDL RECEPTOR-RELATED PR ACCRUAL AND EYE DEVELOPMENT, CELL		1-11 and 13-23			
ZORN A. WNT SIGNALLING: ANTABONISTIC BIOLOGY, 2001, VOL.11, No.15, PAGES R592	1-11 and 13-23				
A WO 9846743 A1 (THE WELL-COME TRUST L (22.10.98) see entire document.		1-11 and 13-23			
(22.10/36) 600 6/1110 6/04/11111					
•					
Further documents are listed in the continuation of Box C.	See patent family annex.				
Special categories of cited documents:	"T" later document published after the inte date and not in conflict with the applie				
"A" document defining the general state of the art which is not considered to be of particular relevance	principle or theory underlying the inve	ention			
"E" earlier application or patent published on or after the international filing date					
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the considered to involve an inventive ste	when the document is			
"O" document referring to an oral disclosure, use, exhibition or other means	combined with one or more other such being obvious to a person skilled in th				
"P" document published prior to the international filing date but later than the priority date claimed	"&" document member of the same patent	family			
Date of the actual completion of the international search	Date of mailing of the international sea				
18 March 2003 (18.03.2003)	08 AUG 200				
Name and mailing address of the ISA/US  Commissioner of Patents and Trademarks  Box PCT	Wilchall A Belvavani Bel	1-Harro			
Washington, D.C. 20231	Talanhana Na. 702/202 0106				
Facsimile No. (703)305-3230	Telephone No. 703/308-0196				

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/15982

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)			
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:			
1. Claim Nos.: because they relate to subject matter not required to be searched by this Authority, namely:			
Claim Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:			
3. Claim Nos.:  because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).			
Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)			
This International Searching Authority found multiple inventions in this international application, as follows: Please See Continuation Sheet			
<ol> <li>As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.</li> <li>As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.</li> <li>As only some of the required additional search fees were timely paid by the applicant, this international search</li> </ol>			
report covers only those claims for which fees were paid, specifically claims Nos.:  No required additional search fees were timely paid by the applicant. Consequently, this international search report			
is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-11 and 13-23			
Remark on Protest The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.			

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# INTERNATIONAL SEARCH REPORT

# BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

- I. Claims 1-11 and 13-23, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP5.
- II. Claims 1-11 and 13-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to LRP6.
- III. Claims 1-11 and 13-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which inhibits Dkk binding to HBM.
- IV. Claims 1-10, 12-14 and 16-23 drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP5.
- V. Claims 1-10, 12-14 and 16-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to LRP6.
- VI. Claims 1-10, 12-14 and 16-20, drawn to a method of regulating LRP5 activity in a subject, comprising administering a composition which enhances Dkk binding to HBM.
- VII. Claims 2 4, 7-11, 13-20, 21-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.
- VIII. Claims 2 4, 7-11, 13-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.
- IX. Claims 2 4, 7-11, 13-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.
- X. Claims 2 4, 7-10, 12-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with RP5, wherein said composition enhances Dkk binding to LRP5.
- XI. Claims 2 4, 7-10, 12-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.
- XII. Claims 2 4, 7-10, 12-20, 24-27, drawn to a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition enhances Dkk binding to HBM.
- XIII. Claims 2 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.
- XIV. Claims 2 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to I RP6.
- XV. Claims 2 4, 7-11, 13-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

# INTERNATIONAL SEARCH REPORT

- XVI. Claims 2 4, 7-10, 12-23, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.
- XIV. Claims 2 4, 7-10, 12-20, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.
- XV. Claims 2 4, 7-10, 12-20, 28-32 drawn to a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.
- XVI. Claims 2 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.
- XVII. XVI. Claims 2 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to IRP6
- XVIII. Claims 2 4, 7-11, 13-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.
- XIX. Claims 2 4, 7-10, 12-23, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.
- XX. Claims 2 4, 7-10, 12-20, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.
- XXI. Claims 2 4, 7-10, 12-20, 33-35 drawn to a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.
- XXII. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of Dkk.
- XXIII. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP5.
- XXIV. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP6.
- XXV. Claims 2, 35 and 36 drawn to a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of HBM.
- XXVI. Claims 2, 37-43, 44-47, drawn to a method of screening for a compound which modulates the interaction of DKK with LRP5.
- XXVII. Claims 2, 37-43, 47, drawn to a method of screening for a compound which modulates the interaction of DKK with LRP6.
- XXVIII. Claims 2, 37-43, 47, drawn to a method of screening for a compound which modulates the interaction of DKK with HBM.
- XXIX. Claims 2,41-43, 48-49, drawn to a method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting proteins.

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XXX. Claims 50-63 drawn to a composition comprising a LRP5 and a pharmaceutical acceptable carrier thereof.

XXXI. Claims 50-59, 63 drawn to a composition comprising a LRP6 and a pharmaceutical acceptable carrier thereof.

XXXII. Claims 50- 59, 63 drawn to a composition comprising a HBM and a pharmaceutical acceptable carrier thereof.

XXXIII. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and LRP5 interaction.

XXXIV. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and LRP6 interaction.

XXXV. Claims 2 and 64 drawn to a method for identifying compound which modulate Dkk and HBM interaction.

XXXVI. Claims 2 and 65 drawn to a method of identifying binding partners for a Dkk protein.

XXXVII. Claims 66-68 drawn to a nucleic acid and a vector encoding a Dkk interacting protein.

XXXVIII. Claims 69-90, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP5.

XXXIX. Claims 69-87, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP6.

XL. Claims 69-87, drawn to a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a HBM.

XLI. Claims 91-92 drawn to a transgenic animal.

XLII. Claims 2 and 93 drawn to a method for identifying potential compound which modulate Dkk activity.

XLIII - LXII. Claim 94, drawn to one specific peptide aptamer of one specific SEQ ID NOs: 171-88; 189-192.

LXIII- LXXIX. Claims 95-97, drawn to an antibody which specifically recognizes and binds to specific peptides of SEQ ID NOs: 110-127.

LXXX. Claims 2, 98-100, drawn to a method of identifying Dkk interacting protein which modulate the interaction of Dkk with Wnt signaling pathway.

LXXXI. Claims 2, 25 and 101-104, drawn to a method for identifying Dkk interacting proteins.

LXXXII. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and LRP5 interaction.

LXXXIII. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and LRP6 interaction.

LXXXIV. Claims 2, 105-106, drawn to a method for identifying compounds which modulate Dkk and HBM interaction.

LXXXV. Claims 2, 25, 107-110, drawn to a method for identifying compound which modulate the interaction of Dkk with Wnt signaling pathway.

LXXXVI. Claims 2, 111, drawn to a method of testing compounds that modulate Dkk-mediated activity in a mammal.

LXXXVII. Claims 2, 112, 113, drawn to method of screening for compound or composition which modulate the interaction of Dkk and Dkk interacting protein.

LXXXVIII-CIX. Claim 114 drawn to antibody which recognizes and binds to one specific SEQ ID NOs: 171-192.

INTERNATIONAL SEARCH REPORT	
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The inventions listed as Groups 1-109 do not relate to a single general inventive concept under PCT Rule 13.1 because, under I	PCT
Rule 13.2, they lack the same or corresponding special technical features for the following reasons:  The invention listed as groups 1-109 do not related to a single general inventive concept under PCT Rule 13.1 because, under F Rule 13.2, hey lack the same or corresponding special technical features for the following reasons:	<b>·</b> ÇT
The special technical features of Group I is considered a method of regulating LRP5 activity in a subject, comprising administration which inhibits Dkk binding to LRP5.	ering a
The special technical features of Group II is considered a method of regulating LRP5 activity in a subject, comprising administ a composition which inhibits Dkk binding to LRP6.	tering
The special technical features of Group III is considered a method of regulating LRP5 activity in a subject, comprising adminis a composition which inhibits Dkk binding to HBM.	
The special technical features of Group IV is considered a method of regulating LRP5 activity in a subject, comprising adminis a composition which enhances Dkk binding to LRP5.  The special technical features of Group V is considered a method of regulating LRP5 activity in a subject, comprising administ	
composition which enhances of Group VI is considered a method of regulating LRP5 activity in a subject, comprising administration of the special technical features of Group VI is considered a method of regulating LRP5 activity in a subject, comprising administration which enhances. Disk binding to HPM	

Form PCT/ISA/210 (second sheet) (July 1998)

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# INTERNATIONAL SEARCH REPORT

The special technical features of Group VII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group VIII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group IX is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group X is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with RP5, wherein said composition enhances Dkk binding to LRP5.

The special technical features of Group XI is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XII is considered a method of regulating Dkk-Wnt pathway activity, comprising administering a composition which modulates Dkk activity, or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XIII is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group XIV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group XV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group XVI is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

The special technical features of Group XIV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XV is considered a method of modulating bone mass in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XVI is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition inhibits Dkk binding to LRP5.

The special technical features of Group XVII is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition inhibits Dkk binding to LRP6.

The special technical features of Group XVIII is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition inhibits Dkk binding to HBM.

The special technical features of Group XIX is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP5, wherein said composition enhances Dkk binding to LRP5.

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The special technical features of Group XX is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with LRP6, wherein said composition enhances Dkk binding to LRP6.

The special technical features of Group XXI is considered a method of modulating lipid levels in a subject comprising administering to the subject a composition which modulates DKK activity or interaction with HBM, wherein said composition enhances Dkk binding to HBM.

The special technical features of Group XXII is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of Dkk.

The special technical features of Group XXIII is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP5.

The special technical features of Group XXIV is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of LRP6.

The special technical features of Group XXV is considered a method of diagnosing low or high bone mass and /or high or low lipid levels in a subject comprising examining expression of HBM.

The special technical features of Group XXVI is considered a method of screening for a compound which modulates the interaction of DKK with LRP5.

The special technical features of Group XXVII is considered a method of screening for a compound which modulates the interaction of DKK with LRP6.

The special technical features of Group XXVIII is considered a method of screening for a compound which modulates the interaction of DKK with HBM.

The special technical features of Group XXIX is considered a method of screening for a compound which modulates the interaction of Dkk with a Dkk interacting proteins.

The special technical features of Group XXX is considered a composition comprising a LRP5 and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXI is considered a composition comprising a LRP6 and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXII is considered a composition comprising a HBM and a pharmaceutical acceptable carrier thereof.

The special technical features of Group XXXIII is considered a method for identifying compound which modulate Dkk and LRP5 interaction.

The special technical features of Group XXXIV is considered a method for identifying compound which modulate Dkk and LRP6 interaction.

The special technical features of Group XXXV is considered a method for identifying compound which modulate Dkk and HBM interaction.

The special technical features of Group XXXVI is considered a method of identifying binding partners for a Dkk protein.

The special technical features of Group XXXVII is considered a nucleic acid and a vector encoding a Dkk interacting protein.

The special technical features of Group XXXVIII is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP5.

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The special technical features of Group XXXIX is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a LRP6.

The special technical features of Group XL is considered a method of detecting a modulatory activity of a compound, wherein the first peptide is a Dkk peptide and the second peptide is a HBM.

The special technical features of Group XLI is considered a transgenic animal.

The special technical features of Group XLII is considered a method for identifying potential compound which modulate Dkk activity.

The special technical features of Group XLIII - LXII is considered one specific peptide aptamer of one specific SEQ ID NOs: 171-88: 189-192.

The special technical features of Group LXIII- LXXIX is considered an antibody which specifically recognizes and binds to specific peptides of SEQ ID NOs: 110-127.

The special technical features of Group LXXX is considered a method of identifying Dkk interacting protein which modulate the interaction of Dkk with Wnt signaling pathway.

The special technical features of Group LXXXI is considered a method for identifying Dkk interacting proteins.

The special technical features of Group LXXXII is considered a method for identifying compounds which modulate Dkk and LRPS interaction.

The special technical features of Group LXXXIII is considered a method for identifying compounds which modulate Dkk and LRP6 interaction.

The special technical features of Group LXXXIV is considered a method for identifying compounds which modulate Dkk and HBM interaction.

The special technical features of Group LXXXV is considered a method for identifying compound which modulate the interaction of Dkk with Wnt signaling pathway.

The special technical features of Group LXXXVI is considered a method of testing compounds that modulate Dkk-mediated activity in a mammal.

The special technical features of Group LXXXVII is considered a method of screening for compound or composition which modulate the interaction of Dkk and Dkk interacting protein.

The special technical features of Group LXXXVIII-CIX. is considered an antibody which recognizes and binds to one specific SEQ ID NOs: 171-192.

Accordingly, Groups I-CIX are not so linked by the same or corresponding special technical feature within meaning of PCT Rule 13.2 so as to form a single general inventive concept.

# INTERNATIONAL SEARCH REPORT Biosis, CAPLUS, SciSearch, Medline, EMBASE, WEST, USPATFULL, PCTFULL search terms; Allen K; Anisowicz, A; Bhat, B; Damagnez, V, Robinson, J; Yaworsky, P; DKK, Dkk1, LRP5, SEQ ID NO:28, protein OST262; osteoporosis.

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